Technical Data Sheet

Purified Active Recombinant Human Caspase-8

Product Information

 Material Number:
 556480

 Size:
 10 μg

Component: 51-66311V

Description: Recombinant Human Active Caspase-8

 Size:
 5 μg (2 ea)

 Concentration:
 0.2 mg/ml

Storage Buffer: 50 mM Tris (pH 8.0) with 100 mM NaCl and 50 mM imidazole

Description

Caspases are cysteine proteases that play a central role in apoptosis. The caspase family was discovered by searching human cDNA libraries for sequences homologous to ced-3, a C. elegans death gene that is required for normal apoptosis during development. The first mammalian homolog of ced-3 to be identified was ICE (interleukin-1β-converting enzyme). Subsequent numerous human ced-3 homologues were rapidly identified which led to multiple names for many of the molecules. To achieve consistency, "caspase" was adopted as a root name for all family members. The name was selected based on two catalytic properties of these enzymes, the "c" reflects a cysteine protease mechanism and "aspase" refers to their unique ability to cleave after aspartic acid. There are at least 10 members, caspase-1 (ICE), caspase-2 (ICH-1), caspase-3 (CPP32, Yama, apopain), caspase-4 (TX, ICH-2, ICErel-II), caspase-5 (ICErel-III), caspase-6 (Mch2), caspase-7 (Mch3, ICE-LAP3, CMH-1), caspase-8 (MACH, FLICE, Mch5), caspase-9 (ICELAP6, Mch6), and caspase-10 (Mch4). Each caspase is synthesized as an inactive proenzyme that is processed by cleavage at asparte residues by another protease or by self-proteolysis. The processed forms consist of large (17-22 kDa) and small (10-12 kDa) subunits which associate to form an active enzyme. The activation of some of these caspases has been shown to occur during apoptosis.

Caspase-3, -6, -7, and -8 have been shown to play a role in apoptosis induced by the death receptors Fas and tumor necrosis factor receptor type 1 (TNFR1). One of their substrates is poly (ADP ribose) polymerase (PARP). PARP is an enzyme that is involved in DNA repair and genomic maintenance. Activated caspases 3, 6, 7 and 8 can all cleave PARP from its 116 kDa form to an 85 kDa residual fragment. The cleavage separates the DNA-binding domain in the N-terminus of PARP from its C-terminus catalytic domain, and results in loss of normal PARP function. The cleavage site in PARP is C-terminal to Asp-216. The upstream sequence of the PARP cleavage site, DEVD (Asp-Glu-Val-Asp), is utilized as a basis for highly specific caspase-3 substrates such as Ac(N-acetyl)-DEVD-AFC (7-amino-4-trifluoromethylcourmarin) and Ac(N-acetyl)-DEVD-AMC (7-amino-4-methylcourmarin) as well as the caspase-3 aldehyde inhibitor Ac-DEVD-CHO.

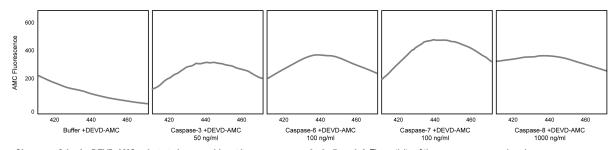
This product is supplied as 2 vials of MN 51-66311V (556481), each containing 5 μg in a volume of 25 μl .

Preparation and Storage

Store product at -80°C prior to use or for long term storage of stock solutions.

Avoid multiple freeze-thaws of product.

Once thawed, the active enzyme is stable at 4°C for at least one week.



Cleavage of the Ac-DEVD-AMC substrate by recombinant human caspases-3, -6, -7, and -8. The activity of the caspases was analyzed by spectrofluorometry using an excitation at 380 nm and an emission wavelength of 430-460 nm (peak is at 440 nm). The concentration of each caspase (50, 100, or 1000 ng/ml used in the reactions is noted in the graphs.

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Application Notes

Application

Fluorescence quantitation	Routinely Tested
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Recommended Assay Procedure:

This product (Active caspase-8) is expressed in E. coli and purified. When expressed in E. coli, caspase-8 spontaneously undergoes autoprocessing to yield the subunits characteristic of the active enzyme. The rate of caspase enzymatic hydrolysis can be measured by the release of AMC from the Ac-DEVD-AMC caspase substrate as emission at 440 nm upon excitation at 380 nm using a spectrofluorometer.

Enzyme Evaluation Protocol

This protocol is used to measure caspase enzyme activity. The synthetic fluorogenic peptide, Ac-DEVD-AMC, is used as the caspase enzyme substrate. The enzyme cleaves the substrate between D and AMC, releasing the fluorescent AMC. AMC release is measured by spectrofluorometry using UV excitation of 380 nm and emission wavelength of 440 nm.

- 1. Add 20 µM of Ac-DEVD-AMC to 1 ml of assay buffer (20 mM PIPES, 100 mM NaCl, 10mM DTT, 1mM EDTA, 0.1% (w/v) CHAPS, 10% sucrose, pH 7.2). Add the appropriate amount of the selected active caspase to the mixture (as indicated in the Enzyme Concentration table).
- 2. Incubate for 1 hr at 37°C.
- 3. Measure the AMC liberated from the Ac-DEVD-AMC using a spectrofluorometer with an excitation wavelength of 380 nm and an emission wavelength of 440 nm.

Caspase	Concentration	Cat. No. and Size
Caspase-3	50ng/ml	556472 (5 µg) / 556471 (10 µg)
Caspase-6	100ng/ml	556475 (5 μg) / 556474 (10 μg)
Caspase-7	100ng/ml	556478 (5 μg) / 556477 (10 μg)
Caspase-8	1000ng/ml	556481 (5 µg) / 556480 (10 µg)
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Suggested Companion Products

Catalog Number	Name	Size	Clone
556449	Ac-DEVD-AMC Caspase-3 Fluorogenic Substrate	1.0 mg	(none)
556552	Ac-IETD-AFC Caspase-8 Fluorogenic Substrate	1.0 mg	(none)

Product Notices

- 1. Since applications vary, each investigator should titrate the reagent to obtain optimal results.
- 2. Please refer to www.bdbiosciences.com/pharmingen/protocols for technical protocols.

References

Nicholson DW, Ali A, Thomberry NA, et al. Identification and inhibition of the ICE/CED-3 protease necessary for mammalian apoptosis. *Nature*. 1995; 376(6535):17-18.(Biology)

Patel T, Gores GJ, Kaufmann SH. The role of proteases during apoptosis. FASEB J. 1996; 10(5):587-597.(Biology)

Stennicke HR, Salvesen GS. Biochemical characteristics of caspases-3, -6, -7, and -8. J Biol Chem. 1997; 272(41):25719-25723.(Biology)

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