Calculation of Results

Calculate the mean absorbance for each set of duplicate standards, controls and samples. Subtract the mean zero standard absorbance from each.

Plot the standard curve on log-log graph paper, with IFN-γ concentration on the x-axis and absorbance on the y-axis. Draw the best fit curve through the standard points.

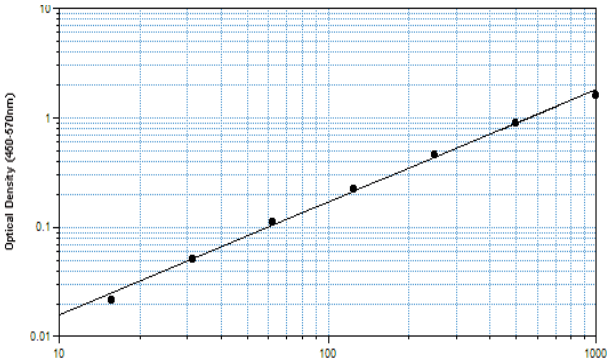
To determine the IFN-γ concentration of the unknowns, find the unknown's mean absorbance value on the y-axis and draw a horizontal line to the standard curve. At the point of intersection, draw a vertical line to the x-axis and read the IFN-γ concentration. If samples were diluted, multiply the IFN-γ concentration by the dilution factor.

Computer data reduction may also be employed, utilizing log-log regression analysis.

Typical Standard Curve

This standard curve is for demonstration only. A standard curve must be run with each assay.

BD OptEIA™ Monkey IFN-γ Set



Specificity

Cross Reactivity: The following factors were tested in the BD OptEIA™ assay at 100 ng/mL and no cross-reactivity was identified.

Recombinant Human
IL-1α, IL-1β, IL-3, IL-4, IL-5, IL-6, IL-7, IL-8, IL-10, IL-13, IL-15, TNF, LT-α (TNF-β), MCP-3, sICAM-1, sCD4, sCD14, sCD25, RANTES, TGF-β1, TNFRI, VEGF, IL-4R, IP-10, GRO-α

Standardization

The standard in this Set is calibrated against purified Baculovirus-expressed recombinant human IFN-γ. In this BD OptEIA™ Set, one monkey IFN-γ unit (U) is defined as one picogram of recombinant human IFN-γ.

Assay Optimization

1. BD OptEIA™ Sets allow flexible assay design to fit individual laboratory needs. To design an immunoassay with different sensitivity and dynamic range, the following parameters can be varied: Capture, Detection Antibody titers, Incubation time, Incubation temperature, Assay Diluent formulation, Buffer pH, ionic strength, protein concentration, Type of substrate, Washing technique (i.e., number of wash repetitions and soak times)
2. "Typical Standard Curve" and 20-plate yield were obtained in the BD Biosciences Pharmingen laboratory, using the recommended procedure and manual plate washing.

Troubleshooting

Poor Precision

Possible Source	*Corrective Action*
• Inadequate washing/ aspiration of wells	• Check function of washing system
• Inadequate mixing of reagents	• Ensure adequate mixing
• Imprecise/ inaccurate pipetting	• Check/ calibrate pipettes
• Incomplete sealing of plate	• Ensure complete seal on plate

Poor Standard Curve

Possible Source	*Corrective Action*
• Improper standard handling/ dilution age of standards	• Ensure correct preparation, stor
• Incomplete washing/ aspiration of wells	• Check function of washing system
• Imprecise/ inaccurate pipetting	• Check/ calibrate pipettes
• Improper buffer/ diluent used pH	• Check buffer/ diluent preparation,

Low Absorbances

Possible Source	*Corrective Action*
• Inadequate reagent volumes added to wells	• Check/ calibrate pipettes
• Incorrect incubation times/ temperature times/reagents warmed to RT	• Ensure sufficient incubation
• Incorrect antibody titration Detector preparation	• Check Capture Ab and Working
• Improper buffer/ diluent used pH	• Check buffer/ diluent preparation,
• Overly high wash/aspiration pressure from automated plate-washer	• Utilize manual washing

Limitations of the Procedure

• Samples that generate absorbance values higher than the standard curve should be diluted with Standard Diluent and re-assayed.
• Interference by drug metabolites, soluble receptors, or other binding proteins in specimens has not been thoroughly investigated. The possibility of interference cannot be excluded.
• BD OptEIA™ Sets are intended for use as an integral unit. Do not mix reagents from different Set batches. Reagents from other manufacturers are not recommended for use in this Set.

2750KI_551492 Rev.3

BD OptEIA™

Technical Data Sheet

Monkey IFN-γ ELISA Set

Cat. No. 551492

Materials Provided

The OptEIA™ Set for monkey interferon- gamma (IFN-γ) contains the components necessary to develop enzyme-linked immunosorbent assays (ELISA) for natural or recombinant IFN-γ in non-human primate (rhesus, cynomolgus, and baboon) serum, plasma, and cell culture supernatants. Sufficient materials are provided to yield approximately 20 plates of 96-wells if the recommended storage, materials, buffer preparation, and assay procedure are followed as specified in this package.

