# Purified Mouse Anti-Human Caspase-2 w/Control

Material Number:	551093	
Reactivity:	QC Testing: Human	
Component:	51-1395GR	
Description:	Purified Mouse Anti-Human Caspase-2 (Ich-1)	
Size:	50 µg (1 ea)	
Concentration:	0.25 mg/ml	
Clone Name:	G310-1248	
Immunogen:	Human Caspase-2 (short)	
Isotype:	Mouse IgG1	
Storage Buffer:	Aqueous buffered solution containing BSA, glycerol, and ≤0.09% sodium	
	azide.	
Component:	51-16526N	
Description:	Jurkat Cell Lysate	
Size:	50 µg (1 ea)	
Concentration:	1.0 mg/ml	
Storage Buffer:	SDS-PAGE buffer (62mM Tris pH 6.8, 2% SDS, 0.9% b-mercaptoethanol,	
	0.003% bromophenol blue, 5% glycerol)	

## Description

Caspase-2 (Ich-1) belongs to the ICE/CED-3 cell death gene family of cysteine proteases. CED-3 is a *C. elegans* death gene that is required for normal programmed cell death during development. ICE (interleukin-1ß-converting enzyme) is the mammalian homolog of CED-3 and is activated when cells become apoptotic. Ich1 mRNA is alternatively spliced into two different forms, Ich-1 long (Ich-1L) and Ich-1 short (Ich-1S). Ich-1L is a 48 kDa protein that has about 50% amino acid sequence homology to both CED-3 and ICE. Ich-1S (35 kDa) is a truncated version of Ich-1L. Ich-1L and Ich-1S have been found to have opposite effects on cell death in vitro. Overexpression of Ich-1L in Rat1 rat fibroblast cells induced programmed cell death, whereas overexpression of Ich-1S suppressed Rat-1 cell death induced by serum starvation. These results suggest that Ich-1 may play a role in both the positive and negative regulation of programmed cell death.

The G310-1248 clone was made using recombinant full-length human caspase-2 (short) [Ich-1S] as the immunogen and can recognize both the short and long forms of human caspase-2.

# **Preparation and Storage**

The monoclonal antibody was purified from tissue culture supernatant or ascites by affinity chromatography. Store both the antibody and Jurkat control lysate undiluted at -20°C.

# **Application Notes**

Application	
Western blot	Routinely Tested
Bioimaging	Tested During Development

## **Recommended Assay Procedure:**

#### Bioimaging

- 1. Seed the cells in appropriate culture medium at ~10,000 cells per well in a BD Falcon<sup>™</sup> 96-well Imaging Plate (Cat. No. 353219) and culture overnight.
- 2. Remove the culture medium from the wells, and fix the cells by adding 100 µl of BD Cytofix<sup>™</sup> Fixation Buffer (Cat. No. 554655) to each well. Incubate for 10 minutes at room temperature (RT).
- 3. Remove the fixative from the wells, and permeabilize the cells using either BD Perm Buffer III, 90% methanol, or Triton<sup>™</sup> X-100: a. Add 100 µl of -20°C 90% methanol, Perm Buffer III (Cat. No. 558050) to each well and incubate for 5 minutes at RT.

OR

- b. Add 100 µl of 0.1% Triton<sup>™</sup> X-100 to each well and incubate for 5 minutes at RT.
- 4. Remove the permeabilization buffer, and wash the wells twice with 100  $\mu$ l of 1× PBS.
- 5. Remove the PBS, and block the cells by adding 100 µl of BD Pharmingen<sup>™</sup> Stain Buffer (FBS) (Cat. No. 554656) to each well. Incubate for 30 minutes at RT.

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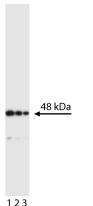




- 6. Remove the blocking buffer and add 50 μl of the optimally titrated primary antibody (diluted in Stain Buffer) to each well, and incubate for 1 hour at RT.
- 7. Remove the primary antibody, and wash the wells three times with 100  $\mu$ l of 1× PBS.
- Remove the PBS, and add the second step reagent at its optimally titrated concentration in 50 μl to each well, and incubate in the dark for 1 hour at RT.
- 9. Remove the second step reagent, and wash the wells three times with 100 µl of 1× PBS.
- 10. Remove the PBS, and counter-stain the nuclei by adding 200  $\mu$ l per well of 2  $\mu$ g/ml Hoechst 33342 (e.g., Sigma-Aldrich Cat. No. B2261) in 1× PBS to each well at least 15 minutes before imaging.
- 11. View and analyze the cells on an appropriate imaging instrument.

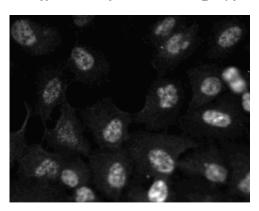
Bioimaging: For more detailed information please refer to http://www.bdbiosciences.com/support/resources/protocols/ceritifed\_reagents.jsp

Western blot: For more detailed information please refer to http://www.bdbiosciences.com/support/resources/protocols/monoclonal\_anti.jsp



ΙZ

Western blot analysis of Caspase-2. Lysate from Jurkat cells were probed with the Caspase-2 antibody at concentrations of 0.5  $\mu$ g/mL (lane 1), 0.25  $\mu$ g/mL (lane 2), and 0.125  $\mu$ g/mL (lane 3). Caspase-2 is identified as a band of 48 kDa.



Immunofluorescent staining of U-2 OS (ATCC HTB-96) cells. Cells were seeded in a 96 well imaging plate (Cat. No. 353219) at ~10,000 cells per well. After overnight incubation, cells were stained using the alcohol perm protocol and the anti-caspase-2 antibody. The second step reagent was FITC goat anti mouse Ig (Cat. No. 554001). Images were taken on a BD Pathway™ 855 bioimaging system using a 20x objective. This antibody also stained HeLa (ATCC CCL-2) and A549 (ATCC CCL-185) cells and worked with both the Triton™ X-100 and alcohol fix/perm protocols (see Recommended Assay Procedure).

#### **Suggested Companion Products**

Catalog Number	Name	Size	Clone
611451	Jurkat Cell Lysate	500 µg	(none)
554001	FITC Goat Anti-Mouse Ig	0.5 mg	Polyclonal
554002	HRP Goat Anti-Mouse Ig	1.0 ml	(none)
353219	BD Falcon <sup>™</sup> 96-well Imaging Plate	NA	(none)
554655	Fixation Buffer	100 ml	(none)
558050	Perm Buffer III	125 ml	(none)
554656	Stain Buffer (FBS)	500 ml	(none)

## **Product Notices**

- 1. Since applications vary, each investigator should titrate the reagent to obtain optimal results.
- 2. This antibody has been developed and certified for the bioimaging application. However, a routine bioimaging test is not performed on every lot. Researchers are encouraged to titrate the reagent for optimal performance.
- 3. Source of all serum proteins is from USDA inspected abattoirs located in the United States.
- 4. Triton is a trademark of the Dow Chemical Company.
- 5. Caution: Sodium azide yields highly toxic hydrazoic acid under acidic conditions. Dilute azide compounds in running water before discarding to avoid accumulation of potentially explosive deposits in plumbing.
- 6. Please refer to www.bdbiosciences.com/pharmingen/protocols for technical protocols.