CD200R antibodies, mouse

For research use only

30 µg equal 100 tests, 150 µg equal 500 tests. One test corresponds to labeling of 10 cells.

Product	Content	Order no.
CD200R-Biotin	150 µg in 1 mL	130-112-456
CD200R-FITC	30 µg in 200 µL	130-112-529
CD200R-PE	30 µg in 200 µL	130-112-530
CD200R-PE	150 µg in 1 mL	130-112-458
CD200R-APC	30 µg in 200 µL	130-112-531
CD200R-APC	150 µg in 1 mL	130-112-459
CD200R-PE-Vio770	30 µg in 200 µL	130-112-532
CD200R-PE-Vio770	150 µg in 1 mL	130-112-460
CD200R-APC-Vio770	30 µg in 200 µL	130-112-533
CD200R-VioBright 515	30 µg in 200 µL	130-112-534
CD200R-Biotin	30 µg in 200 µL	130-112-528

Warnings

Reagents contain sodium azide. Under acidic conditions sodium azide yields hydrazoic acid, which is extremely toxic. Azide compounds should be diluted with running water before discarding. These precautions are recommended to avoid deposits in plumbing where explosive conditions may develop.

Technical data and background information

Antigen	CD200R
Clone	REA850
Isotype	recombinant human IgG1
Isotype control	REA Control antibodies
Alternative names of antigen	OX2R, CD200 Receptor
Entrez Gene ID	57781
Molecular mass of antigen [kDa]	33
Distribution of antigen	neutrophils
Product format	Reagents are supplied in buffer containing stabilizer and 0.05% sodium azide.
Storage	Store protected from light at 2–8 °C. Do not freeze.

Clone REA850 recognizes the mouse CD200R antigen, also known as orexin receptor 2 (OX2R). CD200R is a type I transmembrane glycoprotein expressed by cells of the myeloid lineage, e.g., monocytes, neutrophils, T cells, and mast cells. The interaction between CD200R and its ligand CD200 is involved in immune suppression and controls functions of monocytes and macrophages.

Additional information: Clone REA850 displays negligible binding to Fc receptors.

Reagent requirements

- Buffer: Prepare a solution containing phosphate-buffered saline (PBS), pH 7.2, 0.5% bovine serum albumin (BSA), and 2 mM EDTA by diluting MACS® BSA Stock Solution (# 130-091-376) 1:20 with autoMACS® Rinsing Solution (# 130-091-222). Keep buffer cold (2−8 °C).
  Note: EDTA can be replaced by other supplements such as anticoagulant citrate dextrose formula-A (ACD-A) or citrate phosphate dextrose (CPD). Buffers or media containing Ca^{2+} or Mg^{2+} are not recommended for use.
- (Optional) Fluorochrome-conjugated anti-biotin antibodies, e.g., Anti-Biotin-PE (# 130-090-756) as secondary antibody reagent in combination with biotinylated antibodies.
- (Optional) Propidium Iodide Solution (# 130-093-233) for flow cytometric exclusion of dead cells without fixation.
- (Optional) Fixation and Dead Cell Discrimination Kit (# 130-091-163) for cell fixation and flow cytometric exclusion of dead cells.

