

ZOOM® Strip pH 9-12

Cat. no. ZM0017

Store at -20°C

 $70 \, \mathrm{cm}$

Introduction

The ZOOM® Strip pH 9-12 is a pre-cast immobilized pH gradient (IPG) gel cast on a plastic backing. ZOOM® Strips are easy to use and produce reproducible pH gradients.

ZOOM® Strips pH 9-12 offer a linear pH range of 9-12 that is ideal for isoelectric focusing (IEF) and analysis of basic proteins such as nucleic acid binding proteins, transcription factors, and ribosomal proteins.

Specifications Length of the gel:

| Length of the gen | 7.0 СП |
|----------------------------|-------------------|
| Length of the ZOOM® Strip: | 7.7 cm |
| Gel thickness: | 0.5 mm |
| Width of the ZOOM® Strip: | 3.3 mm |
| pH range: | 9-12 linear |
| Storage: | Store at -20°C |
| Stability: | 3 months at -20°C |
| Number of ZOOM® Strips | 12/package |
| | |

The anode (acidic) end is marked as (+) and cathode (basic) end as (-). Each ZOOM® Strip is individually numbered for easy identification.

Amount of Protein

ZOOM® Strips pH 9-12 are specifically formulated to allow increased protein loads. We recommend loading 50-100 μ g (for silver staining) or 100-200 μ g (for Coomassie® staining) of total protein per ZOOM® Strip pH 9-12.

Part No. ZM0017.pps Rev. Date: 27 May 2004

This product is distributed for laboratory research only. CAUTION: Not for diagnostic use. The safety and efficacy of this product in diagnostic or other clinical uses has not been established.

Preparing Samples

To obtain the best results with ZOOM® Strips pH 9-12, use the sample preparation protocol described below. It is important to perform reduction and alkylation prior to IEF to reduce horizontal streaking.

- Prepare 100X Protease Inhibitor Cocktail by dissolving 1 Protease Inhibitor Cocktail tablet (Roche cat. no. 1873580) in 500 μl 1.1X ZOOM® 2D Protein Solubilizer 1 or 2. Mix well.
- 2. Prepare the Lysis Buffer fresh, just prior to use. You will need 950 μ l Lysis Buffer for each 1 ml of cell/tissue lysate.

| 1.1X ZOOM® 2D Protein Solubilizer 1 or 2 | 909 µl |
|--|--------|
| (cat. nos. ZS10001 or ZS10002) | |
| 1 M Tris Base | 3 µl |
| 100X Protease Inhibitor Cocktail from Step 1 | 10 μl |
| 2 M DTT | 10 μl |
| Deionized water | 18 µl |
| N.C. 11 1 () (2) | |

Mix well and store on ice until use.

- 3. To 50 mg (wet weight) minced tissue or 50 μ l packed *E. coli* (1 x 10¹⁰) cells, add 950 μ l chilled Lysis Buffer from Step 2.
- 4. Sonicate sample on ice for 5-10 rounds of 15 seconds each at ~50% power with cooling samples on ice between sonications.
- 5. Check the pH of the solution. The pH should be 8.4-9.0. Adjust the pH with 1 M Tris Base if needed.
- 6. Incubate on a rotary shaker for 10-15 minutes at room temperature.
- 7. Add $5 \mu l$ 99% N,N-Dimethylacrylamide (DMA, from Aldrich cat. no. 27413-5) to the lysate for alkylation.
- 8. Incubate on a rotary shaker for 30 minutes at room temperature.
- 9. Add 10 μl 2 M DTT to quench any excess DMA.
- 10. Centrifuge at 16,000 x g for 20 minutes at 4°C. Use supernatant for IEF (next page) or aliquot into small volumes and store at -80°C.

Diluting Samples for IEF

Dilute the lysate from Step 10, previous page, for use with the ZOOM® IPGRunner $^{\text{M}}$ System as described below. You will need 140 μ l diluted sample per ZOOM® Strip.

| Lysate from Step 10, previous page | 9 µl |
|---|-----------|
| 1.1X ZOOM® 2D Protein Solubilizer 1 or 2 | 128 µl |
| 2 M DTT | 1.4 µl |
| ZOOM® Carrier Ampholytes, pH 9-11 (cat. no. ZM0024) | 0.4 μl |
| Trace Bromophenol Blue | |
| Deionized water | to 140 ul |

Rehydrating ZOOM® Strips

ZOOM® Strips are supplied dry and are rehydrated in the ZOOM® IPGRunner™ Cassette (cat. no. ZM0003). For detailed instructions on rehydration and assembling the ZOOM® IPGRunner™ Mini-Cell (cat. no. ZM0001), see the ZOOM® IPGRunner™ System Manual (available from our Web site at www.invitrogen.com).

- After diluting your sample as described above, load 140 µl of the diluted sample into each Enclosed Channel of the ZOOM[®] IPGRunner™ Cassette. Leave unused channels empty.
- Peel a ZOOM® Strip pH 9-12 away from the card backing and slide the acidic end (+) of the strip into the cassette channel until the acidic end of each strip touches the end of the channel slot.
- Seal all Sample Loading Wells (including unused wells) with Sealing Tape (supplied with ZOOM® IPGRunner™ Cassettes).
- 4. Incubate ZOOM® Strips in the ZOOM® IPGRunner™ Cassette for 1 hour at room temperature. Then assemble the ZOOM® IPGRunner™ Mini-Cell (see ZOOM® IPGRunner™ Manual) and perform isoelectric focusing (see page 4).

Run Conditions for ZOOM® Strips pH 9-12

To obtain the best results, we recommend using ZOOM® Strips pH 9-12 with the ZOOM® IPGRunner $^{\text{TM}}$ Mini-Cell (cat. no. ZM0001).

- After rehydrating ZOOM® Strips, assemble the ZOOM® IPGRunner™ Mini-Cell as described in the ZOOM® IPGRunner™ Manual.
- 2. Perform IEF using an appropriate protocol as described below:

| Voltage Ramp | Step Voltage |
|--------------------------------|-----------------------|
| 175 V for 15 minutes | 200 V for 20 minutes |
| 175-2000 V ramp for 45 minutes | 450 V for 15 minutes |
| 2000 V for 1 hour | 750 V for 15 minutes |
| | 2000 V for 60 minutes |

 After the run is complete, disassemble the ZOOM® IPGRunner™ Mini-Cell as described in the ZOOM® IPGRunner™ Manual

Product Qualification

ZOOM® Strips pH 9-12 are qualified by subjecting a mixture of proteins to isoelectric focusing under standard focusing conditions. The strips are stained and visualized for proper resolution and migration of protein bands

Limited Use Label License

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