USER GUIDE

novex® by life technologies™

Human/Monkey Extracellular Protein Buffer Reagent for Magnetic Kits

For simultaneous quantitative determination of multiple extracellular proteins in human or monkey serum, plasma, and tissue culture supernatant

Catalog Number LHB0001M

Revision 1.00
Publication Part Number MAN0005492

life technologies™

Contents

Kit Contents and Storage

Storage

All components of the Human/Monkey Extracellular Protein Buffer Reagent for Magnetic Kits are shipped at 2–8°C. Upon receipt, store all kit components at 2–8°C. **Do not freeze**.

Contents

The components and amounts included in the Human/ Monkey Extracellular Protein Buffer Reagent for Magnetic Kits are listed below.

Reagents Provided	100 Test Kit
Wash Solution Concentrate (20X) (contains 0.1% sodium azide)	15 mL × 3 bottles
Assay Diluent (contains 0.1% sodium azide)	15 mL × 1 bottle
Incubation Buffer (contains 0.05% sodium azide)	12 mL × 1 bottle
Biotin Diluent (contains 3.3 mM thymol)	12 mL × 1 bottle
Streptavidin-RPE Concentrate (10X) (contains 0.1% sodium azide)	1 mL × 1 vial
Streptavidin-RPE Diluent (contains 3.3 mM thymol)	12 mL × 1 bottle
96-well Filter Plate	1 × 96-well plate
96-well Flat Bottom Plate	1 × 96-well plate

Introduction

Overview

Purpose

Singleplex and Multiplex Bead Immunoassay Kits from Life Technologies are developed to maximize flexibility in experimental design, permitting the measurement of one or multiple proteins in panels designed by the researcher. The Human/Monkey Extracellular Protein Buffer Reagent for Magnetic Kits contains all the reagents that are intended for use with the Luminex® 100™, 200™ FLEXMAP 3D® or the MAGPIX® instrument. xPONENT® software package for data acquisition and analysis is provided for control of the Luminex® 100/200™, FLEXMAP 3D® and MAGPIX® instrumentation. These instruments are manufactured by Luminex Corporation and are sold by Life Technologies.

Background information

Advances in the field of cell biology have defined a complex and interdependent set of extracellular and intracellular signaling molecules that control normal cell function. There is growing interest among researchers as well as drug discovery groups in simultaneously monitoring multiple components of signaling pathways. Solid phase multiplex protein assays are the tools of choice in these studies as they maximize efficiency by simultaneously profiling several proteins within individual samples.

The Singleplex Multiplex Bead Immunoassays are solid phase protein immunoassays that use spectrally encoded antibody-conjugated beads as the solid support. The spectral beads are suitable for use in singleplex assays or may be mixed for multiplex assays according to the researcher's requirements. Each assay is carefully designed and tested to assure that sensitivity, range, and correlation are maximized. The assay is performed in a 96-well plate format and analyzed with a Luminex® 100™, 200™, FLEXMAP 3D® or MAGPIX® instrument which monitors the spectral properties of the capture beads while simultaneously measuring the quantity of associated fluorophore.

Continued on next page

Overview, Continued

Background information, continued

Standard curves generated with this assay system extend over several orders of magnitude of concentrations, while the sensitivity and quantitation of the assays are comparable to ELISAs (Enzyme Linked-Immunosorbent Assays). Assay standards are calibrated to National Institute for Biological Standards and Controls (NIBSC) reference preparations, when available, to assure accurate and reliable results.

The Human/Monkey Extracellular Protein Buffer Reagent for Magnetic Kits is designed for the quantitative determination of proteins in serum, plasma, and tissue culture supernatant.

Visit **www.lifetechnologies.com/luminex** for a current listing of available multiplex bead immunoassays and reagents.

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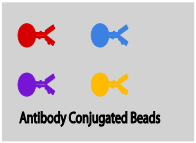
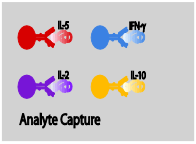
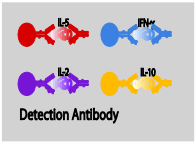
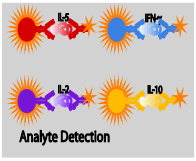
Overview, Continued

Assay overview









The xMAP® technology combines the efficiencies of multiplexing up to 100 different proteins for simultaneous analysis, with reproducibility similar to ELISA. This assay uses 6.5 µm polystyrene beads which contain magnetite.

The beads are internally dyed with red and infrared fluorophores of differing intensities. Each bead is given a unique number, or bead region, allowing differentiation of one bead from another.

Beads of defined spectral properties which have been conjugated to protein-specific capture antibodies are added into the wells of a microplate along with samples (including standards of known protein concentration, control samples, and test samples), Proteins in the samples will bind to the capture antibodies over the course of a 2 hour incubation.

The beads are then washed by either using a filter plate, manually with the aid of a magnetic separator, or with the aid of automated magnetic bead washing equipment.

After washing the beads, protein-specific biotinylated detector antibodies are added to the assay and incubated for 1 hour. During this incubation, the protein-specific biotinylated detector antibodies bind to the appropriate bead-immobilized proteins.

