

Technical Data Sheet

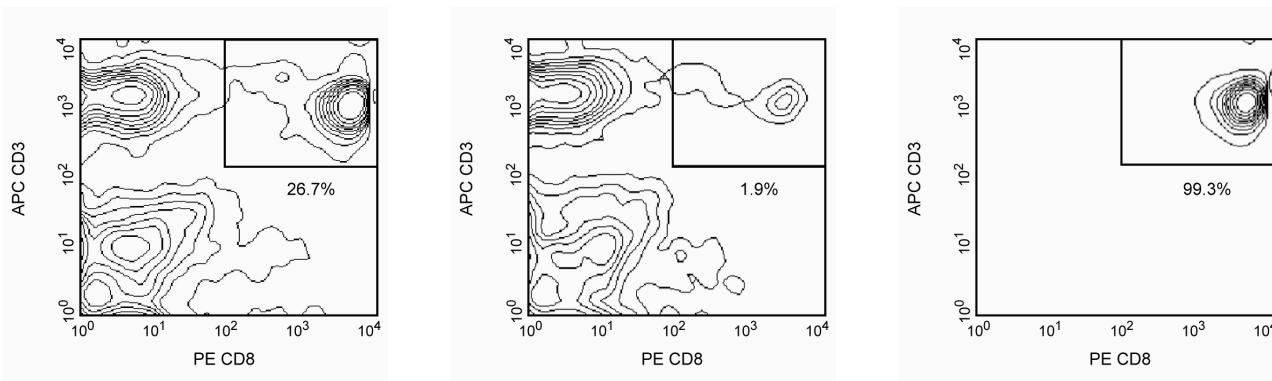
Anti-Human CD8 Magnetic Particles - DM

Product Information

Material Number:	557766
Size:	5.0 ml
Clone:	SK1
Reactivity:	QC Testing: Human Tested in Development: Rhesus Macaque
Storage Buffer:	Aqueous buffered solution containing BSA and ≤0.09% sodium azide.

Description

BD IMag™ anti-human CD8 Particles - DM are magnetic nanoparticles that have monoclonal antibody conjugated to their surfaces. These particles are optimized for the positive selection or depletion of CD8-bearing leukocytes using the BD IMagnet™. CD8 is expressed on the peripheral MHC class I-restricted suppressor/cytotoxic T-lymphocyte subset, on a subset of NK cells, and on the majority of thymocytes. The SK1 mAb has been reported to cross-react with lymphocytes of chimpanzee and cynomolgus, pig-tailed, and rhesus macaque. Peripheral Blood Mononuclear Cells (PBMC) are labeled with BD IMag™ Anti-Human CD8 Magnetic Particles - DM according to the Magnetic Labeling Protocol. This labeled cell suspension is then placed within the magnetic field of the BD IMagnet™ (Cat. No. 552311). Labeled cells migrate toward the magnet (positive fraction), leaving the unlabeled cells in suspension so they can be drawn off (negative fraction). The tube is then removed from the magnetic field for resuspension of the positive fraction. The separation is repeated twice to increase the purity of the positive fraction. The magnetic separation steps are diagrammed in the Separation Flow Chart. After the positive fraction is washed, the small size of the magnetic particles allows the positive fraction to be further evaluated in downstream applications such as flow cytometry.



Positive selection of human CD8+ T lymphocytes from PBMC. Leukocytes were labeled with BD IMag™ Anti-Human CD8 Magnetic Particles - DM as described in the protocol. After labeling, the cells were separated using the BD IMagnet™, and the negative (CD8-) and positive (CD8+) fractions were collected. Please refer to the Separation Flow Chart to identify the separated cell populations represented in this figure. For flow cytometric analysis, fresh PBMC (left panel), the negative fraction (middle panel) and the positive fraction (right panel) were stained with PE mouse anti-human CD8 (Cat. No. 555367) and APC mouse anti-human CD3 (Cat. No. 555335). The percent CD8+/CD3+ cells in each sample is given.

Preparation and Storage

Store undiluted at 4°C.

Application Notes

Application

Cell separation	Routinely Tested
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Recommended Assay Procedure:

Magnetic Labeling Protocol

1. Prepare PBMC from anti-coagulated human (or rhesus macaque) blood, preferably by density gradient centrifugation using Ficoll-Paque™.*
2. Dilute BD IMag™ Buffer (10X) (Cat. No. 552362) 1:10 with sterile distilled water or prepare 1X BD IMag™ buffer by supplementing Phosphate Buffered Saline with 0.5% BSA, 2 mM EDTA, and 0.09% sodium azide. Store at 4°C.
3. Count the cells, wash them with an excess volume of 1X BD IMag™ buffer, and carefully aspirate all the supernatant.
4. Vortex the BD IMag™ Anti-Human CD8 Magnetic Particles - DM thoroughly, and add 50 µl of particles for every 10⁷ total cells.†
5. **MIX THOROUGHLY.** Incubate at room temperature for 30 minutes.‡
6. Bring the BD IMag™-particle labeling volume up to 1-8 x 10⁷ cells/mL with 1X BD IMag™ buffer, and immediately place the tube on the BD IMagnet™. Incubate for 8-10 minutes.

