

CD300e (IREM-2) antibodies human

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Warnings

Reagents contain sodium azide. Under acidic conditions sodium azide yields hydrazoic acid, which is extremely toxic. Azide compounds should be diluted with running water before discarding. These precautions are recommended to avoid deposits in plumbing where explosive conditions may develop.

1. Description

This product is for research use only.

Components

Monoclonal CD300e (IREM-2) antibodies, human conjugated to:

Conjugate	Order no. 1 mL (100 tests)	Order no. 300 μL (30 tests)
FITC	130-101-776	130-101-790
PE	130-101-777	130-101-791
APC	130-101-773	130-101-788
VioBlue®	130-101-779	130-101-796
PE-Vio770™	130-101-778	130-101-793
APC-Vio770™	130-101-774	130-101-789
PerCP-Vio700	130-101-775	130-101-792
Biotin	130-101-813	130-101-836

Clone UP-H2 (isotype: mouse IgG1κ).

Capacity 1 mL: 100 tests or up to 109 total cells

 $300~\mu\text{L}{:}~30~tests$ or up to $3{\times}10^{8}$ total cells.

Product format Antibodies are supplied in buffer containing

stabilizer and 0.05% sodium azide.

Storage Store protected from light at 2–8 °C. Do not

freeze. The expiration date is indicated on the

vial label.

1.1 Background information

- Antigen: CD300e (IREM-2)
- Synonym: IREM-2; LMIR6; CMRF35-A5
- Expression patterns: The monoclonal antibody UP-H2 reacts with human CD300e, also known as IREM-2, LMIR6, or CMRF35-A5. This 34 kDa activating receptor of the immunglobulin superfamily is expressed on mature myeloid dendritic cells and monocytes and is down-regulated on immature dendritic cells. CD300e interacts with DAP-12, which mediates acivating signals. It is reported that the alloreactive response of naive T cells is enhanced by IREM-2 activation of mDCs.

1.2 Applications

 Identification and enumeration of CD300e (IREM-2)* cells by flow cytometry.

1.3 Recommended antibody dilution

The recommended antibody dilution for all CD300e (IREM-2) conjugates is 1:11 for up to 10^7 cells/100 μL of buffer for labeling of cells and subsequent analysis by flow cytometry.

Cells should be stained prior to fixation, if formaldehyde is used as a fixative.

1.4 Reagent requirements

- Buffer: Prepare a solution containing phosphate-buffered saline (PBS), pH 7.2, 0.5% bovine serum albumin (BSA), and 2 mM EDTA by diluting MACS* BSA Stock Solution (#130-091-376) 1:20 with autoMACS* Rinsing Solution (#130-091-222). Keep buffer cold (2-8 °C).
 - ▲ Note: EDTA can be replaced by other supplements such as anticoagulant citrate dextrose formula-A (ACD-A) or citrate phosphate dextrose (CPD). Buffers or media containing Ca²⁺ or Mg²⁺ are not recommended for use.
- (Optional) FcR Blocking Reagent, human (#130-059-901) to avoid Fc receptor-mediated antibody labeling.
- (Optional) Tandem Signal Enhancer, human (# 130-099-888) to reduce non-specific binding of tandem dye-conjugated antibodies to human cells, especially to monocytes.
- (Optional) Conjugated anti-biotin antibodies, e.g., Anti-Biotin-PE (# 130-090-756) as secondary antibody reagent in combination with CD300e (IREM-2)-Biotin.
- (Optional) For antibodies for additional staining or for isotype control, refer to www.miltenyibiotec.com/antibodies.
- (Optional) Propidium Iodide Solution (#130-093-233) for flow cytometric exclusion of dead cells without fixation.
- (Optional) Fixation and Dead Cell Discrimination Kit (#130-091-163) for cell fixation and flow cytometric exclusion of dead cells.

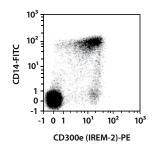
2. General protocol for immunofluorescent staining

Volumes given below are for **up to 10^7** nucleated cells. When working with fewer than 10^7 cells, use the same volumes as indicated. When working with higher cell numbers, scale up all reagent volumes and total volumes accordingly (e.g. for 2×10^7 nucleated cells, use twice the volume of all indicated reagent volumes and total volumes).

- 1. Determine cell number.
- 2. Centrifuge cell suspension at 300×g for 10 minutes. Aspirate supernatant completely.
- 3. Resuspend up to 10⁷ nucleated cells per 100 μL of buffer.
- 4. Add $10 \mu L$ of the CD300e (IREM-2) antibody.
- 5. Mix well and incubate for 10 minutes in the dark in the refrigerator (2–8 °C).
 - ▲ Note: Higher temperatures and/or longer incubation times may lead to nonspecific cell labeling. Working on ice requires increased incubation times.
- 6. Wash cells by adding 1–2 mL of buffer and centrifuge at 300×g for 10 minutes. Aspirate supernatant completely.
- 7. (Optional) If CD300e (IREM-2)-Biotin was used, resuspend the cell pellet in 100 μ L of buffer, add 10 μ L of anti-biotin antibody, and continue as described in steps 5 and 6.
- Resuspend cell pellet in a suitable amount of buffer for analysis by flow cytometry or fluorescence microscopy.

3. Example of immunofluorescent staining with CD300e (IREM-2) antibodies

Human peripheral blood mononuclear cells (PBMCs) were stained with CD300e (IREM-2) antibodies conjugated to PE as well as with CD14-FITC (# 130-080-701) and analyzed by flow cytometry using the MACSQuant* Analyzer. Cell debris and dead cells were excluded from the analysis based on scatter signals and propidium jodide fluorescence.



For more examples please refer to the respective product page at www.miltenyibiotec.com/antibodies.

4. References

- Aguilar, H. et al. (2004) Molecular characterization of a novel immune receptor restricted to the monocytic lineage. J. Immunol. 173: 6703–6711.
- Brckalo, T. et al. (2010) Functional analysis of the CD300e receptor in human monocytes and myeloid dendritic cells. Eur. J. Immunol. 40 (3): 722–732.

Refer to www.miltenyibiotec.com for all data sheets and protocols.

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