

## Revised: 03-November-2004

# EnzChek<sup>®</sup> Protease Assay Kits

E6638 EnzChek<sup>®</sup> Protease Assay Kit \*green fluorescence\*

E6639 EnzChek<sup>®</sup> Protease Assay Kit \*red fluorescence\*

# Quick Facts

### Storage upon receipt:

- -20°C
- Desiccate
- Protect from light

## **Ex/Em of Digestion Products:**

- 505/513 nm for kit E6638
- 589/617 nm for kit E6639

### Note:

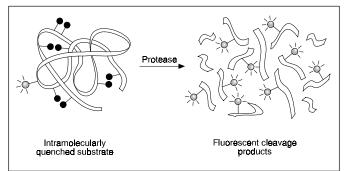
- Ensure that the substrate is fully reconstituted before use
- Avoid freeze-thaw cycles after reconstituting

# Introduction

Molecular Probes' EnzChek<sup>®</sup> Protease Assay Kits (E6638, E6639) are fast, simple and direct fluorescence-based assays for detecting metallo-, serine, acid and sulfhydryl proteases. Detecting low levels of protease activity is important in quality-control testing, high-throughput screening and basic research. However, current methods for detecting protease activity, such as the fluorescein thiocarbamoyl (FTC)–casein protease assay, require extensive manipulation and are therefore prone to error. In the FTC-casein assay, unhydrolyzed protein must be precipitated with trichloroacetic acid, separated by centrifugation, transferred for measurement and then pH-adjusted for fluorescein signal enhancement.<sup>1</sup>

Our two EnzChek Protease Assay Kits contain casein derivatives that are heavily labeled with the pH-insensitive greenfluorescent BODIPY<sup>®</sup> FL (E6638) or red-fluorescent BODIPY<sup>®</sup> TR-X (E6639) dyes, resulting in almost total quenching of the conjugate's fluorescence. Protease-catalyzed hydrolysis releases highly fluorescent BODIPY FL or BODIPY TR-X dye–labeled peptides (Figure 1). The accompanying increase in fluorescence, which can be measured with a spectrofluorometer, minifluorometer or microplate reader, is proportional to protease activity.

In contrast to the FTC-casein assay, these EnzChek assays do not involve any separation steps and can be used to continuously measure the kinetics of a variety of exo- and endopeptidases over



*Figure 1.* Principle of protease detection used in our EnzChek Protease Assay Kits (E6638, E6639).

a wide pH range. They can also be used to measure the total substrate turnover at a fixed time following addition of the enzyme. Furthermore, we have found that our protease assays are up to 100 times more sensitive and much easier to perform than the FTC-casein assay.<sup>2</sup>

In addition to their utility for detecting protease contamination of culture media and other experimental samples, BODIPY FL casein and BODIPY TR-X casein appear to have significant potential as general nontoxic, pH-insensitive markers for phagocytic cells in culture. Moreover, preliminary reports indicate that BODIPY FL casein is useful for monitoring proteolytic activity in frozen tissue sections with a fluorescence microplate reader.<sup>3</sup>

The BODIPY FL and BODIPY TR-X casein substrates can be used interchangeably, depending on whether green or red fluorescence is desired. The peptide hydrolysis products of BODIPY FL casein exhibit green fluorescence that is optimally excited by the argon-ion laser, permitting flow sorting of cells that have phagocytosed this reagent. The red-fluorescent BODIPY TR-X dye–labeled peptides, with excitation and emission spectra similar to those of the Texas Red<sup>®</sup> fluorophore, should be useful for multilabeling experiments or measurements in the presence of green autofluorescence.

# Materials

## Kit Contents

- **BODIPY FL casein or BODIPY TR-X casein** (Component A), five vials that each contain 200 µg substrate lyophilized from phosphate-buffered saline (PBS)
- **20X Digestion buffer** (Component B), 13 mL of 200 mM Tris-HCl, pH 7.8, containing 2 mM sodium azide

Each kit provides sufficient reagents for approximately 100 assays when using a standard fluorometer or 1000 assays when using a fluorescence microplate reader.

#### Storage and Handling

Upon receipt, each kit should be stored frozen at  $-20^{\circ}$ C. Allow reagents to warm to room temperature before opening vials. When stored properly, these reagents are stable for six months to one year.

Reconstituted BODIPY casein substrates may be stored at 4°C for 2–4 weeks. We recommend the addition of sodium azide at a final concentration of 2 mM to act as a preservative. If longer storage is required, freeze at -20°C. PROTECT FROM LIGHT. AVOID REPEATED FREEZING AND THAWING.