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7. With the tube on the BD IMagnet™, carefully aspirate off the supernatant. This supernatant contains the negative fraction.
8. Remove the tube from the BD IMagnet™, and add 1 ml of 1X BD IMag™ buffer to the same volume as in Step 6. Gently resuspend cells by pipetting up and down, and return the tube to the BD IMagnet™ for another 2-4 minutes.
9. With the tube on the BD IMagnet™, carefully aspirate off the supernatant and discard.
10. Repeat Steps 8 and 9.
11. After the final wash step, resuspend the positive fraction in an appropriate buffer or media, and proceed with desired downstream application(s).

NOTES:

* Hints for successful cell preparation:

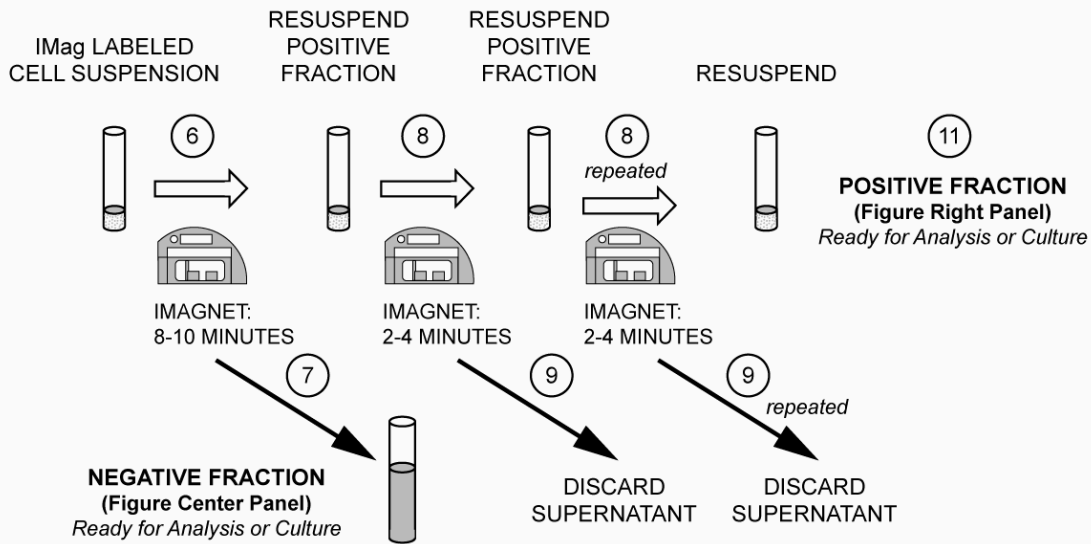
- Draw the blood into a tube containing EDTA (e.g. BD Vacutainer® EDTA tube, Cat. No. 366457 or 367661).
- Remove the platelet rich plasma by centrifuging once at 220-240 × g.
- Wash 2-3 times in PBS after the density gradient separation.
- Remove clumps of cells and/or debris by passing the suspension through a 70-µm nylon cell strainer.

† The BD IMag™ particles may need to be titrated to optimize the separation of rhesus macaque leukocytes.

‡ Avoid nonspecific labeling by working quickly and adhering to the recommended incubation times.

SEPARATION FLOW CHART

(The circled numbers correspond to the steps of the following Protocol.)



Suggested Companion Products

Catalog Number	Name	Size	Clone
552362	Buffer (10X)	100 ml	(none)
552311	Cell Separation Magnet	each	(none)
555335	APC Mouse Anti-Human CD3	100 tests	UCHT1
555367	PE Mouse Anti-Human CD8	100 tests	RPA-T8

Product Notices

1. Since applications vary, each investigator should titrate the reagent to obtain optimal results.
2. Please refer to www.bdbiosciences.com/pharming/en/protocols for technical protocols.
3. BD IMag™ particles are prepared from carboxy-functionalized magnetic particles which are manufactured by Skold Technology and are licensed under US patent number 7,169,618.
4. Ficoll-Paque is a trademark of Amersham Biosciences Limited.
5. Caution: Sodium azide yields highly toxic hydrazoic acid under acidic conditions. Dilute azide compounds in running water before discarding to avoid accumulation of potentially explosive deposits in plumbing.

6. Source of all serum proteins is from USDA inspected abattoirs located in the United States.

References

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