



CAL-LYSE[™] Lysing Solution

Whole blood lysing solution for flow cytometric applications

Catalog Numbers GAS-010, GAS-010S-100

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Introduction

Kit Contents and Storage

Sufficient Cal-Lyse[™] Lysing Solution is provided for the lysis of 250 samples **Reagent provided** (GAS-010) or 1000 samples (GAS-010S-100) of anticoagulated whole blood. Cat. no. Amount Storage GAS-010 25 mL (250 tests) Room temperature. Do not freeze. GAS-010S-100 100 mL (1000 tests) Room temperature. Do not freeze. The composition of the lysing solution includes the following ingredients: Polyvinylpyrrolidone Ethylene-bis (oxyrthylenenitruol) -tetra acetic acid Sodium phosphate Formaldehyde Deionized water Cal-Lyse[™] Lysing Solution contains formaldehyde as a fixative. Formaldehyde Warning is toxic, allergenic and a suspected carcinogen. Avoid ingestion, inhalation and contact with eyes, skin and clothing. Storage Store at room temperature. Do not freeze. **Evidence of** The reagent should not be used if any evidence of deterioration such as cloudiness or substantial loss of reactivity is observed. deterioriation The normal appearance of Cal-Lyse[™] Lysing Solution solution is a clear, 0 particulate-free liquid. Do not use this reagent if discoloration occurs or a visible precipitate forms. 0 We recommend periodically checking the reagent against your standard. Cal-Lyse[™] is a lysing solution formulated for lysis of erythrocytes in samples of Intended use anticoagulated human blood. Cal-Lyse[™] Lysing Solution is intended to be used as an aid in the enumeration of leukocytes from various human biological sources including blood and bone marrow that have been stained with monoclonal antibodies for analysis using flow cytometric methods. Results should be put within the context of other diagnostic tests, clinical history by a certified professional. Important: Cal-Lyse[™] Lysing Solution is not a concentrate, and should not be diluted prior to use. For optimal results, use only as directed. For In Vitro Diagnostic Use.

About the System

ic antigenic ochrome- he surfaces			
To perform analysis of leukocytes in the sample, cells bearing specific antigenic determinants are identified by incubating, blood samples with fluorochrome-conjugated monoclonal antibodies. Monoclonal antibodies bind to the surfaces of viable blood cells that express the corresponding antigens.			
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Methods

Procedure	
Statement of warning	 Do not pipet by mouth. Samples should be handled as if capable of transmitting infection. Appropriate disposal methods should be used.
	 Do not use reagent beyond the stated expiration date of the product.
	• Cal-Lyse [™] Lysing Solution is not a concentrate, and should not be diluted prior to use. For optimal results, use only as directed.
	• Deviations from the recommended procedure enclosed within this product insert may invalidate the results of testing
	• The reagent should not be used if any evidence of deterioration such as cloudiness or substantial loss of reactivity is observed.
Required materials that are	• Centrifuge with swinging bucket rotor capable of $1000 \times g$
not supplied	Vortex mixer
	• 3–5 mL conical tubes
	 Micropipette capable of dispensing 100 µL volumes
	Blood collection tubes with anticoagulant (EDTA or heparin)
	Phosphate Buffered Saline (PBS)
	Flow cytometer

