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## Warnings

Reagents contain sodium azide. Under acidic conditions sodium azide yields hydrazoic acid, which is extremely toxic. Azide compounds should be diluted with running water before discarding. These precautions are recommended to avoid deposits in plumbing where explosive conditions may develop.

## 1. Description

This product is for research use only.

<b>Components</b>	<b>10 Whole Blood Columns</b> columns and plungers, sterile packed.
	<b>50 mL Whole Blood Column Elution Buffer</b> contains stabilizer and 0.09% sodium azide.
<b>Storage</b>	Store Whole Blood Columns dry at 10–35 °C and protected from light at room temperature. Store Whole Blood Column Elution Buffer at 2–8 °C. The expiration date is indicated on the box label. Do not use after this date.

### 1.1 Background information

The patented MACS® Column Technology is based on the use of MACS MicroBeads, MACS Columns and MACS Separators. Whole Blood Columns have been developed for the gentle isolation of cells labeled with MACS Whole Blood MicroBeads. As MACS MicroBeads are extremely small, superparamagnetic particles, a high-gradient magnetic field is required to retain the labeled cells. Whole Blood Columns contain an optimized matrix to generate this strong magnetic field when placed in a permanent magnet such as the MidiMACS™ Separator, QuadroMACS™ Separator, VarioMACS™ Separator, SuperMACS™ II Separator, or MultiMACS™ Cell24 Separator Plus.

## 1.2 Technical specifications

	Max. volume of whole blood per single column
Manual use	0.25–15 mL
Use with MultiMACS Cell24 Separator Plus	10 mL

- Columns are “flow stop” and do not run dry.
- Void volume: 600 µL. Reservoir volume: 7.5 mL.
- Typical flow rate for phosphate-buffered saline (PBS) containing 0.5% bovine serum albumin (BSA): 2.7–3.3 mL/min.
- Whole Blood Columns are for single use only.
- Use Whole Blood Elution Buffer to elute positive cell fraction from column.

## 1.3 Applications

Whole Blood Columns have been developed for positive selection of leukocyte subsets from anticoagulated whole blood labeled with MACS Whole Blood MicroBeads in combination with a MACS Separator.

▲ Do not use Whole Blood Columns in combination with magnetic particles other than MACS Whole Blood MicroBeads. Magnetic forces in the column are very high and may damage biological material if other beads are used.

▲ To remove clumps and to prevent aggregates in the sample, pass cells through 30 µm nylon mesh (Pre-Separation Filters, 30 µm, # 130-041-407) before separation.

## 1.4 Reagent and instrument requirements

- Separation buffer: Prepare a solution containing phosphate-buffered saline (PBS), pH 7.2, 0.5% bovine serum albumin (BSA), and 2 mM EDTA by diluting MACS BSA Stock Solution (# 130-091-376) 1:20 with autoMACS® Rinsing Solution (# 130-091-222). Keep buffer cold (2–8 °C).
  - ▲ **Note:** The recommended buffer is PBS supplemented with EDTA and BSA. The suitability of other buffers has to be tested experimentally.
  - ▲ **Note:** Use degassed buffer only! Degas buffer by applying vacuum, preferentially with buffer at room temperature. Excessive gas in running buffer will form bubbles in the matrix during separation. This may lead to clogging of the column and decrease the quality of separation.
- MACS Whole Blood MicroBeads for magnetic labeling of cells.
- MidiMACS Separator, QuadroMACS Separator, VarioMACS Separator, SuperMACS II Separator, or MultiMACS Cell24 Separator Plus.
- LS Column Adapter (# 130-090-544) for use with VarioMACS Separator or Adapter for MS, LS, and LD Columns for use with SuperMACS II Separator.