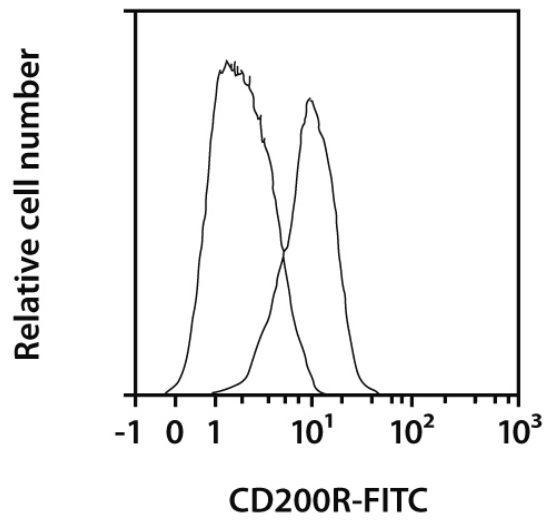
Protocol for cell surface staining

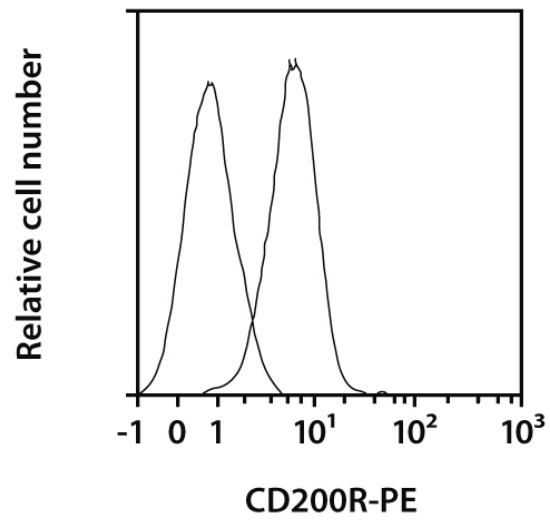
- The recommended antibody dilution for labeling of cells and subsequent analysis by flow cytometry is 1:50 for up to 10^6 cells/100 µL.
- Volumes given below are for up to 10^6 nucleated cells. When working with fewer than 10^6 cells, use the same volumes as indicated. When working with higher cell numbers, scale up all reagent volumes and total volumes accordingly.

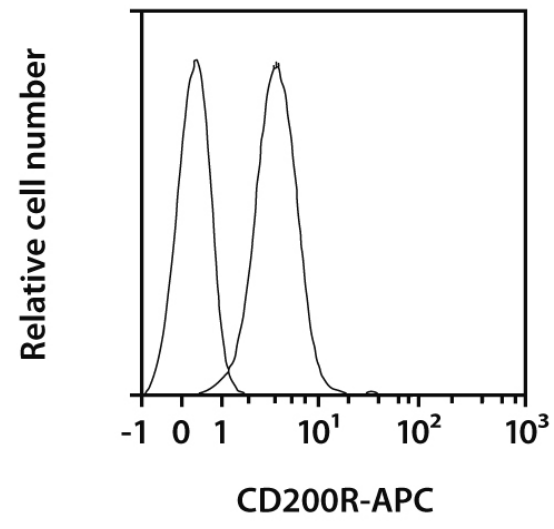
1. Determine cell number.
2. Centrifuge cell suspension at 300×g for 10 minutes. Aspirate supernatant completely.
3. Resuspend up to 10^6 nucleated cells per 98 µL of buffer.
4. Add 2 µL of the antibody.
5. Mix well and incubate for 10 minutes in the dark in the refrigerator (2−8 °C).
   Note: Higher temperatures and/or longer incubation times may lead to non-specific cell labeling. Working on ice requires increased incubation times.
6. Wash cells by adding 1−2 mL of buffer and centrifuge at 300×g for 10 minutes. Aspirate supernatant completely.
7. (Optional) If biotinylated antibody was used, resuspend the cell pellet in buffer and stain with fluorochrome-conjugated anti-biotin antibody according to the manufacturer's recommendations.
8. Resuspend cell pellet in a suitable amount of buffer for analysis by flow cytometry or fluorescence microscopy.