### Materials Required but Not Provided

- Deionized water (dH<sub>2</sub>O)
- Phosphate-buffered saline (PBS) (E6638 only)
- 0.1 M sodium bicarbonate, pH 8.3 (E6639 only)
- Specific buffers for detection of enzymes requiring activation compounds or a unique pH environment, if applicable (see note A)
- An appropriate enzyme standard of known specific activity, if applicable (see step 2.1)

## **Experimental Protocol**

#### **Reagent Preparation**

The solution volumes recommended in this section provide sufficient reagents for 20 assays using a fluorometer and standard 2.0 mL cuvettes, or 200 assays using a fluorescence microplate reader and 200  $\mu$ L per microplate well.

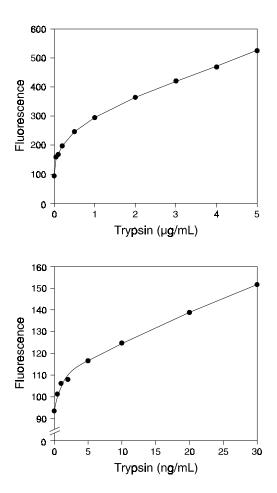
**1.1** If using the EnzChek Protease Assay Kit for green fluorescence (E6638), prepare a 1.0 mg/mL stock solution of the BODIPY FL casein by adding 0.2 mL of PBS directly to one of the vials containing the lyophilized substrate. If using the EnzChek Protease Assay Kit for red fluorescence (E6639), prepare a 1.0 mg/mL stock solution of the BODIPY TR-X casein by adding 0.2 mL of 0.1 M sodium bicarbonate, pH 8.3, to one of the vials containing the lyophilized substrate. In either case, mix well and allow sufficient time at room temperature for the substrate to dissolve fully.

**1.2** Prepare 1X digestion buffer. Dilute 2.5 mL of the 20X digestion buffer with dH<sub>2</sub>O to a final volume of 50 mL (note **A**).

**1.3** Prepare a 10  $\mu$ g/mL working solution of the BODIPY casein. Add 0.2 mL of the stock solution prepared in step 1.1 to 19.8 mL of the 1X digestion buffer prepared in step 1.2.

### Protease Activity Standard Curve

**2.1** When quantitating the activity of purified protease preparations, generate a protease activity standard curve. If possible, use an appropriate enzyme standard of known specific activity that closely matches the protease activity being determined. A standard curve may not be relevant for samples containing one or more unknown proteases. In this case, protease activity may be expressed as fluorescence change per unit sample. Also, for



**Figure 2.** Sample standard curve obtained with the EnzChek Protease Assay Kit (E-6638). The top panel shows fluorescence versus trypsin concentration ( $\mu$ g/mL) measured with a filter fluorometer (excitation 485 ± 10 nm, emission 530 ± 12.5 nm). The bottom panel displays a standard curve at lower trypsin concentrations (ng/mL), obtained using the same fluorometer but with a higher gain setting.

simple detection of protease activity or contamination, a standard curve may not be necessary; proceed to step 3.3.

**2.2** Determine the range of enzyme response. Titrate at least four concentrations of enzyme and one buffer-only control in 1.0 mL (or 100  $\mu$ L for microplate assays) of the 1X digestion buffer prepared in step 1.2. Add 1.0 mL (or 100  $\mu$ L for microplate assays) of the BODIPY casein working solution prepared in step 1.3.

**2.3** Incubate the samples for one hour, protected from light (notes **B**, **C**).

**2.4** Read the fluorescence in a fluorometer or fluorescence microplate reader. BODIPY FL and BODIPY TR-X dye–labeled peptides have excitation/emission maxima of approximately 505/513 nm and 589/617 nm, respectively. We have found that standard fluorescein filters (e.g., excitation =  $485 \pm 12.5$  nm, emission =  $530 \pm 15$  nm) can be used to detect BODIPY FL dye–labeled peptides, whereas longer wavelength filters (e.g., excitation =  $590 \pm 10$  nm, emission =  $645 \pm 20$  nm ) can be used to detect BODIPY TR-X dye–labeled peptides.

**2.5** Plot the data to show fluorescence versus protease concentration (Figure 2) (note **D**).

#### Sample Analysis

**3.1** To detect enzyme activity in a sample, dilute the sample to 1.0 mL (or 100  $\mu$ L for microplate assays) in 1X digestion buffer prepared in 1.2. Add 1.0 mL (or 100  $\mu$ L for microplate assays) of the BODIPY casein working solution prepared in step 1.3.