After removal of excess biotinylated detector antibodies by washing, streptavidin conjugated to the fluorescent protein, R-Phycoerythrin (Streptavidin-RPE), is added and allowed to incubate for 30 minutes. The Streptavidin-RPE binds to the biotinylated detector antibodies associated with the immune complexes on the beads, forming a four-member solid phase sandwich.

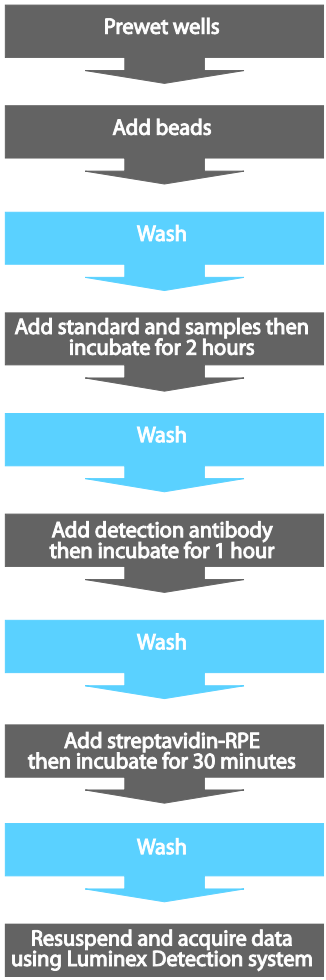
After washing to remove unbound Streptavidin-RPE, the beads are analyzed with the Luminex detection system. By monitoring the spectral properties of the beads and the amount of associated R-Phycoerythrin (RPE) fluorescence, the concentration of one or more proteins can be determined.

Experimental Overview

Experimental outline

The experimental outline for using the Human/Monkey Extracellular Protein Buffer Reagent for Magnetic Kits with human or monkey extracellular singleplex magnetic kits is shown below.

Note: Pre-wet step required only with the filter bottom plate.



Methods

Before Starting

Materials required but not provided

- Any of the following Life Technologies Luminex® xMAP® systems with data acquisition and analysis software:
  - Luminex® 200™ system (Cat. no. MAP0200)
  - FLEXMAP 3D® system (Cat. no. FM3D000)
  - MAGPIX® system (Cat. no. MPX0001)

  Contact Life Technologies for instrument and software placement services, see page 27.
- Bead washing equipment. This kit may be used with any of the following:
  - Magnetic 96-Well Separator (Cat. no. A14179).
  - Automated magnetic bead washing equipment
  - EveryPrep™ Universal Vacuum Manifold (Cat. no. K2111-01) or equivalent filtration vacuum manifold.
- Sonicating water bath
- Vortex mixer
- Orbital shaker (small diameter rotation recommended)
- Calibrated, adjustable, precision pipettes, preferably with disposable plastic tips (a manifold multi-channel pipette is desirable)
- Distilled or deionized water
- Glass or polypropylene tubes
- Aluminum foil

Procedural notes

Review the procedural notes below before starting the protocol.

- **All phases of the assay are performed using the filter plate or flat bottom plate provided with the kit. Do not invert the plates during the assay**.
- The filter plate is provided for use when washing steps are performed with a vacuum manifold. **Do not exceed 5 mm Hg**. With the filter bottom plate, contents are emptied from the bottom of the plate during washing.

Continued on next page

Before Starting, Continued

Procedural notes, continued

- The flat bottom plate is provided for use when washing steps are performed with a magnetic separator. With the flat bottom plate, contents are removed from the top of the plate during washing. Washing with the flat bottom plate may be performed manually or with the aid of automated washing equipment.
- Do not freeze any component of this kit. Store kit components at 2–8°C when not in use. Allow all reagents to warm to room temperature before use (air-warm all reagents at room temperature for at least 30 minutes, or alternatively, in a room temperature water bath for 20 minutes (except plate and standard vials).
- **The fluorescent beads are light-sensitive**. Protect the beads from light to avoid photobleaching of the embedded dye. Use aluminum foil to cover test tubes used in the assay. Cover microplates containing beads with an opaque or aluminum foil-wrapped plate cover. Since the amber vial does not provide full light protection, keep the vial covered in the box or in a dark drawer when not in use.
- Do not expose beads to organic solvents.
- Do not place filter plates on absorbent paper towels during loading or incubations, as liquid may be lost due to contact wicking. An extra plate cover is a recommended surface to rest the filter plate. Following plate washing, remove excess liquid and blot from the bottom of the plate by pressing the plate on clean paper towels.
- When pipetting reagents, maintain a consistent order of addition from well-to-well to ensure equal incubation times for all wells.
- To prevent filter tearing, avoid touching the filter plate membrane with pipette tips.
- Do not use reagents after kit expiration date.
- It is recommended that in-house controls be included with every assay. If control values fall outside pre-established ranges, the assay may be suspect. Contact Life Technologies Technical Support for product and technical assistance.
- Do not mix or substitute reagents with those from other lots or sources.

Continued on next page

Before Starting, Continued

Procedural notes, continued

- Handle all blood components and biological materials as potentially hazardous. Follow standard precautions as established by the Centers for Disease Control and Prevention and by the local Occupational Safety and Health Administration when handling and disposing of infectious agents.
- This kit contains materials with small quantities of sodium azide. Sodium azide reacts with lead and copper plumbing to form explosive metal azides. Upon disposal, flush drains with a large volume of water to prevent azide accumulation. Avoid ingestion and contact with eyes, skin and mucous membranes. In case of contact, rinse affected area with plenty of water. Observe all federal, state and local regulations for disposal.

