

Points to consider

when culturing human Embryonic Stem Cells with STEMPRO® hESC SFM

- Starter culture
 - This must be a high quality culture. There must be a high density of cells, and they must be primarily undifferentiated. The starter culture should be cells maintained on Geltrex™ in MEF-CM, i.e., not hESCs on MEFs.
- Passaging
 - Passaging the cells is the most likely point of difficulty. It is absolutely critical to achieve high plating/survival of colony pieces. The colony pieces must be a bit smaller than typical collagenase passaging of cells on Geltrex™/MEF-CM .
 - Some cell death at passaging is normal, but wide-scale cell death is not (ie <20% survival), and typically indicates a poor split.
- Timing of passaging
 - This is critical. Do not passage the cells too early, they will plate poorly and differentiate. The cultures need to grow to near-confluence, i.e., a day or two longer than when the colonies are just touching. We routinely harvest 5-8 million cells per 60 mm dish.
- hESCs in STEMPRO® hESC SFM are very sensitive to over exposure to collagenase. This will cause poor plating and induce differentiation. Do not expose longer than 3 minutes. Do not use lower concentrations of collagenase and treat for longer periods.
- Density
 - The cultures must be maintained at a high density. This means 200+ colonies in a 60 mm dish. If the density of colonies drops, the culture will tend to deteriorate: cells will differentiate, and the culture will take longer between splits. If this happens, leave the culture longer to proliferate to near-confluence before splitting.
- The “knife edge”
 - Just as with hESCs grown in other conditions, cells in STEMPRO® hESC SFM sit on a knife edge of proliferation vs. differentiation. The cultures should be fed every day. Do not exhaust the medium by not feeding. Badly differentiated areas should be scraped out with a pipette tip. Mistreated cultures will differentiate.
 - Passage the cells 2-3 times in the previous ratio of MEF conditioned media to defined media.
- We strongly recommend that you always take these precautions
 - Make a frozen stock of the cells in MEF conditioned media prior to adaptation
 - Keep a culture going of the cells in each prior condition when starting the next level of adaptation as a fall-back if the cells do not survive in the next passage.
- Technical support
 - Please contact Tech Services at Invitrogen
United States TECH-LINESM: 1 800 955 6288
Canada TECH-LINESM: 1 800 757 8257

Outside the U.S. and Canada, refer to the GIBCO products catalogue for the TECH-LINE in your region.
- You may also contact your Invitrogen Sales Representative or our World Wide Web site at www.invitrogen.com.

References

In this section we list references to articles that may further help you culture and understand human Embryonic Stem Cells.

Amit, M, Shariki, C, Margulets, V, et al. (2004) Feeder layer- and serum-free culture of human embryonic stem cells. *Biol Reprod*: 70: 837-45.

Babaie, Y, Herwig, R, Greber, B, et al. (2006) Analysis of OCT4 dependent transcriptional networks regulating self renewal and pluripotency in human embryonic stem cells. *Stem Cells*.

Boyer, L A, Lee, T I, Cole, M F, et al. (2005) Core transcriptional regulatory circuitry in human embryonic stem cells. *Cell*: 122: 947-56.

Ding, V, Choo, a B, and Oh, S K. (2006) Deciphering the importance of three key media components in human embryonic stem cell cultures. *Biotechnol Lett*: 28: 491-5.

Hovatta, O. (2006) Derivation of human embryonic stem cell lines, towards clinical quality. *Reprod Fertil Dev*: 18: 823-8.

James, D, Levine, a J, Besser, D, et al. (2005) TGFbeta/activin/nodal signaling is necessary for the maintenance of pluripotency in human embryonic stem cells. *Development*: 132: 1273-82.

Li, Y, Powell, S, Brunette, E, et al. (2005) Expansion of human embryonic stem cells in defined serum-free medium devoid of animal-derived products. *Biotechnol Bioeng*: 91: 688-98.

Mallon, B S, Park, K Y, Chen, K G, et al. (2006) Toward xeno-free culture of human embryonic stem cells. *Int J Biochem Cell Biol*: 38: 1063-75.

Richards, M, and Bongso, A. (2006) Propagation of human embryonic stem cells on human feeder cells. *Methods Mol Biol*: 331: 23-41.

Rosler, E S, Fisk, G J, Ares, X, et al. (2004) Long-term culture of human embryonic stem cells in feeder-free conditions. *Dev Dyn*: 229: 259-74.

Thomson, J A, Itskovitz-Eldor, J, Shapiro, S S, et al. (1998) Embryonic stem cell lines derived from human blastocysts. *Science*: 282: 1145-7.

Wang, G, Zhang, H, Zhao, Y, et al. (2005) Noggin and bFGF cooperate to maintain the pluripotency of human embryonic stem cells in the absence of feeder layers. *Biochem Biophys Res Commun*: 330: 934-42.

Yao, S, Chen, S, Clark, J, et al. (2006) Long-term self-renewal and directed differentiation of human embryonic stem cells in chemically defined conditions. *Proc Natl Acad Sci U S A*: 103: 6907-12.

Zeng, X, Miura, T, Luo, Y, et al. (2004) Properties of pluripotent human embryonic stem cells BG01 and BG02. *Stem Cells*: 22: 292-312.

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