Procedure, Continued

Sample preparation	Two methods of sample preparation are provided. The first procedure includes a wash step to remove red cell debris and any unbound antibody, while the second procedure does not include the wash step. Note : Failure to completely remove erythrocytes from blood samples may interfere with the subsequent flow cytometric enumeration of antibody- stained leukocytes.				
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Antibody staining	1.	Collect blood into an appropriate anticoagulant (EDTA or heparin).			
and wash lysis	2.	Pipette 100 µL of thoroughly mixed blood into a conical tube.			
procedure	3.	Add antibody as indicated in the manufacturer's package insert to appropriately labeled tubes from step 2. Mix gently.			
	4.	Incubate all tubes for 15 minutes at room temperature ($22 \pm 3^{\circ}$ C) in the dark.			
	5.	Add 100 μL of Cal-Lyse [™] Lysing Solution to all tubes.			
	6.	Incubate all tubes for 10 minutes at room temperature ($22 \pm 3^{\circ}$ C) in the dark.			
	7.	Add 1.0 mL of deionized water to all tubes. Immediately vortex tubes.			
	8.	Incubate all tubes for 10 minutes at room temperature			
	9.	Add 3–4 mL of distilled water to each tube, and vortex. Incubate tubes for 5–10 minutes at room temperature.			
	10.	Centrifuge tubes for 5 minutes at $300 \times g$. Remove supernatant.			
	11.	Resuspend the cells in all tubes in 1 mL of PBS or Sheath Fluid.			
	12.	Analyze on a flow cytometer according immediately, or store samples at 2°C–8°C in the dark and analyze within 24 hours.			
Antibody staining	1.	Collect blood into an appropriate anticoagulant (EDTA or heparin).			
without wash lysis	2.	Pipette 100 µL of thoroughly mixed blood into a conical tube.			
procedure	3.	Add antibody as indicated in the manufacturer's package insert to appropriately labeled tubes from step 2. Mix gently.			
	4.	Incubate all tubes for 15 minutes at room temperature ($22 \pm 3^{\circ}$ C) in the dark.			
	5.	Add 100 μL of Cal-Lyse [™] Lysing Solution to all tubes.			
	6.	Incubate all tubes for 10 minutes at room temperature ($22 \pm 3^{\circ}$ C) in the dark.			
	7.	Add 1.0 mL of deionized water to all tubes. Immediately vortex tubes.			
	8.	Incubate all tubes for 10 minutes at room temperature.			
	9.	Analyze on a flow cytometer according immediately, or store samples at 2°C–8°C in the dark and analyze within 24 hours.			

Appendix A

Interpretation of Results

Flow cytometry

Analyze antibody-stained cells on an appropriate flow cytometer according to the manufacturer's instructions. The right angle light scatter, or side scatter (SSC), versus forward angle light scatter (FSC) is collected to reveal the lymphocyte cell cluster. A gate is drawn for the lymphocyte cluster (lymphocyte bitmap). The fluorescence attributable to the fluorochromeconjugated monoclonal antibody is collected, and the percentage of antibodystained cells is determined. A fluorochrome-conjugated isotypic control may be used to estimate and correct for nonspecific binding to lymphocytes. The isotypic control must be of the same heavy chain immunoglobulin class as the specific antibody. The isotypic control should be used at approximately the same antibody concentration as the specific antibody. An analysis region is set to exclude background fluorescence and to include positively stained cells.

The following histograms are representative of cell staining from a normal donor, gated on the lymphocyte region and stained with a representative T cell-specific monoclonal antibody (CD3 FITC), and a B cell-specific monoclonal antibody (CD19 R-PE). Cells were processed with the "Antibody staining and wash lysis procedure". Red blood cells were lysed with Cal-Lyse[™] Lysing Solution.



Limitations of the Procedure

- 1. The values obtained from normal individuals may vary from laboratory to laboratory; therefore, it is recommended that each laboratory establish its own normal range.
- 2. When using the whole blood method, red blood cells found in some abnormal donors, as well as nucleated red cells found in normal and abnormal donors, may be resistant to lysis. Longer red cell lysis periods may be needed to avoid the inclusion of unlysed red cells.
- 3. Blood samples should not be refrigerated or retained at ambient temperature for an extensive period (longer than 24–30 hours) prior to incubation with monoclonal antibodies.

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Appendix B

Technical Support

Obtaining support	For the latest services and support information for all locations, go to www.lifetechnologies.com At the website, you can:					
	• Access worldwide telephone and fax numbers to contact Technical Support and Sales facilities					
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Explanation of Symbols

Symbol	Description	Symbol	Description	Symbol	Description
	Manufacturer	REF	Catalog number	LOT	Batch code
IVD	<i>In vitro</i> diagnostic medical device	\square	Use by	X	Temperature limitation
EC REP	European Community authorized representative	i	Consult instructions for use	Â	Caution, consult accompanying documents

Notes

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