Recommended plate plan

It is recommended to design a plate plan before starting the assay. A plate plan template is provided on page 31. The following is a suggested plate plan:

	1	2	3	4	5	6	7	8	9	10	11	12
A	B	Std 4										
B	B	Std 4										
C	Std 1	Std 5										
D	Std 1	Std 5										
E	Std 2	Std 6										
F	Std 2	Std 6										
G	Std 3	Std 7										
H	Std 3	Std 7										

B= blank (Assay Diluent), Standards 1 through 7, highest concentration to lowest.
The remainder of the plate is available for controls and samples which may be run as a singlet or in duplicate, as desired.

Note: Running all standards, samples and controls in duplicate is recommended.

Preparing Reagents

Introduction

Review the information in this section before starting. The Human/Monkey Extracellular Protein Buffer Reagent for Magnetic Kits is intended for use with human or monkey extracellular singleplex magnetic kits from Life Technologies. Prepare components of the buffer kit according to instructions below. **Note:** Bring all reagents and samples to room temperature before use.

Preparing wash solution

Upon storage at 2–8°C, a precipitate may form in the 20X Wash Solution Concentrate. If this occurs, warm the 20X Wash Solution Concentrate to 37°C and mix until the precipitate is dissolved.

1. Prepare a 1X Working Wash Solution for use with a 96-well plate by transferring the entire contents of the Wash Solution Concentrate bottle to a 500 mL container (or equivalent) and then add 285 mL of deionized water. Mix well.
2. The 1X Working Wash Solution is stable for up to 2 weeks when stored at 2–8°C.

Note: To prepare smaller volumes of 1X Working Wash Solution, mix 1 part of 20X concentrate with 19 parts of deionized water. Mix well.

Continued on next page

Preparing Reagents, Continued

Guidelines for sample preparation

- Serum, plasma, and tissue culture supernatants are suitable for use with Singleplex and Multiplex Bead Immunoassays from Life Technologies. Additional sample types may be suitable but have not been thoroughly validated. If possible, avoid the use of hemolyzed or lipemic sera. The appropriate sample types are defined on the Technical Data Sheet included with this multiplex panel.
- Collect samples in pyrogen/endotoxin-free tubes. Centrifuge, separate, and transfer samples to polypropylene tubes for storage.
- Analyze samples shortly after collection or thawing. Freeze samples after collection if samples will not be tested immediately. Avoid multiple freeze-thaw cycles of frozen samples. Thaw completely and mix well (**do not vortex)** prior to analysis.
- Clarify all samples by centrifugation (1000 × *g* for 10 minutes) and/or filter prior to analysis to prevent clogging of the filter plates.
- In the event that the sample concentrations exceed the standard curve, dilute samples and reanalyze. Dilute the serum or plasma samples in Assay Diluent and dilute tissue culture supernatants in the corresponding tissue culture medium.

Continued on next page

Preparing Reagents, Continued

Guidelines for standard curve preparation

- Each human or monkey extracellular singleplex magnetic kits comes with 2 complete sets of standard vials so that 2 runs of the assay can be made with freshly prepared standards.
- Reconstitute the protein standard within 1 hour of performing the assay.
- Before performing standard mixing and serial dilutions confirm reconstitution volumes on the Technical Data Sheet included with the Extracellular Protein Buffer Reagent Kit.
- The concentrations of the protein components of the standard are indicated on the Technical Data Sheet.
- Perform standard dilutions in glass or polypropylene tubes.
- When using serum or plasma samples, reconstitute the standard with Assay Diluent provided.
- If using other sample types (e.g., tissue culture supernatant), reconstitute the standard with a mixture, composed of 50% Assay Diluent and 50% of the matrix which closely resembles the sample type (50%/50% mixture). For example: When the sample type is RPMI medium containing 5% FBS, the standards should be reconstituted in a mixture composed of 50% Assay Diluent and 50% RPMI containing 5% FBS.

Reconstituting lyophilized standards

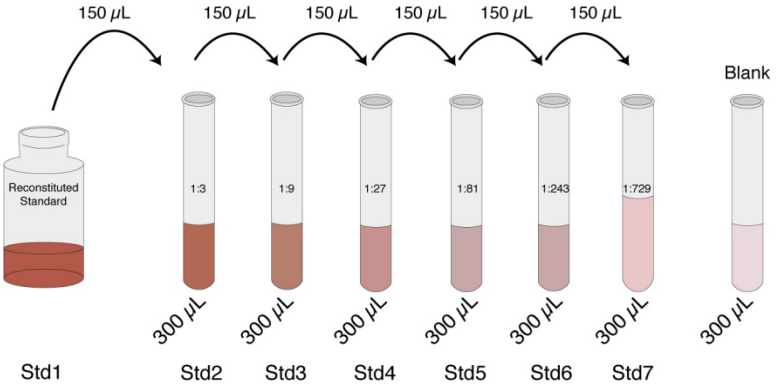
1. To the standard vials, add the suggested reconstitution volume of the appropriate diluent (see next page). **Do not vortex. When mixing or reconstituting protein solutions, always avoid foaming.**
2. Replace the vial stopper and allow the vial to stand undisturbed for 10 minutes.
3. Gently swirl and invert the vial 2–3 times to ensure complete reconstitution and allow the vial to sit at room temperature for an additional 5 minutes.