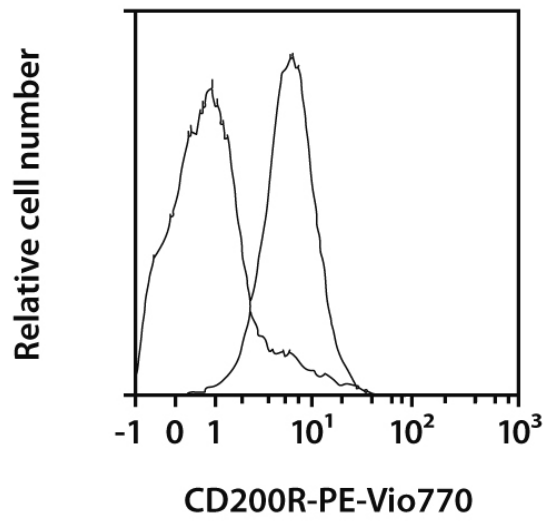
Examples of immunofluorescent staining

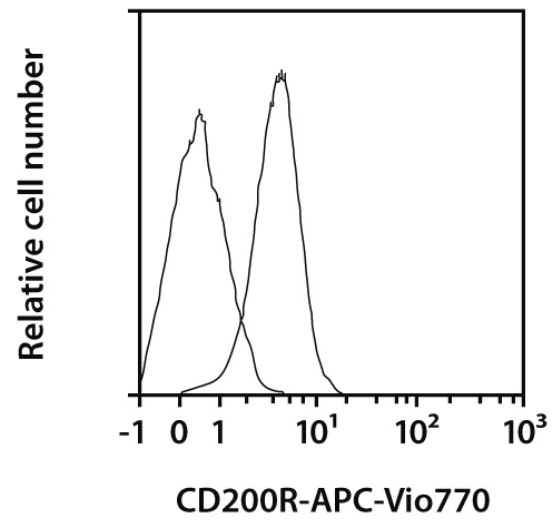
Thioglycollate-induced peritoneal exudate macrophages from C57BL/6 mice were stained with CD200R antibodies or with the corresponding REA Control antibodies (left peak) as well as with Anti-F4/80 antibodies. Flow cytometry was performed using the MACSQuant®Analyzer. Cell debris and dead cells were excluded from the analysis based on scatter signals and propidium iodide fluorescence or 4',6-diamidino-2-phenylindole (DAPI) fluorescence, as in the case of tandem conjugates.

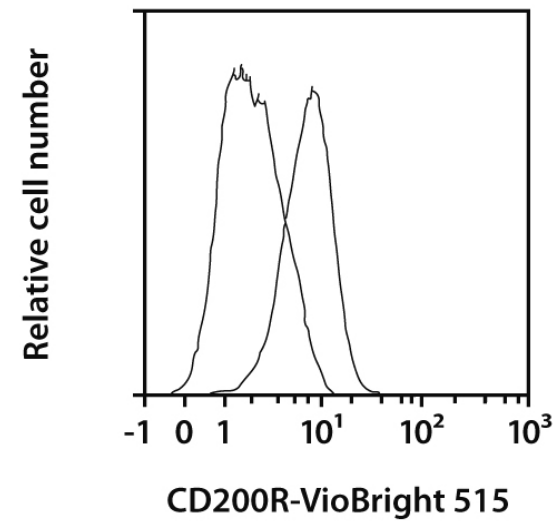


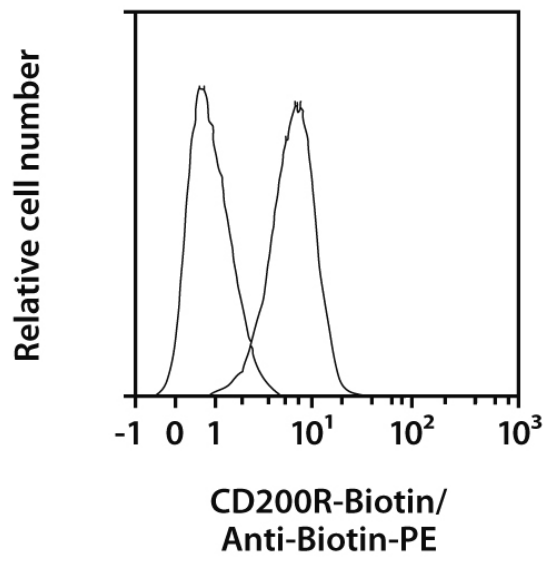












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