Capture Antibody

Anti-Monkey IFN-γ monoclonal antibody

Detection Antibody

Biotinylated Anti-Monkey IFN-γ monoclonal antibody

Enzyme Reagent

Streptavidin-horseradish peroxidase conjugate (SAv-HRP)

Standards

Recombinant IFN-γ, lyophilized

Instruction / Analysis Certificate

(lot-specific)

United States
877.232.8995

Canada
888.259.0187

Europe
32.53.720.211

Japan
0120.8555.90

Asia/Pacific
65.6861.0633

Latin America/Caribbean
55.11.5185.9995

BD Biosciences

BD Biosciences Pharmingen
10975 Torreyana Road
San Diego, CA 92121
Customer/Technical Service
Tel 877.232.8995 (US)
Fax 858.812.8888
www.bdbiosciences.com

Recommended buffers, solutions

Note: Do not use sodium azide in these preparations. Sodium azide inactivates the horseradish peroxidase enzyme.

The BD OptEIA™ Reagent Set B (Cat. No 550534) containing Coating Buffer, Assay Diluent, Substrate Reagents A and B, Stop Solution and 20X Wash Buffer Concentrate is recommended.

1. **Coating Buffer** - 0.1 M Sodium Carbonate, pH 9.5
   7.13 g $NaHCO_3$, 1.59 g Na_2CO_3; q.s. to 1.0 L; pH to 9.5 with 10N NaOH.
   Freshly prepare or use within 7 days of preparation, stored at 2-8°C.
2. **Assay Diluent**- PBS* with 10% FBS#, pH 7.0. The BD Pharmingen™ Assay Diluent (Cat. No. 555213) is recommended.
   *Phosphate-Buffered Saline: 80.0 g NaCl. 11.6 g Na_2HPO_4, 2.0 g KH_2PO_4, 2.0 g KCL, q.s. to 10 L; pH to 7.0.
   #Fetal Bovine Serum: Hyclone Cat. No. SH30088 (heat-inactivated) recommended.
   Freshly prepare or use within 3 days of preparation, with 2-8°C storage.
3. **Wash Buffer** - PBS* with 0.05% Tween-20. Freshly prepare or use within 3 days of preparation, stored at 2-8°C.
4. **Substrate Solution** - Tetramethylbenzidine (TMB) and Hydrogen Peroxide. The BD Pharmingen™ TMB Substrate Reagent Set (Cat. No. 555214) is recommended.
5. **Stop Solution** - 1 M H_3PO_4 or 2 N H_2SO_4

Additional Materials Required

1. 96-well Nunc-Immuno™ polystyrene Maxisorp ELISA flat bottom plates (ThermoFisher Scientific Cat. No. 442404) are recommended
2. Microplate reader capable of measuring absorbance at 450 nm
3. Precision pipettes
4. Graduated cylinder, one liter
5. Deionized or distilled water
6. Wash bottle or automated washer
7. Log-log graph paper or automated data reduction
8. Tubes to prepare standard dilutions
9. Laboratory timer
10. Plate sealers or parafilm

Storage Information

1. Store unopened reagents at 2-8°C. Do not use reagents after expiration date, or if turbidity is evident.
2. Before use, bring all reagents to room temperature (18-25°C). Immediately after use, return to proper storage conditions.
3. Lyophilized standards are stable until expiration date. See below for reconstituted standard storage information.

Specimen Collection and Handling

Specimens should be clear, non-hemolyzed and non-lipemic.

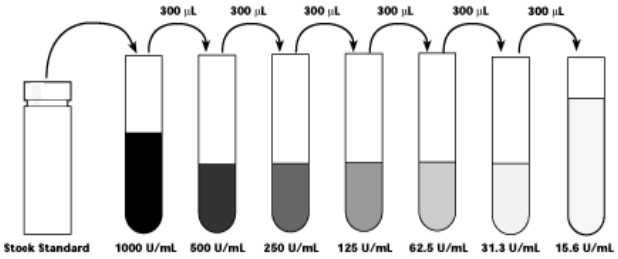
Cell culture supernatants: Remove any particulate material by centrifugation and assay immediately or store samples at ≤ -20°C. Avoid repeated freeze-thaw cycles.

Serum: Use a serum separator tube and allow samples to clot for 30 minutes, then centrifuge for 10 minutes at 1000 x g. Remove serum and assay immediately or store samples at ≤ -20° C. Avoid repeated freeze-thaw cycles.

Plasma: Collect plasma using citrate, EDTA, or heparin as anticoagulant. Centrifuge for 10 minutes at 1000 x g within 30 minutes of collection. Assay immediately or store samples at ≤ -20° C. Avoid repeated freeze-thaw cycles.