Continued on next page

Preparing Reagents, Continued

Preparing standard curve

The standard curve is made by serially diluting the reconstituted standard in Assay Diluent for serum and plasma samples or a mixture of 50% Assay Diluent and 50% tissue culture medium for tissue culture supernatant samples. See below. **Do not vortex.** Mix by gently pipetting up and down 5–10 times.



Serum/plasma: Assay Diluent
Tissue culture: 50% Assay Diluent / 50% Tissue Culture Medium

Discard all remaining reconstituted and diluted standards after completing assay. Return the Assay Diluent to the kit.

Online tool

Go to **www.lifetechnologies.com/luminex** under Multiplex Solution Tools, click Luminex® Calculation Worksheet for auto calculation of all assay dilutions.

Continued on next page

Preparing Reagents, Continued

Important

Before preparing Antibody Beads confirm concentration of beads on the Antibody Bead vial label. **The fluorescent beads are light-sensitive. Protect antibody conjugated beads from light during handling.**

Preparing 1X Antibody Beads

The Antibody Bead Concentrate (sold separately in the Antibody Bead Kit) is supplied as a **10X concentrate and must be diluted prior to use.** The fluorescent beads are light-sensitive. **Protect antibody conjugated beads from light during handling.**

1. Immediately before dispensing, vortex the 10X Antibody Bead Concentrate for 30 seconds followed by sonication in a sonicating water bath for 30 seconds.
2. Prepare 1X Antibody Bead stock by diluting 2.5 µL of 10X beads in 25 µL of Working Wash Solution (page 8) per assay well. Each well requires 25 µL of the diluted beads. See the following table for examples of how to combine volumes.

Note: Dilution factor is 1:11 for extra pipetting volume

Number of Wells	Vol. 10X AntibodyBead Concentrate	Vol. Working Wash Solution
24	0.06 mL	0.6 mL
32	0.08 mL	0.8 mL
40	0.10 mL	1.0 mL
48	0.12 mL	1.2 mL
56	0.14 mL	1.4 mL
64	0.16 mL	1.6 mL
72	0.18 mL	1.8 mL
80	0.20 mL	2.0 mL
88	0.22 mL	2.2 mL
96	0.24 mL	2.4 mL

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Assay Procedure

Washing methods

This assay may be washed using a vacuum manifold (requires the filter bottom plate provided), or by aid of a magnetic separator (requires the flat bottom plate provided).

Incomplete washing adversely affects assay results. Perform all wash steps with the Wash Solution supplied with the kit.

Filter Plate Method:

1. To wash beads, place the filter plate on the vacuum manifold and aspirate the liquid with gentle vacuum (**do not exceed 5 mm Hg**). Excessive vacuum can cause the membrane to tear, resulting in antibody bead loss. Prevent any vacuum surge by opening and adjusting the vacuum on the manifold before placing the plate on the manifold surface.
2. Stop the vacuum pressure as soon as the wells are empty.

   Note: If solution remains in the wells during vacuum aspiration, **do not detach the bottom of the 96-well filter plate.** In some cases, minor clogs in the filter plate may be dislodged by carefully pressing the bottom of the plate under the clogged well with the pointed end of a 15 mL plastic conical tube. Place the filter plate on a clean paper towel and use a gloved thumb or a 1 mL Pasteur pipette bulb to plunge the top of the clogged well. Empty all clogged wells entirely before continuing the washes.

   Do not attempt to repetitively pull vacuum on plates with clogged wells. This can compromise the unclogged wells and bead loss may occur.
3. Release the vacuum and remove the plate from the manifold. **Note**: Do not attempt to pull the plate off the vacuum manifold while the vacuum is still on or filter plate damage may occur.
4. Lightly tap or press the filter plate onto clean paper towels (hold the plate in the center for tapping) to remove excess fluid from the bottom of the filter plate. **Do not invert plate.**
5. Following the final aspiration and plate taps, use a clean absorbent towel to blot the bottom of the filter plate before addition of next liquid phase or data acquisition step.
6. **Do not leave plate on absorbent surface when adding reagents**.

Continued on next page

Assay Procedure, Continued

Washing methods, continued

Magnetic Separator Method:

1. Place 96-well Flat Bottom Plate provided with the kit onto the magnetic separator.
2. Incubate the plate on the magnet for 60–90 seconds so that the beads are immobilized on the bottom of the plate.
3. Lift the magnet while keeping the plate securely in place on top, and invert the magnet and plate (held securely together) over an appropriate disposal container.
4. Gently shake out excess fluid.
5. Keep the magnet and plate (held securely together) **upside down**, and blot the plate on a short stack of paper towels.
6. Tap the magnet and plate on dry paper towels several times to assure equal evacuation from all wells.
7. Return the magnet and plate to the right-side up position on the benchtop.
8. Remove the plate from the magnet to add additional wash solution, and for incubation.
9. Repeat for any additional washes as needed.

Guidelines for Automated Plate Washers:

Some optimization of the automated plate washer set up may be required.

