Human Tau (Total) ELISA Kit

Catalog nos. KHB0041
KHB0042

Pub. No. MAN0005237 **Rev** 2.0

Description

The Human (Hu) Tau (Total) ELISA Kit is a solid-phase sandwich Enzyme-Linked Immunosorbent Assay (ELISA) designed to detect and quantify the level of Human Tau in human cerebrospinal fluid (CSF), buffered solution, or cell culture medium. The assay will recognize both natural and recombinant Human Tau.

Contents and storage

The components included in the ELISA kit are listed below. Upon receipt, store the kit at 2°C to 8°C.

Components	Cat. no. KHB0041 96 tests	Cat. no. KHB0042 192 tests
Human Tau Antibody Coated Wells. 96 well plate.	1 plate	2 plates
Human Tau Biotin Conjugate. Contains 0.1% sodium azide.	11 mL	2 × 11 mL
Human Tau (Total) Standard, recombinant Human Tau-352 expressed in *E. coli*. Contains 0.1% sodium azide. Refer to vial label for quantity and reconstitution volume.	2 vials	4 vials
Wash Buffer Concentrate (25X).	100 mL	100 mL
Standard Diluent Buffer. Contains 0.1% sodium azide.	25 mL	2 × 25 mL
Streptavidin-HRP (100X). Contains 3.3 mM thymol.	0.125 mL	2 × 0.125 mL
Streptavidin-HRP Diluent. Contains 3.3 mM thymol.	25 mL	25 mL
Stabilized Chromogen, Tetramethylbenzidine (TMB).	25 mL	25 mL
Stop Solution.	25 mL	25 mL
Adhesive Plate Covers.	3	6

CAUTION! This kit contains materials with small quantities of sodium azide. Sodium azide reacts with lead and copper plumbing to form explosive metal azides. Upon disposal, flush drains with a large volume of water to prevent azide accumulation. Avoid ingestion and contact with eyes, skin and mucous membranes. In case of contact, rinse affected area with plenty of water. Observe all federal, state, and local regulations for disposal.

Materials required but not provided

- Distilled or deionized water
- Microtiter plate reader with software capable of measurement at or near 450 nm
- Plate washer–automated or manual (squirt bottle, manifold dispenser, or equivalent)
- Calibrated adjustable precision pipettes and glass or plastic tubes for diluting solutions

General Guidelines

- Review the **Procedural guidelines** and **Plate washing directions** in the *ELISA Technical Guide* available at **thermofisher.com/techresources** for details prior to starting the procedure.
- Reagents are lot-specific. Do not mix or interchange different reagent lots from various kit lots.

Prepare 1X Wash Buffer

1. Allow the Wash Buffer Concentrate (25X) to reach room temperature and mix to redissolve any precipitated salts.
2. Dilute 16 mL of Wash Buffer Concentrate (25X) with 384 mL of deionized or distilled water. Label as 1X Wash Buffer.
3. Store the concentrate and 1X Wash Buffer in the refrigerator. Use the diluted buffer within 14 days.

Sample preparation guidelines

- Collect samples in pyrogen/endotoxin-free tubes.
- Freeze samples after collection if samples will not be tested immediately. Avoid multiple freeze-thaw cycles of frozen samples. Thaw completely and mix well (do not vortex) prior to analysis.
- Avoid the use of hemolyzed or lipemic sera. If large amounts of particulate matter are present in the sample, centrifuge or filter sample prior to analysis.

