

ORDERING INFORMATION

Catalog Number: AF1573

Lot Number: JLE01

Size: 100 µg

Formulation: 0.2 µm filtered solution in PBS with 5% trehalose

Storage: -20° C

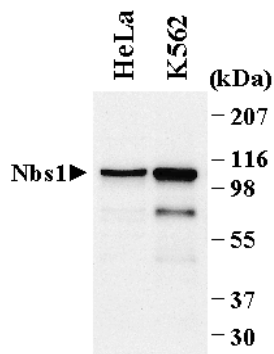
Reconstitution: sterile PBS and 0.02% NaN₃

Specificity: human Nbs1

Immunogen: *E. coli*-derived recombinant human Nbs1 (amino acids 498 - 754)

Ig Type: affinity-purified goat IgG

Applications: Western blot
Immunoprecipitation



Extracts from exponentially growing human cell lines (HeLa and K562) were prepared, resolved by SDS-PAGE, and transferred to a PVDF membrane. The membrane was immunoblotted with 0.5 µg/mL goat anti-Nbs1 antibody.

Background

The Nijmegen Breakage Syndrome 1 (Nbs1) protein is a member of the Mre11/Rad50/Nbs1 (MRN) protein complex that binds to DNA double-strand breaks in cells exposed to DNA damaging agents. In addition, the MRN complex colocalizes with replication forks during DNA replication. The MRN complex plays an important role in routine cell cycle progression and genotoxic stress responses by facilitating DNA repair. In fact, mutation of the *nbs1* gene and resultant loss of Nbs1 protein expression in humans results in the chromosomal instability disease, Nijmegen Breakage Syndrome.

Preparation

Goat antibodies were raised against purified, *E. coli*-derived, recombinant human Nbs1 (rhNbs1). Polyclonal antibody was affinity-purified on a column derivatized with rhNbs1, and further purified by isolating the IgG fraction.

Formulation

Lyophilized from a 0.2 µm filtered solution in phosphate-buffered saline (PBS) with 5% trehalose.

Reconstitution

Reconstitute the antibody with 100 µl of sterile PBS containing 0.02% NaN₃.

Storage

The reconstituted antibody should be aliquoted and stored at -20° C in a manual defrost freezer until use. **Avoid repeated freeze/thaw cycles.**

Specificity

The antibody detects human Nbs1.

Application

Western blot - An antibody concentration of 0.5 - 1.0 µg/mL is recommended.

Immunoprecipitation - Use 2 µg antibody per 500 µg cell extract.

Protocols for Immunoprecipitation and Immunoblotting:

Immunoprecipitation

Wash Buffer

50 mM Tris, pH 7.4
150 mM NaCl
1 % NP-40
1 mM DTT

Cell Lysis Buffer

Wash Buffer containing:
3 µg/mL aprotinin
2 µg/mL leupeptin
2 µg/mL pepstatin A

Cell lysates for immunoprecipitation: Wash cells twice with cold PBS and extract cell protein by solubilization of 1×10^6 - 5×10^6 cells in 1 ml cold Cell Lysis Buffer. Solubilize cells for 15 minutes on ice, followed by centrifugation at 6000 x g for 5 minutes to clear insoluble material. Measure protein concentration and bring volume up to 1 ml per sample with Cell Lysis Buffer.

Immunoprecipitation: Add 2 µg goat anti-Nbs1 antibody per 500 µg cell extract and incubate 1 hour on ice with occasional inversion. Add Protein G agarose (20 µL of a 50% suspension) to each sample and rotate 1 hour at 4° C. Pellet the Protein G-absorbed complexes and wash twice with Wash Buffer. Suspend the washed pellet in 25 - 50 µL 2X SDS gel sample buffer (see above) and incubate 5 minutes in boiling water bath. Pellet Protein G agarose and resolve the supernatant by SDS-PAGE.

Western blotting

Blotting Buffer

25 mM Tris, pH 7.5
0.15 M NaCl
0.05% Tween 20

Blocking Solution

5% nonfat dry milk
in blotting buffer
pH to 7.5

1. Transfer the electrophoresed proteins onto a PVDF membrane and incubate the membrane for 1 hour at room temperature in Blocking Solution.
2. Incubate the membrane for 2 hours at room temperature or overnight at 2 - 8° C in Blocking Solution containing 0.5 - 1.0 µg/mL goat anti-Nbs1 antibody.
3. Wash the membrane at room temperature for 30 minutes with 3 or more changes of Blotting Buffer. Changing the membrane containers often reduces background.
4. Incubate the membrane at room temperature for 1 hour in Blocking Solution containing a 1:2,000 dilution of HRP-conjugated donkey anti-goat Ig (R&D Systems, Catalog # HAF109).
5. Wash the membrane for 30 minutes with 3 or more changes of Blotting Buffer.
6. Detect with ECL Reagent.

Cell lysates for western blotting: To prepare total cell lysates, solubilize cells in 2X SDS gel sample buffer (20 mM dithiothreitol, 6% SDS, 0.25 M Tris, pH 6.8, 10% glycerol, and bromophenyl blue) and sonicate with a probe sonicator using 3 - 4 bursts of 5 - 10 seconds each. Heat extracts in a boiling water bath for 5 minutes and load onto polyacrylamide gels. Samples may be diluted with 1X SDS sample buffer to the desired concentration.

Optimal dilutions should be determined by the individual laboratory.