BD Transduction Laboratories[™] Bioimaging Certified Reagent

Technical Data Sheet

Purified Mouse Anti-ERK1/2 (pT202/pY204)

Product Information

Material Number: Alternate Name: Size: Concentration: Clone: Immunogen: Isotype: Reactivity:

Target MW: Storage Buffer: 612359 p44/42 MAPK; Extracellular signal-Regulated Kinase 1/2 (pT202/Y204) 150 μg 250 μg/ml 20A Phosphorylated Rat ERK1 (T202/Y204) Peptide Mouse IgG1 QC Testing: Human Tested in Development: Mouse, Rat 44/42 kDa Aqueous buffered solution containing BSA, glycerol, and ≤0.09% sodium azide.

Description

The members of the Mitogen-Activated Protein Kinase (MAPK) family are components of a key signal transduction cascade that links events at the cell surface to responses in the nucleus. The signaling cascade is found in species as varied as yeast and humans, with many of the proteins being well conserved. In mammals the most widely studied members of the cascade are the Extracellular signal-Regulated Kinases, ERK1 (p44 MAPK) and ERK2 (p42 MAPK). ERK1 and ERK2 share 85% homology and are activated by extracellular signals such as growth factors, hormones, and phorbol esters. Activation occurs through a series of phosphorylations by kinases activating other kinases and eventually leading to phosphorylation of the ERKs. Growth factor stimulation leads to activation of Ras and Raf, leading to phosphorylation of MEK1 (MAPK/ERK kinase) which, in turn, activates the ERKs via dual phosphorylation. Once activated, the ERKs phosphorylate other cytoplasmic signalling molecules, cell-surface receptors, microtubule-associated proteins, and transcription factors in the nucleus. Thus, the active ERK has myriad downstream effectors that implicate it in the control of cell proliferation and differentiation, as well as regulation of the cytoskeleton. Furthermore, studies have shown that elevated ERK activity is associated with some cancers.

The 20A monoclonal antibody recognizes the phosphorylated threonine 202 and tyrosine 204 (pT202/pY204) of human ERK1 and pT184/pY186 of human ERK2. The orthologous phosphorylation sites in murine ERK1 and ERK2 are T203/Y205 and T183/Y185.





Left panel (western blot): A431 cells (ATCC CRL-1555) were either left untreated (lane 1) or treated (lane 2) with 100 ng/mL EGF for 5 minutes at 37°C. The top panel was probed with Purified Mouse Anti-ERK1 (Cat. No. 610030) showing ERK1 at 44 kDa, which may also be crossreactive to ERK2 at 42 kDa. The bottom panel was probed with ERK1/2 (pT202/pY204) (Cat. No. 612358/612359). Western blotting with Purified Mouse Anti-ERK1/2 (pT202/pY204) displays bands for human ERK1 (pT202/pY204) at 44 kDa and ERK2 (pT184/pY186) at 42 kDa. In some instances, investigators may also observe an unidentified 50 kDa band.

Right panel (Immunofluorescence staining): A549 (ATCC CCL-185) cells were seeded in a 96 well imaging plate (Cat. No. 353219) at ~10,000 cells per well. After overnight incubation, cells were either mock treated (media alone, left) or exposed to PMA (right) for 30 minutes. After treatment cells were stained using the alcohol perm protocol and the Purified Mouse Anti-ERK1/2 (pT202/pY204) antibody. The second step reagent was Alexa Fluor® 488 Anti-Mouse IgG. The image was taken on a BD Pathway™ 855 Bioimager with a 20x objective. This antibody also stains HeLa (ATCC CCL-2) and U-2 OS (ATCC HTB-96) cells and and worked with both the Triton™ X-100 and alcohol perm protocols (see Recommended Assay Procedure).

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Preparation and Storage

Store undiluted at -20°C.

The monoclonal antibody was purified from tissue culture supernatant or ascites by affinity chromatography.

Application Notes

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Western blot	Routinely Tested
Bioimaging	Tested During Development
Intracellular staining (flow cytometry)	Tested During Development

Recommended Assay Procedure:

Bioimaging

- 1. Seed the cells in appropriate culture medium at ~10,000 cells per well in a Falcon® 96-well Imaging Plate (Cat. No. 353219) and culture overnight.
- 2. Remove the culture medium from the wells, and fix the cells by adding 100 µl of BD Cytofix[™] Fixation Buffer (Cat. No. 554655) to each well. Incubate for 10 minutes at room temperature (RT).
- 3. Remove the fixative from the wells, and permeabilize the cells using either BD Perm Buffer III, 90% methanol, or Triton™ X-100:
 - a. Add 100 µl of -20°C 90% methanol or Perm Buffer III (Cat. No. 558050) to each well and incubate for 5 minutes at RT.

OR

- b. Add 100 µl of 0.1% Triton[™] X-100 to each well and incubate for 5 minutes at RT.
- 4. Remove the permeabilization buffer, and wash the wells twice with 100 μ l of 1× PBS.
- 5. Remove the PBS, and block the cells by adding 100 µl of BD Pharmingen[™] Stain Buffer (FBS) (Cat. No. 554656) to each well. Incubate for 30 minutes at RT.
- 6. Remove the blocking buffer and add 50 µl of the optimally titrated primary antibody (diluted in Stain Buffer) to each well, and incubate for 1 hour at RT.
- 7. Remove the primary antibody, and wash the wells three times with 100 μ l of 1× PBS.
- 8. Remove the PBS, and add the second step reagent at its optimally titrated concentration in 50 µl to each well, and incubate in the dark for 1 hour at RT.
- 9. Remove the second step reagent, and wash the wells three times with 100 μ l of 1× PBS.
- 10. Remove the PBS, and counter-stain the nuclei by adding 200 µl per well of 2 µg/ml Hoechst 33342 (e.g., Sigma-Aldrich Cat. No. B2261) in 1× PBS to each well at least 15 minutes before imaging.
- 11. View and analyze the cells on an appropriate imaging instrument.

Suggested Companion Products

Catalog Number	Name	Size	Clone	
611447	A431 Cell Lysate	500 μg	(none)	
611448	A431 + EGF Cell Lysate	500 µg	(none)	
610030	Purified Mouse Anti-ERK1	50 µg	MK12	
353219	BD Falcon [™] 96-well Imaging Plate		(none)	
554655	Fixation Buffer	100 mL	(none)	
558050	Perm Buffer III	125 mL	(none)	
554656	Stain Buffer (FBS)	500 mL	(none)	

Product Notices

- 1. Since applications vary, each investigator should titrate the reagent to obtain optimal results.
- This antibody has been developed and certified for the bioimaging application. However, a routine bioimaging test is not performed on every 2. lot. Researchers are encouraged to titrate the reagent for optimal performance.
- 3. Triton is a trademark of the Dow Chemical Company.

Europe

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- 4. Source of all serum proteins is from USDA inspected abattoirs located in the United States.
- 5. Caution: Sodium azide vields highly toxic hydrazoic acid under acidic conditions. Dilute azide compounds in running water before discarding to avoid accumulation of potentially explosive deposits in plumbing.
- Please refer to www.bdbiosciences.com/pharmingen/protocols for technical protocols. 6.

References

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