# Platinum<sup>®</sup> Tfi DNA Polymerase

### Cat. no: 73684-050

## Conc. 5 U/µl

## Kit Size: 500 units Store at -20°C (non-frost-free)

## Description

Platinum<sup>®</sup> *Tfi* DNA Polymerase is recombinant *Tfi* DNA polymerase complexed with a proprietary antibody mix that inhibits polymerase activity at ambient temperature, allowing room-temperature reaction setup. Activity is restored after the initial denaturation step in PCR cycling at 94°C, providing an automatic "hot start" for increased specificity, sensitivity, and yield.

*Tfi* DNA Polymerase is purified from *E. coli.* expressing cloned mutants of the *Thermus filiformis* DNA polymerase gene. This enzyme has both  $5' \rightarrow 3'$  polymerase and  $5' \rightarrow 3'$  exonuclease activity, but lacks  $3' \rightarrow 5'$  exonuclease activity. *Tfi* DNA polymerase is heat-stable and will synthesize DNA at elevated temperatures from single-stranded templates in the presence of a primer (Shandilya et al., 2004).

Platinum<sup>®</sup> *Tfi* DNA Polymerase can be used in protocols that currently use Platinum<sup>®</sup> *Taq* DNA Polymerase without modification. PCR performance is comparable to that of Platinum<sup>®</sup> *Taq* in yield, specificity, fidelity, and robustness. Like Platinum<sup>®</sup> *Taq*, Platinum<sup>®</sup> *Tfi* DNA Polymerase has a nontemplate-dependent terminal transferase activity that adds a single deoxyadenosine (A) to the 3' ends of PCR products.

Amount
100 µl
$4 \times 1.3$ ml
1 ml

Part no. 73684.pps

Rev. date: 30 June 2006

## **Guidelines for PCR**

General PCR parameters and troubleshooting information are documented in Innis, et al (Innis et al., 1990). PCR reactions should be assembled in a DNA-free environment using clean, dedicated automatic pipettors and aerosol resistant barrier tips. Always keep the control DNA and other templates to be amplified isolated from the other components.

## **General Recommendations for PCR Optimization**

The protocol on the following page provides general guidelines for PCR amplification. Optimal reaction conditions—including incubation times and temperatures, and amounts of polymerase, primers, MgCl<sub>2</sub>, and template DNA—may vary.

- For genomic DNA, 1.0 unit of Platinum<sup>®</sup> *Tfi* DNA Polymerase is sufficient for amplifying most targets less than 1kb. Increasing the amount of enzyme to 2.0 units may improve yield.
- For plasmid DNA, 1.0 unit is optimal.
- A general scheme for PCR optimization should start with adjusting the annealing temperature. The optimal annealing temperature should be  $5-10^{\circ}$  lower than the  $T_m$  of the primers used. For higher specificity, it may be necessary to gradually increase the annealing temperature in steps of  $2-3^{\circ}$ C.

## Unit Definition

One unit of Platinum<sup>®</sup> *Tfi* DNA Polymerase incorporates 10 nmol of deoxyribonucleotide into acid-precipitable material in 30 minutes at 74°C.

### Basic PCR Protocol

Due to Platinum<sup>®</sup> *Tfi* DNA Polymerase's "hot-start" capability, the reaction can be set up at room temperature.

1. Program the thermal cycler as follows (note that the annealing temperature will vary depending on the T<sub>m</sub> of your primers):

Initial denaturation: 94°C for 2 minutes

25-40 cycles of:

Denaturation:  $94^{\circ}$ C for 15–30 seconds Annealing: T<sub>m</sub> of primers minus 5–10°C for 30 seconds Extension:  $68^{\circ}$ C for 1 minute per kb of PCR product

Final extension: 68°C for 10 minutes

 Add the following components to a DNase/RNase-free microcentrifuge tube. For multiple reactions, prepare a master mix of common components to minimize reagent loss and enable accurate pipetting.

<u>Component</u>	Volume	<b>Final Concentration</b>
5X Platinum <sup>®</sup> Tfi Reaction Buffer	10 µl	1X
10 mM dNTP mix, PCR grade	1 µl	200 µM each
50 mM MgCl <sub>2</sub>	1.5 µl	1.5 mM
Primer mix (10 µM each)	1 µl	0.2 μM each
Template DNA	≥ 1 µl	as required
Platinum <sup>®</sup> Tfi DNA Polymerase	0.2–0.4 µl	1–2 units*
Autoclaved distilled water	to 50 µl	n/a

\*May use up to 2 units for genomic DNA. See note on previous page.

- 3. Cap the tube, tap gently to mix, and centrifuge briefly to collect the contents.
- 4. Place the tube in the thermal cycler and run the program from Step 1. After cycling, maintain the reaction at 4°C. Samples can be stored at -20°C until use.
- Analyze the amplification products by agarose gel electrophoresis. We recommend using E-Gel<sup>®</sup> 1.2% gels and TrackIt<sup>™</sup> 100 bp or 1kb Plus DNA ladders (see Additional Products on page 4).

#### **Quality Control**

Platinum<sup>®</sup> *Tfi* DNA Polymerase is evaluated in a DNA polymerization activity assay that measures the percent of polymerase inhibition versus an uninhibited control. Platinum<sup>®</sup> *Tfi* DNA Polymerase is also functionally tested for amplification and the absence of double- and single-stranded endonuclease activity, as well as the absence of contaminating exonuclease activity.

#### Additional Products

Product	Amount	Catalog no.
10 mM dNTP Mix, PCR Grade	100 µl	18427-013
10 mM dNTP Mix, PCR Grade	1 ml	18427-088
E-Gel® 1.2% Starter Pak	6 gels plus PowerBase™	G6000-01
E-Gel® 1.2% 18-Pak	18 gels	G5018-01
TrackIt <sup>™</sup> 100 bp DNA Ladder	100 applications	10488-058
TrackIt <sup>™</sup> 1kb Plus DNA Ladder	100 applications	10488-085

#### References

- Innis, M. A., Gelfand, D. H., Sninsky, J. J., and White, T. S. (eds) (1990) PCR Protocols: A Guide to Methods and Applications, Academic Press, San Diego, CA
- Shandilya, H., Griffiths, K., Flynn, E. K., Astatke, M., Shih, P. J., Lee, J. E., Gerard, G. F., Gibbs, M. D., and Bergquist, P. L. (2004) Extremophiles 8, 243-251

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