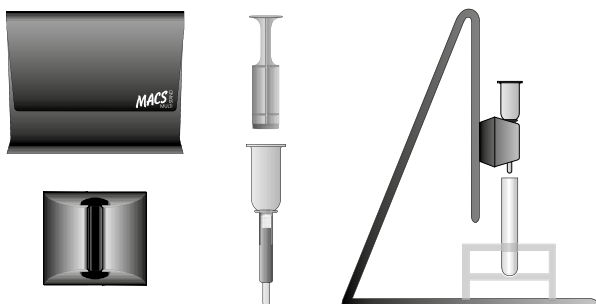
- MACS Acrylic Tube Rack (# 130-041-406) or MACS 15 mL Tube Rack (# 130-091-052).
- (Optional) Pre-Separation Filters, 30  $\mu\text{m}$  (# 130-041-407) to remove cell clumps.
- (Optional) Red Blood Cell Lysis Solution (10 $\times$ ) (# 130-094-183).

## 2. Use of Whole Blood Column Kit with manual separators

### 2.1 Reagent and column preparation

1. Adjust Whole Blood Column Elution Buffer to room temperature.
2. Insert Whole Blood Column with the column wings to the front into an MACS Separator.

#### A) Use with MidiMACS™ or QuadroMACS™ Separator



Attach MidiMACS™ Separator or QuadroMACS™ Separator to the MultiStand and place Whole Blood Column in the separator. Place a collection tube under the Whole Blood Column.

▲ **Note:** Check that the ejection blocks in the gap of the magnet are attached before placing the MACS Column into the magnetic field of the MidiMACS or QuadroMACS Separator.

▲ **Note:** Be careful when attaching the QuadroMACS Separator to the MultiStand to avoid trapping your fingers (for details refer to the QuadroMACS Starting Kit data sheet).

#### B) Use with VarioMACS™ or SuperMACS™ II Separator

For use of Whole Blood Columns with the VarioMACS™ or SuperMACS™ II Separator, please refer to the respective data sheet.

3. Prepare Whole Blood Column by rinsing with **separation buffer**: apply 3 mL of **separation buffer** on top of the column and let the buffer run through. Whole Blood Columns are “flow stop” and do not run dry.
4. Discard effluent and change collection tube. The Whole Blood Column is now ready for magnetic separation.

▲ **Note:** Use column immediately after filling to avoid formation of air bubbles caused by warming up. Do not store columns after filling.

▲ **Note:** The time for filling the column with buffer is dependent on the storage conditions, temperature, and humidity. Therefore, the time may vary from a few seconds to several minutes. This filling time has no influence on the quality of the separation.



## 2.2 Magnetic separation using Whole Blood Columns

▲ For details on magnetic labeling, refer to the MACS Cell Separation Reagent data sheets.

▲ Always wait until the column reservoir is empty before proceeding to the next step.

1. Apply magnetically labeled cell suspension onto the prepared Whole Blood Column. Collect flow-through containing unlabeled cells.

▲ **Note:** The reservoir of the Whole Blood Column contains a maximum of 7.5 mL. Samples greater than 7.5 mL should be applied in aliquots to the column.

2. Wash Whole Blood Column with 3 $\times$ 3 mL **separation buffer**. Collect unlabeled cells that pass through and combine with the effluent from step 1.

▲ **Note:** Perform washing steps by adding buffer aliquots only when the column reservoir is empty.

3. Remove Whole Blood Column from the separator and place it on a new collection tube.

4. Pipette 5 mL **Whole Blood Column Elution Buffer** onto the Whole Blood Column. Immediately flush out the magnetically labeled cells by firmly pushing the plunger into the column.

5. (Optional) To increase the purity of the magnetically labeled fraction the eluted fraction can be enriched over a new, freshly prepared MS Column (# 130-042-201, for up to 10<sup>7</sup> magnetically labeled cells) or LS Column (# 130-042-401, for up to 10<sup>8</sup> magnetically labeled cells).

Remaining erythrocytes can be lysed using Red Blood Cell Lysis Solution (10 $\times$ ) (# 130-094-183).

## 3. Use of the Whole Blood Columns with the MultiMACS™ Cell24 Separator Plus

Up to 9 Whole Blood Columns can be used in a single run with the Single-Column Adapter.

For further details please refer to the MultiMACS™ Cell24 Separator Plus user manual.

### Warranty

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