1. The program used for automated washing should include a 90 second incubation to allow the beads to immobilize on the bottom of the plate. After the beads are immobilized, the liquid may be aspirated using automated washing equipment. A suggested probe height of 4.8 mm is recommended.
2. For each successive wash step, refill the wells with washing solution, and remove the plate from the magnet. Allow the beads to incubate for 60 seconds before replacing the plate on the magnetic separator for 90 seconds and aspirating the well contents with the automated plate washing equipment.

Online tool

Go to **www.lifetechnologies.com/luminex** under Multiplex Solution Tools for further guidelines on automated plate washers and Luminex magnetic bead-based kits.

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Reverse pipetting recommendation

To reduce bubbles and loss of reagents due to residual fluid left in pipette tips, use the recommended reverse pipetting technique.
Note: Do not reverse pipette volumes <20 µL.

1. To reverse pipette, set the pipette to the appropriate volume needed.
2. Press the push-button slowly to the first stop and then press on past it. **Note:** the amount past the first stop will depend on the volume of liquid available to aspirate from.
3. Immerse the tip into the liquid, just below the meniscus.
4. Release the push-button slowly and smoothly to the top resting position to aspirate the set volume of liquid. Drag the tip up the side of the tube or reservoir to remove excess volume from the outside of the tips.
5. Place the end of the tip against the inside wall of the recipient vessel at an angle above the fluid level.
6. Press the push button slowly and smoothly to the first stop. Some liquid will remain in the tip, this should not be dispensed.
7. Remove the tip, keeping the pipette pressed to the first stop and return to step 3 above if reusing tips and contamination is not an issue.

Continued on next page

Assay Procedure, Continued

Analyte capture

1. Choose the filter bottom plate when washing with a vacuum manifold. Choose the flat bottom plate when washing manually with a magnetic separator or with automated magnetic bead washing equipment. An adhesive plate cover may be used to seal any unused wells; this will keep the wells dry for future use.
2. **The filter bottom plate requires pre-wetting before use in the assay.** Pre-wet the designated wells of the filter bottom plate by adding 200 µL of Working Wash Solution. Incubate the plate 30 seconds at room temperature. Aspirate the Working Wash Solution from the wells using the vacuum manifold.
3. The flat bottom plate may be used without the pre-wetting step.
4. Vortex the 1X Antibody Bead Solution for 30 seconds, then sonicate for at least 30 seconds immediately prior to use in the assay. **The magnetic beads settle rapidly. It is important that the 1X Antibody Bead Solution is well-mixed immediately prior to use.**
5. Pipette 25 µL of the 1X Antibody Bead Solution into each well. Once the beads are added to the plate, **keep the plate protected from light**.
6. Add 200 µL Working Wash Solution to the wells. Allow the beads to soak for 15–30 seconds.
7. Wash the wells two times, aspirating the Working Wash Solution at the end of each washing step **Note:** When using the filter plate, blot the bottom of the plate on clean paper towels to remove any residual liquid. Place the filter plate on a plate cover or non-absorbent surface before all incubations.
8. Pipette 50 µL Incubation Buffer into each well.
9. To wells designated for the standard curve, pipette 100 µL of appropriate standard dilution.
10. To the wells designated for the sample, pipette 50 µL Assay Diluent followed by 50 µL sample to each well or 50 µL in-house controls, if used.

Continued on next page

Assay Procedure, Continued

Analyte capture, continued

11. Cover microplate with an aluminum foil-wrapped plate cover. Incubate the plate for 2 hours at room temperature on an orbital shaker. Shaking should be sufficient to keep beads suspended during the incubation (500–600 rpm). Larger radius shakers will need a lower speed and smaller radius shakers will typically handle higher speeds without splashing.
12. Ten to fifteen minutes prior to the end of this incubation, prepare the biotinylated detector antibody, and then proceed to Analyte Detection, Step 1.

Preparing 1X Biotinylated Antibody

The Biotinylated Antibody is supplied as a **10X concentrate and must be diluted prior to use.**

To prepare a 1X Biotinylated Antibody stock, dilute 10 µL of 10X Biotinylated Antibody in 100 µL of Biotin Diluent per assay well. Each well requires 100 µL of the diluted Biotinylated Antibody. See table below for examples of volumes to combine.

Note: Dilution factor is 1:11 for extra pipetting volume.

Number of Wells	Vol. 10X Biotinylated Antibody Concentrate	Vol. Biotin Diluent
24	0.24 mL	2.4 mL
32	0.32 mL	3.2 mL
40	0.40 mL	4.0 mL
48	0.48 mL	4.8 mL
56	0.56 mL	5.6 mL
64	0.64 mL	6.4 mL
72	0.72 mL	7.2 mL
80	0.80 mL	8.0 mL
88	0.88 mL	8.8 mL
96	0.96 mL	9.6 mL

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Assay Procedure, Continued

Analyte detection

1. After the 2 hour capture bead incubation, remove the liquid from wells with the vacuum manifold (**filter bottom plate**), or with magnetic washing equipment (**flat bottom plate**).
2. Wash the plate by adding 200 µL of Working Wash Solution to the wells. Allow the beads to soak for 90 seconds. Remove the liquid with the vacuum manifold, or with magnetic washing equipment. Repeat this washing step for a total of 2 washes (The bottom of the filter plate should be blotted on clean paper towels to remove residual liquid after the second wash).
3. Add 100 µL of prepared 1X Biotinylated Detector Antibody (page 17) to each well and incubate the plate for 1 hour at room temperature on an orbital shaker. Shaking should be sufficient to keep the beads suspended during incubation (500–600 rpm).
4. Prepare the Luminex® 100™, 200™, FLEXMAP 3D® or MAGPIX® instrument during this incubation step. Refer to the Technical Data Sheet for all bead regions and standard concentration values.
5. Ten to fifteen minutes prior to the end of the detector incubation step, prepare the Streptavidin-RPE, and then proceed with Assay Reading, Step 1.