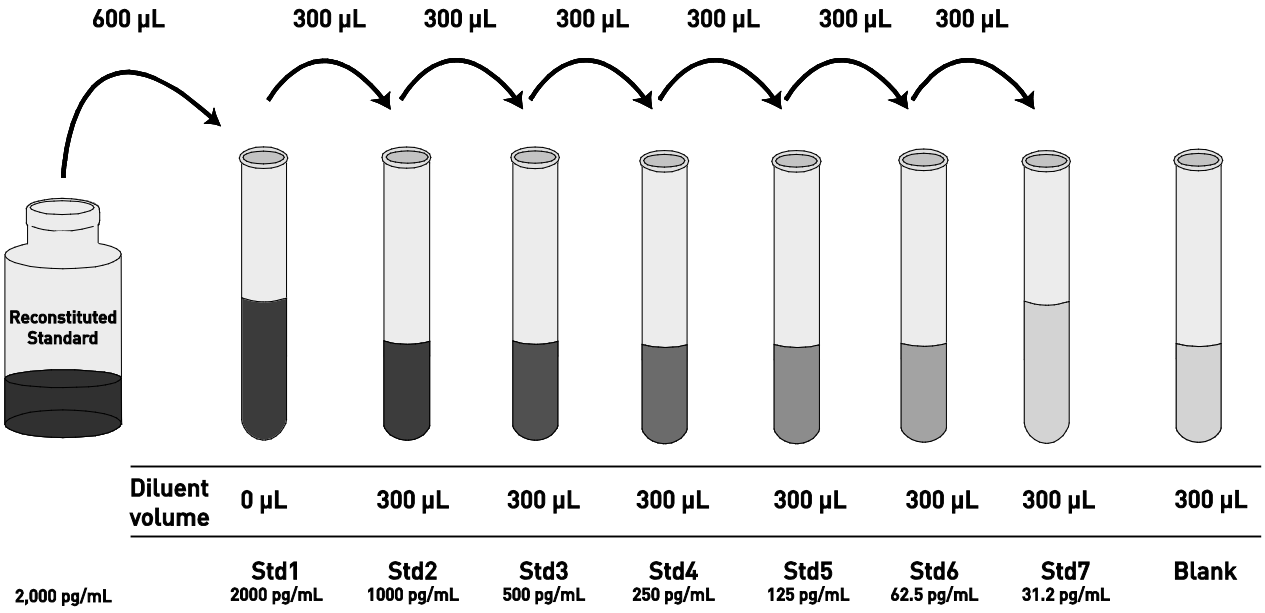
Dilute samples

- Dilute **serum**, **plasma**, **buffered solution**, and **cell culture medium** two-fold in Standard Diluent Buffer.
- Because conditions may vary, it is recommended that each investigator determine the optimal dilution to be used for each application.

Dilute standards

Note: Use glass or plastic tubes for diluting standards.

1. Reconstitute Human Tau Standard to 2,000 pg/mL with Standard Diluent Buffer. Refer to the standard vial label for instructions. Swirl or mix gently and allow the contents to sit for 10 minutes to ensure complete reconstitution. Label as 2,000 pg/mL Human Tau. **Use the standard within 1 hour of reconstitution**.
2. Add 600 µL Reconstituted Standard to one tube and label as 2,000 pg/mL Human Tau.
3. Add 300 µL Standard Diluent Buffer to each of 6 tubes labeled as follows: 1,000, 500, 250, 125, 62.5, and 31.2 pg/mL Human Tau.
4. Make serial dilutions of the standard as described below in the dilution diagram. Mix thoroughly between steps.
5. Discard any remaining reconstituted standard. Return the Standard Diluent Buffer to the refrigerator.



Prepare 1X Streptavidin-HRP solution

Note: Prepare Streptavidin-HRP within 15 minutes of usage.

The Streptavidin-HRP (100X) is in 50% glycerol, which is viscous. To ensure accurate dilution:

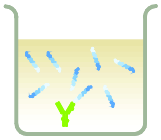
1. Allow the Streptavidin-HRP (100X) to reach room temperature and mix gently.
2. For each 8-well strip used in the assay, pipet 10 µL Streptavidin-HRP (100X) solution, wipe the pipette tip with a clean absorbent paper to remove any excess solution, and dispense the solution into a tube containing 1 mL of HRP Diluent. Mix thoroughly.
3. Return the unused Streptavidin-HRP (100X) solution to the refrigerator.

ELISA procedure

Allow all reagents to reach room temperature before use. Mix all liquid reagents prior to use. **Total assay time is 4 hours.**

IMPORTANT! Perform a standard curve with each assay.

Determine the number of 8-well strips required for the assay. Insert the strips in the frames for use. Re-bag any unused strips and frames, and store at 2 to 8°C for future use.

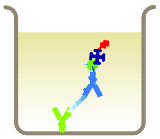


Bind antigen

1. Add 100 µL of Standard Diluent Buffer to zero wells except the chromogen blanks.
2. Add 100 µL of standards to the appropriate wells. For all samples (CSF, buffered solution, cell culture medium and controls), add 50 µL of Standard Diluent Buffer to each well followed by 50 µL of sample.
3. Cover the plate with the plate cover and incubate for 2 hours at room temperature.
4. Thoroughly aspirate the solution and wash wells 4 times with 1X Wash Buffer.

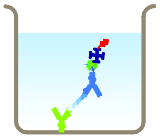
Add Biotin Conjugate

5. Add 100 µL Human FGF-b Biotin Conjugate solution into each well except the chromogen blanks.
6. Tap side of the plate to mix. Cover the plate with plate cover and incubate for 1 hour at room temperature.
7. Thoroughly aspirate the solution and wash wells 4 times with 1X Wash Buffer.



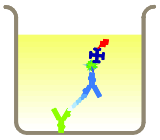
Add Streptavidin-HRP

8. Add 100 µL Streptavidin-HRP (see page 2) into each well except the chromogen blanks.
9. Cover the plate with the plate cover and incubate for 30 minutes at room temperature.
10. Thoroughly aspirate the solution from the wells and wash wells 4 times with 1X Wash Buffer.



Add chromogen

11. Add 100 µL Stabilized Chromogen to each well. The substrate solution will begin to turn blue.
12. Incubate for 30 minutes at room temperature **in the dark**.
    Note: TMB should not touch aluminum foil or other metals.



