GoTaq® qPCR Master Mix

INSTRUCTIONS FOR USE OF PRODUCTS A6001 AND A6002.

Quick PROTOCOL

Use of the GoTaq® qPCR Master Mix

The protocol for a 50µl reaction is outlined below. Component volumes may be scaled as appropriate. This protocol assumes that 20% of the reaction volume is DNA template (e.g., 10µl of DNA template added to 40µl of reaction mix). If the volume of DNA template is more or less than 10µl, adjust the volume of Nuclease-Free Water accordingly so that the final reaction volume is 50µl.

Preparing the DNA Template

1. Prepare standards and experimental template dilutions in nuclease-free water.
2. Add 10µl of DNA (or water for no-template control reactions) to the appropriate wells of the reaction plate.

Preparing the qPCR Reaction Mix

1. Prepare the reaction mix by combining GoTaq® qPCR Master Mix, Nuclease-Free Water and primers in the order listed. See Notes 1 and 2.

Component	Volume per 50µl Reaction	Final Concentration
GoTaq® qPCR Master Mix, 2X	25µl	1X
Nuclease-Free Water	to a final volume of 40µl	
upstream and downstream PCR primers	_____µl	0.2µM or 0.05–0.9µM each

Notes

1. See the *GoTaq® qPCR Master Mix Technical Manual* #TM318 for a list of instruments that require addition of CXR Reference Dye.
2. Some instruments such as the BioRad instruments require addition of a normalization dye (e.g., fluorescein).

2. Carefully pipet 40µl of reaction mix into each reaction well.
3. Seal the plate, and centrifuge at low speed for 1 minute.
4. Program the thermal cycler with the desired thermal cycling conditions as per the manufacturer's instructions.
5. Place the plate in thermal cycler, and press "Start".

When the run is complete, analyze the data using your usual procedures.

For complete protocol information see the *GoTaq® qPCR Master Mix Technical Manual* #TM318, available online at: **www.promega.com/tbs/**

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Printed in USA. Revised. 11/10
Part #9FB103