**3.2** Incubate sample for one hour, protected from light (notes **B**, **C**).

**3.3** Read the fluorescence in a fluorometer or fluorescence microplate reader. BODIPY FL and BODIPY TR-X dye–labeled peptides have excitation/emission maxima of approximately 505/513 nm and 589/617 nm, respectively. We have found that standard fluorescein filters (e.g., excitation =  $485 \pm 12.5$  nm, emission =  $530 \pm 15$  nm) can be used to detect BODIPY FL dye–labeled peptides, whereas longer wavelength filters (e.g., excitation =  $590 \pm 10$  nm, emission =  $645 \pm 20$  nm) can be used to detect BODIPY TR-X dye–labeled peptides.

**3.4** If the protease sample has a high fluorescence background, prepare an additional control without the BODIPY casein prepared in step 1.3. Then, subtract the fluorescence background of the substrate-free control from the sample containing the substrate to determine the true fluorescence increase due to protease activity.

# **Protease Detection Limits**

Using the EnzChek protease assay with a fluorescence microplate reader, we have determined the protease detection limits for a number of proteases. In these assays, 200  $\mu$ L reaction mixtures were incubated in a 96-well microplate for one hour at room temperature, protected from light. The fluorescence was then measured in a fluorescence microplate reader, using excitation and emission filters of 485 ± 12.5 nm and 530 ± 15 nm, respectively, for detection of BODIPY FL dye–labeled peptides and 590 ± 10 nm and 645 ± 20 nm for detection of BODIPY TR-X dye–labeled peptides. Table 1 shows the approximate detection limits for a variety of enzymes when assayed at 22°C (note **E**).

## Notes

**[A]** The digestion buffer provided (pH 7.8) is recommended for detecting the protease activity of most proteolytic enzymes with activity optima from pH 7.4 to 8.0. However, if you are working with an enzyme that requires activation compounds or a unique pH environment, then prepare the specific buffer required in place of the digestion buffer.

Table 1. Detection	limits of the	EnzChek protease	assav.
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Enzyme	Class	Detection	Buffer
(Source)		Limit (Units)	Conditions
Elastase, Type IV	Serine	$2.2 \times 10^{-3}$	10 mM Tris-HCl,
(porcine pancreas)	Protease		pH 8.8
Chymotrypsin, Type II (bovine pancreas)	Serine Protease	$5.0  imes 10^{-5}$	10 mM Tris-HCl, pH 7.8
Thermolysin (B. proteolyticus rokko)	Acid Protease	$4.4 \times 10^{-5}$	10 mM Tris-HCl, pH 7.8
Trypsin, Type IX	Serine	$1.3 \times 10^{-2}$	10 mM Tris-HCl,
(porcine pancreas)	Protease		pH 7.8
Papain	Sulfhydryl	$2.1  imes 10^{-4}$	10 mM MES,
(papaya latex)	Protease		pH 6.2
Pepsin (porcine stomach mucosa)	Acid Protease	$2.1 \times 10^{-3}$	10 mM HCl, pH 2.0
Elastase (Pseudomonas aeruginosa)	Metallo- protease	$1.0 \times 10^{-3}$	20 mM sodium phosphate, pH 8.0
Cathepsin D	Acid Protease	$2.0  imes 10^{-4}$	20 mM sodium citrate, pH 5.0
Elastase	Serine	$1.0  imes 10^{-3}$	10 mM Tris, HCl
(human leukocyte)	Protease		pH 7.5

The detection limit is defined as the amount of enzyme required to cause a 10–20% change in fluorescence compared to the control sample at 22°C. Enzyme unit definitions are standard definitions for each individual enzyme. Detection limits were determined with BODIPY FL casein and with BODIPY TR-X casein; both substrates yielded similar results. Detection limits may vary with instrumentation.

**[B]** Sensitivity may be increased by incubating for up to 24 hours.

**[C]** The exact time interval is not critical. However, it is important that all reactions, experimental samples and controls be incubated for approximately the same time. For consistent incubation periods, it may be desirable to initiate the reactions with offset starting times to allow sufficient time for reading the fluorescence of each at the end of the reaction.

[D] Standard curves will vary with enzyme type.

**[E]** Enzyme activity may vary depending on incubation buffers and temperature, as well as the storage conditions and number of freeze-thaw cycles to which the enzyme preparation has been subjected.

## References

1. Anal Biochem 143, 30 (1984); 2. Anal Biochem 251, 144 (1997); 3. Derek Winslow, "Measurement of Proteolytic Activity in Whole Tissue Sections using Quenched Fluorescent Substrates: An Analogue of *In-Situ* Zymography?" Focus on Fluorescence: Industrial Applications Symposium, Leiden, The Netherlands, November 17, 1997.

<b>Product List</b> Current prices may be obtained from our Web site or from our Customer Service Department.					
Cat #	ProductName	Unit Size			
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