

## Technical Data Sheet

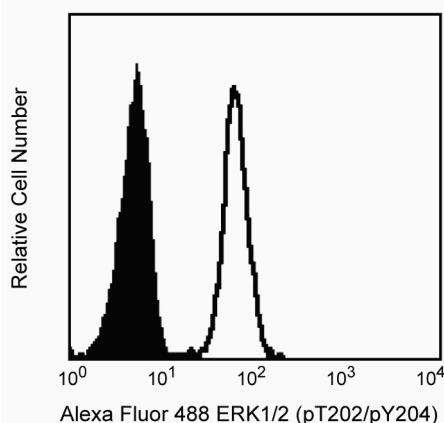
## Lyse/Fix Buffer 5X

## Product Information

Material Number:	558049
Size:	250 mL
Storage Buffer:	Aqueous buffered solution containing formaldehyde and proprietary ingredients.

## Description

BD Phosflow™ Lyse/Fix Buffer is intended to be used for the lysing and fixing of whole blood for use with intracellular flow cytometry. For example, after cell stimulation with kinase activators, such as phorbol esters, the BD Phosflow™ Lyse/Fix buffer can be used for lysing red blood cells and for fixing leukocytes in one step. This allows investigators to rapidly inactivate kinases, phosphatases and/or proteases, so that the in vivo phosphorylation state of the cell can be examined. The BD Phosflow™ Lyse/Fix Buffer has been reported to preserve the light scattering properties of the cell and may be used on human and mouse whole blood. Whole blood lysis has been shown to be as effective as density gradient centrifugation in the preparation of PBMCs for lymphocyte subset analysis. In many cases, phospho-specific antibodies may be used simultaneously with the staining of cell surface markers (e.g CD markers). Not all cell surface markers, however, are compatible with the BD Phosflow™ Lyse/Fix Buffer. For a current listing of compatible cell surface markers for use with BD Phosflow™ Lyse/Fix Buffer, investigators are encouraged to visit the following website: [http://www.bdbiosciences.com/docs/Validated\\_Cell\\_Surface\\_Markers.xls](http://www.bdbiosciences.com/docs/Validated_Cell_Surface_Markers.xls).



**Flow cytometric analysis of ERK1/2 (pT202/pY204) on human whole blood treated with BD Phosflow™ Lyse/Fix Buffer.** Freshly isolated human whole blood was left either untreated (shaded histogram) or treated with 40 nM PMA at 37°C for 10 minutes (open histogram). Leukocytes were spun down and washed once with Hank's solution, followed by permeabilization with BD Phosflow™ Perm Buffer II (Cat. No. 558052) for 30 minutes. After washing the cells twice with BD Pharmingen™ Stain Buffer (Cat. No. 554656), the cells were stained with Alexa Fluor® 488 mouse anti-ERK1/2 (pT202/pY204) (Cat. No. 612592). Represented is the data on the lymphocyte gated population.

## Preparation and Storage

Store undiluted at room temperature.

## Application Notes

## Application

Intracellular staining (flow cytometry)	Routinely Tested
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## Recommended Assay Procedure:

**Flow cytometry:**

1. Collect normal human blood in the presence of EDTA or heparin and treat with appropriate activators where applicable.
2. Dilute the required amount of BD Phosflow™ Lyse/Fix Buffer (5X concentrate) 1:5 with deionized or distilled water (at room temperature) and then pre-warm the solution to 37°C. The 1X working solution should be made fresh for each experiment and any remaining solution at the end of the experiment should be discarded.
3. Mix one volume of blood with 20 volumes of the pre-warmed BD Phosflow™ Lyse/Fix Buffer (1X). Mix well by vigorously inverting the tubes 8-10 times, and then incubate the tubes in a 37°C water bath for 10 min.
4. Spin down the cells at 500x g for 8 min in a table-top centrifuge, aspirate the supernatant, and wash the cells once with Hank's solution.
5. Vortex to loosen the cells and permeabilize the cells by adding the appropriate permeabilization reagent, such as cold BD Phosflow™ Perm Buffer II, slowly while vortexing. Incubate on ice for 30 min and then spin down the cells.
6. Wash the cells twice and resuspend with BD Pharmingen™ Staining Buffer (Cat. No. 554656).
7. Stain the cells with appropriate antibodies for 30-60 min at room temperature, wash and prepare for flow cytometric analysis.

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**Danger:** BD Phosflow™ Lyse/Fix Buffer contains 7.15% methanol (w/w), 20.35% formaldehyde (w/w) and 15.65% diethylene glycol (w/w).

**Hazard Statements:**

Combustible liquid.

Harmful if swallowed or in contact with skin.

Toxic if inhaled.

Causes skin irritation.

Causes serious eye damage.

May cause an allergic skin reaction.

Suspected of causing genetic defects.

May cause cancer. Route of exposure: Inhalative.

May cause damage to organs. May cause respiratory irritation.

May cause damage to the kidneys through prolonged or repeated exposure. Route of exposure: Oral.

**Precautionary Statements:**

Wear protective gloves / eye protection.

Wear protective clothing.

Do not breathe mist/vapours/spray.

IF ON SKIN (or hair): Remove/Take off immediately all contaminated clothing. Rinse skin with water/shower.

IF IN EYES: Rinse cautiously with water for several minutes. Remove contact lenses, if present and easy to do.

Continue rinsing.

IF INHALED: Remove victim to fresh air and keep at rest in a position comfortable for breathing.

**Suggested Companion Products**

Catalog Number	Name	Size	Clone
612592	Alexa Fluor® 488 Mouse Anti-ERK1/2 (pT202/pY204)	50 Tests	20A
554656	Stain Buffer (FBS)	500 mL	(none)
558052	Perm Buffer II	125 mL	(none)

**Product Notices**

1. The Alexa Fluor®, Pacific Blue™, and Cascade Blue® dye antibody conjugates in this product are sold under license from Molecular Probes, Inc. for research use only, excluding use in combination with microarrays, or as analyte specific reagents. The Alexa Fluor® dyes (except for Alexa Fluor® 430), Pacific Blue™ dye, and Cascade Blue® dye are covered by pending and issued patents.
2. Please refer to [www.bdbiosciences.com/pharmingen/protocols](http://www.bdbiosciences.com/pharmingen/protocols) for technical protocols.

**References**

Ashmore LM, Shopp GM, Edwards BS. Lymphocyte subset analysis by flow cytometry. Comparison of three different staining techniques and effects of blood storage. *J Immunol Methods*. 1989; 118(2):209-215. (Biology)

De Paoli P, Reitano M, Battistin S, Castiglia C, Santini G. Enumeration of human lymphocyte subsets by monoclonal antibodies and flow cytometry: a comparative study using whole blood or mononuclear cells separated by density gradient centrifugation. *J Immunol Methods*. 1984; 72(2):349-353. (Biology)

Renzi P, Ginns LC. Analysis of T cell subsets in normal adults. Comparison of whole blood lysis technique to Ficoll-Hypaque separation by flow cytometry. *J Immunol Methods*. 1987; 98(1):53-56. (Biology)

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