Continued on next page

Assay Procedure, Continued

Preparing Streptavidin-RPE

The Streptavidin-RPE is supplied as a **10X concentrate and must be diluted prior to use. Protect Streptavidin-RPE from light during handling.**

To prepare a 1X Streptavidin-RPE stock, dilute 10 µL of 10X Streptavidin-RPE in 100 of µL Streptavidin-RPE Diluent per assay well. Each well requires 100 µL of the diluted Streptavidin-RPE. See table below for examples of volumes to combine.

Note: Dilution factor is 1:11 for extra pipetting volume.

Number of Wells	Vol. 10X Streptavidin-RPE Concentrate	Vol. Streptavidin-RPE Diluent
24	0.24 mL	2.4 mL
32	0.32 mL	3.2 mL
40	0.40 mL	4.0 mL
48	0.48 mL	4.8 mL
56	0.56 mL	5.6 mL
64	0.64 mL	6.4 mL
72	0.72 mL	7.2 mL
80	0.80 mL	8.0 mL
88	0.88 mL	8.8 mL
96	0.96 mL	9.6 mL

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Assay Procedure, Continued

Assay reading

1. Remove the liquid from the wells using the vacuum manifold (**filter bottom plate**), or with magnetic washing equipment (**flat bottom plate**).
2. Wash the plate by adding 200 µL of Working Wash Solution to the wells. Allow the beads to soak for 90 seconds. Remove the liquid with the vacuum manifold, or with magnetic washing equipment. Repeat this washing step for a total of 2 washes (Blot the bottom of the filter plate on clean paper towels to remove residual liquid after the second wash).
3. Add 100 µL of prepared 1X Streptavidin-RPE to each well and incubate the plate for 30 minutes at room temperature on an orbital shaker. Shaking should be sufficient to keep the beads suspended during incubation (500–600 rpm).
4. Remove the liquid from wells with the vacuum manifold, or with magnetic washing equipment.
5. Wash the plate by adding 200 µL of Working Wash Solution to the wells. Allow the beads to soak for 90 seconds. Remove the liquid with the vacuum manifold, or with magnetic washing equipment. Repeat this washing step 2 more times for a total of 3 washes (Blot the bottom of the filter plate on clean paper towels to remove residual liquid after the second wash).
6. Add 125 µL of Working Wash Solution to each well. Shake the plate on an orbital shaker (500–600 rpm) for 2–3 minutes to resuspend the beads.

   Note: If the plate cannot be read on the day of the assay, cover and store the plate in the dark overnight at 2–8°C for reading the following day without significant loss of fluorescent intensity. Remove the Working Wash Solution from stored plates and add 125 µL fresh Working Wash Solution. Place the plate on an orbital shaker for 2–3 minutes at 500–600 rpm prior to analysis.
7. Uncover the plate and insert the plate into the XY platform of the Luminex® 100™, 200™, FLEXMAP 3D®, or MAGPIX® instrument, and analyze the samples.
8. Determine the concentration of samples from the standard curve using curve fitting software. The five parameter logistic algorithm with a weighted function ($1/y^2$) is recommended.

Instrument Setup

Luminex® 100™ and 200™

Helpful guidelines for Luminex® 100™ and 200™ instrument users with xPONENT® software.

- Assign the appropriate **Bead Region** (refer to the kit-specific technical data sheet) to each analyte.
- We recommend that the user **count 100 events/bead region**.
- Set **Sample Size** to 75 µL.
- For Life Technologies kits using MagPlex™ beads, we recommend that the doublet discriminator gates be set at 7800 –20,000 as the initial setting. Adjustment of these values may be required for individual instruments.
- Collect **Median Fluorescent Intensity (MFI)**.

Note: Use the default setting low PMT for the Luminex® 100™ and 200™ instruments.

Luminex® FLEXMAP 3D®

Helpful guidelines for Luminex® FLEXMAP 3D® instrument users.

- Assign the appropriate **Bead Region** (refer to the kit-specific technical data sheet) to each analyte.
- We recommend that the user **count 100 events/bead region**.
- Set **Sample Size** to 75 µL.
- For Life Technologies kits using MagPlex™ beads, we recommend that the doublet discriminator gates be set at 7800 –20,000 as the initial setting. Adjustment of these values may be required for individual instruments.
- Collect **Median Fluorescent Intensity (MFI)**.

Note: Use the default setting high PMT for the FLEXMAP 3D® instrument.

Instrument Setup, Continued

Luminex® MAGPIX®

Helpful guides for Luminex® MAGPIX® instrument users.