Standards Preparation and Handling

1. **Reconstitution:** After warming lyophilized standard to room temperature, carefully open vial to avoid loss of material. Reconstitute lyophilized standard with 1.0 mL of deionized water to yield a stock standard. Allow the standard to equilibrate for at least 15 minutes before making dilutions. Vortex gently to mix.
2. **Storage/ handling of reconstituted standard:** After reconstitution, **immediately** aliquot standard stock in polypropylene vials at 50 µl per vial and freeze at -80°C for up to 6 months. If necessary, store at 2-8° C for up to 8 hours prior to aliquotting/freezing. Do not leave reconstituted standard at room temperature.
3. **Standards Preparation for Assay:**
   a. Prepare a 1000 U/mL standard from the stock standard. Vortex to mix. (See dilution instructions on Instruction/Analysis Certificate.)
   b. Add 300 µL Assay Diluent to 6 tubes. Label as 500 U/mL, 250 U/mL, 125 U/mL, 62.5 U/mL, 31.3 U/mL, and 15.6 U/mL.
   c. Perform serial dilutions by adding 300 µL of each standard to the next tube and vortexing between each transfer. Assay Diluent serves as the zero standard (0 U/mL).



Serial dilutions within the plate may also be performed by pipetting 100 µL of Assay Diluent into each standard well except the highest (1000 U/mL), then adding 100 µL of the 1000 U/mL standard to both that well and the 500 U/mL well, mixing the well contents by rinsing the pipette tip, and adding 100 µL of the 500 U/mL standard to the 250 U/mL well. Continue these dilutions to the 15.6 U/mL standard well, out of which the extra 100 µL should be discarded.

Working Detector Preparation

(Note: One-step incubation of Biotin/Streptavidin reagents.) Add required volume of Detection Antibody to Assay Diluent. Within 15 minutes prior to use, add required quantity of Enzyme Reagent, vortex or mix well. For recommended dilutions, see lot-specific Instruction/Analysis Certificate. For a full 96-well plate, prepare 12 mL of Working Detector. Discard any remaining Working Detector after use.

Warnings and Precautions

1. Reagents which contain preservatives may be toxic if ingested, inhaled, or in contact with skin.
2. Handle all serum and plasma specimens in accordance with NCCLS guidelines for preventing transmission of blood-borne infections.
3. Capture Antibody contains < 0.1% sodium azide. Sodium azide yields highly toxic hydrazoic acid under acidic conditions. Dilute azide compounds in running water before discarding to avoid accumulation of potentially explosive deposits in plumbing.
4. Detection Antibody contains BSA and ProClin®-150 as a preservative.
5. Enzyme Reagent contains BSA and ProClin®-150 as a preservative.
6. Source of all serum proteins is from USDA inspected abattoirs located in the United States.

Recommended Assay Procedure

1. Coat microwells with 100 µL per well of Capture Antibody diluted in Coating Buffer. For recommended antibody coating dilution, see lot-specific Instruction/Analysis Certificate. Seal plate and incubate overnight at 4° C.
2. Aspirate wells and wash 3 times with ≥ 300 µL/well Wash Buffer. After last wash, invert plate and blot on absorbent paper to remove any residual buffer.
3. Block plates with ≥ 200 µL/well Assay Diluent. Incubate at RT for 1 hour.
4. Aspirate/wash as in step 2.
5. Prepare standard and sample dilutions in Assay Diluent. See "Standards Preparation and Handling".
6. Pipette 100 µL of each standard, sample, and control into appropriate wells. Seal plate and incubate for 2 hours at RT.
7. Aspirate/ wash as in step 2, but with 5 total washes.
8. Add 100 µL of Working Detector (Detection Antibody + Streptavidin-HRP reagent) to each well. Seal plate and incubate for 1 hour at RT.
9. Aspirate/ wash as in step 2, but with 7 total washes. NOTE: In this final wash step, soak wells in wash buffer for 30 seconds to 1 minute for each wash.
10. Add 100 µL of Substrate Solution to each well. Incubate plate (without plate sealer) for 30 minutes at room temperature in the dark.
11. Add 50 µL of Stop Solution to each well.
12. Read absorbance at 450 nm within 30 minutes of stopping reaction. If wavelength correction is available, subtract absorbance at 570 nm from absorbance 450 nm.

Assay Procedure Summary

1. Add 100 µL diluted Capture Ab to each well. Incubate overnight at 4° C.
2. Aspirate and wash 3 times.
3. Block plates: 200 µL Assay Diluent to each well. Incubate 1 hr RT
4. Aspirate and wash 3 times.
5. Add 100 µL standard or sample to each well. Incubate 2 hr RT.
6. Aspirate and wash 5 times.
7. Add 100 µL Working Detector (Detection Ab + SAv-HRP) to each well. Incubate 1 hr RT
8. Aspirate and wash 7 times (with 30 sec to 1 min soaks)
9. Add 100 µL Substrate Solution to each well. Incubate 30 min RT in dark
10. Add 50 µL Stop Solution to each well. Read at 450 nm within 30 min with λ correction 570 nm.