Add stop solution

13. Add 100 µL of Stop Solution to each well. Tap side of the plate gently to mix. The solution in the wells changes from blue to yellow.

Read the plate and generate the standard curve

1. Read the absorbance at 450 nm. Read the plate within 2 hours after adding the Stop Solution.
2. Use curve-fitting software to generate the standard curve. A four parameter algorithm provides the best standard curve fit. Optimally, the background absorbance may be subtracted from all data points, including standards, unknowns and controls, prior to plotting.
3. Read the concentrations for unknown samples and controls from the standard curve. Multiply value(s) obtained for sample(s) by the appropriate factor to correct for the sample dilution.
4. Dilute samples producing signals greater than that of the highest standard in Standard Diluent Buffer and reanalyze. Multiply the concentration by the appropriate dilution factor.

Performance characteristics

Standard curve (example)

The following data were obtained for the various standards over the range of 0-2,000 pg/mL Human Tau.

Standard Human Tau (pg/mL)	Optical Density (450 nm)
2000	2.96
1000	1.65
500	0.93
250	0.54
125	0.35
62.5	0.26
31.2	0.20
0	0.15

Specificity

Buffered solutions of a panel of substances at 20,000 pg/mL were assayed with the Human Tau kit. The following substances were tested and found to have no cross-reactivity: **Human** b Amyloid 1-40, b Amyloid 1-42, a-Synuclein. **Human** b-Synuclein and **Mouse** tau, measured in mouse brain homogenate, showed less than 1.3% cross-reactivity.

Sensitivity

The minimum detectable dose of Human Tau is <10 pg/mL. This was determined by adding two standard deviations to the mean O.D. obtained when the zero standard was assayed 40 times, and calculating the corresponding concentration.

Performance characteristics, continued

Intra-assay precision

Samples of known Human Tau concentration were assayed in replicates of 16 to determine precision within an assay.

Parameters	Sample 1	Sample 2	Sample 3
Mean (pg/mL)	58	236	931
SD	3.4	7.9	40.8
%CV	5.9	3.4	4.4

SD = Standard Deviation; CV = Coefficient of Variation

Inter-assay precision

Samples were assayed 48 times in multiple assays to determine precision between assays.

Parameters	Sample 1	Sample 2	Sample 3
Mean (pg/mL)	60	243	994
SD	5.9	12.1	44.8
%CV	9.9	5.0	4.5

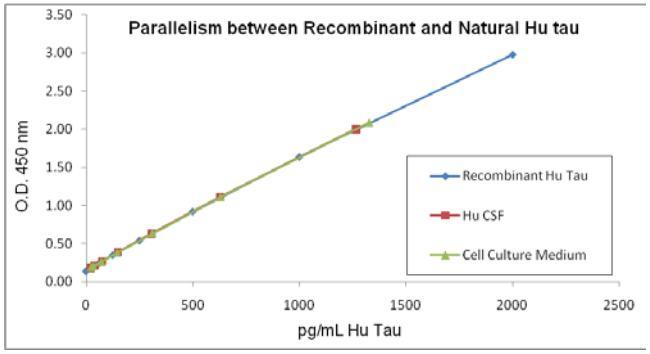
SD = Standard Deviation; CV = Coefficient of Variation

Recovery

The recovery of Human Tau added to human cerebrospiral fluid (CSF) averaged 101%. The recovery of Human Tau added to tissue culture medium containing 1% fetal bovine serum averaged 97%, while the recovery of Human Tau added to tissue culture medium containing 10% fetal calf serum averaged 96%.

Parallelism

Human CSF and tissue culture medium containing 10% fetal bovine serum were spiked with recombinant Human Tau and serially diluted in *Standard Diluent Buffer*. The optical density of each dilution was plotted against the standard curve. Parallelism was demonstrated by the figure below and indicated that the standard accurately reflects Tau content in samples.



Linearity of dilution

Human CSF and tissue culture medium containing 10% fetal bovine serum were spiked with Human Tau and serially diluted in *Standard Diluent Buffer* over the range of the assay. Linear regression analysis of samples versus the expected concentration yielded a correlation coefficient of 0.99 for the spiked Human CSF and 0.96 for the spiked culture medium containing 10% fetal bovine serum.

Dilution	CSF			Cell Culture		
	Measured (pg/mL)	Expected (pg/mL)	% Expected	Measured (pg/mL)	Expected (pg/mL)	% Expected
1/2	1265	—	—	1327	—	—
1/4	629	633	99	630	664	95
1/8	305	314	97	311	315	99
1/16	150	153	98	146	156	94
1/32	75	75	100	71	73	98
1/64	39	38	104	33	36	93

Important licensing information

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Product label explanation of symbols and warnings

REF	Catalog Number	LOT	Batch code		Temperature limitation		Use by		Manufacturer		Consult instructions for use		Caution, consult accompanying documents

For support visit thermofisher.com/support or contact techsupport@lifetech.com.

thermofisher.com

23 November 2015