- Assign the appropriate **Bead Region** (refer to the kit-specific technical data sheet) to each analyte.
- We recommend that the user count **100 events/bead region**.
- Set **Sample Size** to 50 µL.
- The MAGPIX® instrument is designed use MagPlex™ beads.
- Collect **Median Fluorescent Intensity (MFI).**

Performance Characteristics and Limitations of the Procedure

Performance characteristics

Refer to analyte specific Technical Data Sheet for performance claims.

Procedure limitations

- Do not extrapolate the standard curve beyond the highest or lowest standard point; the dose-response and data collected in these regions may be non-linear and should be considered inaccurate. **Note:** In some cases, further dilution of the standard beyond 7 points may be possible to extend the low end of the standard curve.
- Dilute samples that are greater than the highest standard with Assay Diluent or appropriate matrix diluent; reanalyze these samples and multiply results by the appropriate dilution factor.
- **Samples are diluted in the assay 1:2 (50 µL of sample and 50 µL of diluent) relative to the standards. Be sure to account for this dilution factor during sample calculations.**
- The influence of various drugs, aberrant sera (hemolyzed, hyperlipidemic, jaundiced, etc.) and the use of biological fluids in place of serum, plasma and tissue culture supernatant samples have not been thoroughly investigated. The rate of degradation of analytes in various matrices may not have been investigated. The immunoassay literature contains frequent references to aberrant signals seen with some sera, attributed to heterophilic antibodies. Though such samples have not been seen to date, the possibility of this occurrence cannot be excluded.

Troubleshooting

Refer to the table below to troubleshoot problems encountered with the use of Multiplex Bead Kits on the Luminex® platform.

To troubleshoot problems with the Luminex® instrument, refer to the manual supplied with the instrument.

For more troubleshooting solutions, visit **www.lifetechnologies.com/luminex**.

Observation	Cause	Solution
In-house controls perform differently in subsequent assays	Incorrect concentration entered in data analysis software	The standard proteins included in the Bead Kits are calibrated to NIBSC preparations, whenever possible. This calibration assures lot-to-lot consistency in performance. However, the concentration of the reconstituted standards may vary with each new lot of standard. Therefore, it is important to check the concentration of the standard listed on the Technical Data Sheet, and to verify all concentration values entered into the data analysis software.
	Improper reconstitution or dilution of the standard	Check standard reconstitution and dilution as described on page 11.

Continued on next page

Troubleshooting, Continued

Observation	Cause	Solution
During washing steps, the vacuum manifold does not aspirate the liquid from wells of the filter plate	The filter plate is clogged	Dislodge the clog by gently pushing the pointed end of a 15 mL plastic conical tube into the bottom of the plate under the clogged well. This procedure clears the small opening in the plastic casing.
		Dislodge by placing the filter plate on a clean paper towel and use a gloved thumb or a 1 mL Pasteur pipette bulb to plunge the top of the clogged well.
		To prevent filter plate clogging, clarify samples by centrifugation at 1000 × *g* for 10 minutes prior to analysis. Some samples may also require filtration prior to analysis.
	Lack of a tight seal	Hold the filter plate firmly against the vacuum manifold to form a tight seal. If only a partial plate is being analyzed, cover the empty wells with a self-adhesive plate seal.
Leaky filter plate	Solution remains on the bottom of the wells after vacuum aspiration, causing wicking and leakage of well contents during next incubation	After final wash step and plate taps, use a clean absorbent towel to blot the bottom of the filter plate before addition of next liquid phase or data acquisition step.
	Filter plate membrane tearing	Excessive vacuum can cause the membrane to tear, resulting in antibody bead loss. Prevent any vacuum surge by opening and adjusting the vacuum on the manifold before placing the filter plate on the manifold surface.

Continued on next page

Troubleshooting, Continued

Observation	Cause	Solution
During data analysis, insufficient and/or erratic bead count is observed	Magnetic bead settling	Make sure that the rate of plate shaking is sufficient to keep the beads suspended during incubations and prior to analysis.
	Bead aggregation	Make sure to vortex the beads for 30 seconds and then sonicate the beads for at least 30 seconds prior to beginning the assay, to break up any bead aggregates.
		Empty wells and add fresh wash buffer. Shake for 2–3 minutes to resuspend the beads.
	Loss of beads due to the filter plate membrane tearing	To prevent membrane tearing, place pipette tips on the side of the well, rather than straight down onto the membrane when dispensing liquid into the wells.
		Turn the vacuum manifold on before placing the filter plate on the top to prevent vacuum surge. When evaluating a new vacuum manifold, adjust the vacuum force so that 3 seconds are required to empty 0.2 mL from the wells of a plate.
	Clog in instrument or probe	Remove probe, sonicate for 5 minutes, rinse the probe and reinstall. Run an unclog protocol. See instrument manual.
	Probe height set incorrectly	Readjust the instrument probe height. If it is too low, it could puncture the filter plate membrane. If it is too high, air could be pulled up with the liquid which may appear as bead fragments to the instrument.

Appendix

Technical Support

Obtaining support

For the latest services and support information for all locations, go to **www.lifetechnologies.com**

At the website, you can:

- Access worldwide telephone and fax numbers to contact Technical Support and Sales facilities
- Search through frequently asked questions (FAQs)
- Submit a question directly to Technical Support (**techsupport@lifetech.com**)
- Search for user documents, SDSs, vector maps and sequences, application notes, formulations, handbooks, certificates of analysis, citations, and other product support documents
- Obtain information about customer training
- Download software updates and patches

Life Technologies Corporation
7335 Executive Way
Frederick, MD USA 21704
Tel (Toll Free): 1 800 955 6288
E-mail: **techsupport@lifetech.com**

Safety Data Sheets (SDS)

Safety Data Sheets (SDSs) are available at **www.lifetechnologies.com/support**.

Certificate of Analysis

The Certificate of Analysis provides detailed quality control and product qualification information for each product. Certificates of Analysis are available on our website. Go to **www.lifetechnologies.com/support** and search for the Certificate of Analysis by product lot number, which is printed on the box.

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Life Technologies Corporation and/or its affiliate(s) warrant their products as set forth in the Life Technologies' General Terms and Conditions of Sale found on Life Technologies' website at **www.lifetechnologies.com/termsandconditions**. If you have any questions, please contact Life Technologies at **www.lifetechnologies.com/support**.

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Limited use label license: Fluorescently labeled microsphere beads

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References

The references below demonstrate the success customers achieve when using Multiplex Assays from Life Technologies. For a complete list, visit **www.lifetechnologies.com/luminex.**

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Protocol Summary

Note: Pre-wet step required only with the filter bottom plate.

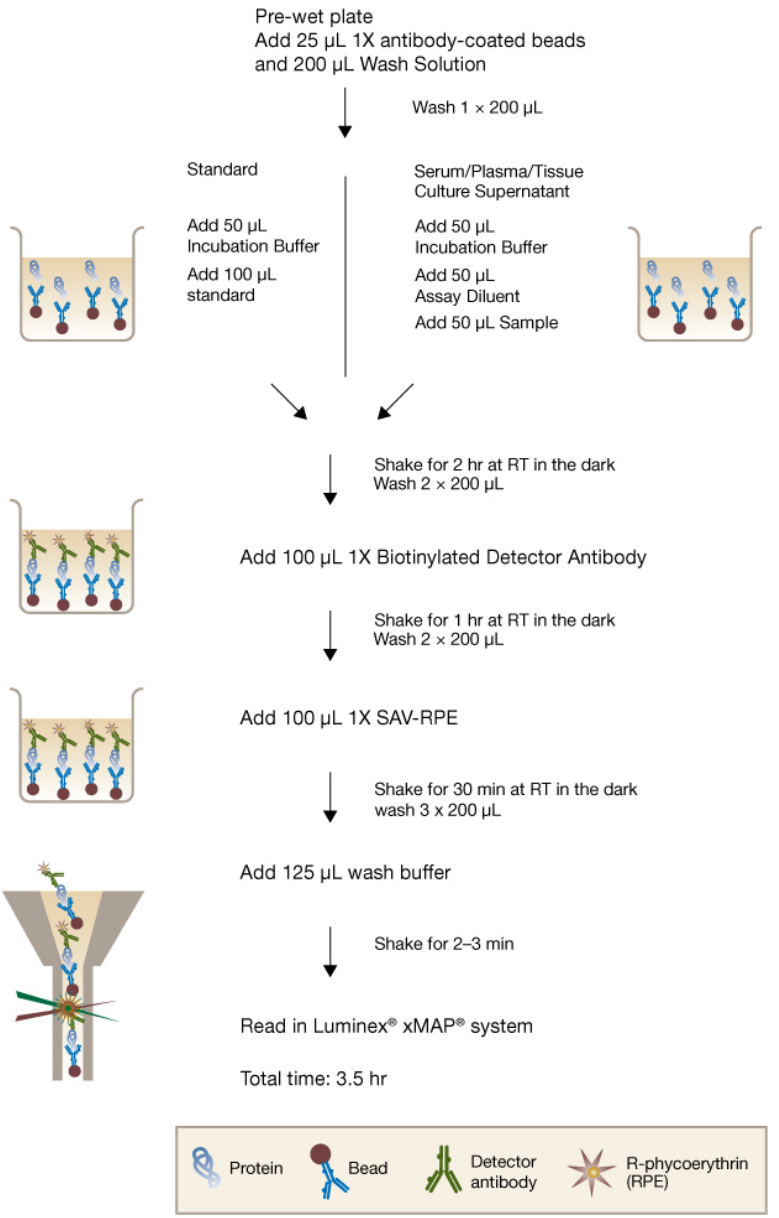

Plate Plan Template

Kit Catalog number: _____

Lot Number: _____

Plate ID: _____

Date: _____

	1	2	3	4	5	6	7	8	9	10	11	12
A												
B												
C												
D												
E												
F												
G												
H												

Explanation of Symbols

Symbol	Description	Symbol	Description
REF	Catalogue Number	LOT	Batch code
RUO	Research Use Only	IVD	*In vitro* diagnostic medical device
	Use by		Temperature limitation
	Manufacturer	EC REP	European Community authorised representative
[-]	Without, does not contain	[+]	With, contains
	Protect from light		Consult accompanying documents
	Directs the user to consult instructions for use (IFU), accompanying the product.		

9 April 2012

Notes

Notes

Headquarters
5791 Van Allen Way | Carlsbad, CA 92008 USA
Phone +1 760 603 7200 | Toll Free in USA 800 955 6288

For support visit
www.invitrogen.com/support or email techsupport@invitrogen.com

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