PRODUCT INFORMATION & MANUAL

Human sTNF-R (60kDa) Instant ELISA

BMS203INST

Enzyme-linked immunosorbent assay for quantitative detection of human sTNF-R (60kDa).
For research use only.
Not for diagnostic or therapeutic procedures.
128 Tests

eBioscience
An Affymetrix Company

Human sTNF-R (60kDa) Instant ELISA

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1 Intended Use

The human sTNF-R (60kDa) Instant ELISA is an enzyme-linked immunosorbent assay for the quantitative detection of soluble human 60 kDa Tumor Necrosis Factor-Receptor levels in cell culture supernatants, human serum, plasma, or other body fluids. The human sTNF-R (60kDa) Instant ELISA is **for research use only. Not for diagnostic or therapeutic procedures.**

2 Summary

Tumor Necrosis Factor (TNF) was originally discovered in sera of animals and was found to cause hemorrhagic necrosis of some transplantable mouse and human tumors and to exhibit primarily cytotoxic activities against tumor but not normal cells in vitro. The TNF family consists of two proteins designated TNF-α, also called cachectin, and TNF-β, also called lymphotoxin, which are pleiotropic cytokines that can mediate a wide variety of biological effects.

Both TNF-α and TNF-β have been shown to interact with a cell through specific high affinity receptors with a few hundred up to more than 20,000 copies per cell. TNF-receptors have been demonstrated on a wide variety of human somatic cells including fibroblasts, endothelial cells, adipocytes, liver membranes, granulocytes and several tumor cell lines. Normal and malignant human myeloid cells as well as mitogen-stimulated lymphocytes express similar numbers of TNF receptors (400 - 1,900 per cell), whereas resting lymphoid cells have fewer, red blood cells and platelets have no detectable TNF receptors.

In most cases no correlation is observed between receptor number and sensitivity to TNF. Based on gel filtration experiments the receptor appears to be a complex of different proteins with a molecular weight of 350 kDa. In a variety of cell lines two different types of TNF receptors with 75-80 and 55-60 kDa respectively have been identified.

The cDNAs encoding the two different TNF receptors have been cloned. The exact mechanism of signal transduction after binding of TNF to the receptor is still unclear. The extracellular fragment of the 60 kDa TNF receptor, with a molecular mass of about 30 kDa was purified, partially sequenced, and the respective cDNA was cloned. This TNF binding

protein is liberated from the intact molecule by proteolytic cleavage and comprises most of the extracellular portion of the receptor, including all three N-glycosylation sites.

The present assay, slightly modified from the assay established by Adolf and Apfler, provides a simple, rapid, and highly sensitive method for the determination of soluble TNF-R (60 kDa) levels in body fluids or cell culture supernatants. This assay will help to clarify the possible diagnostic and prognostic value of circulating sTNF-R (60 kDa) in various neoplastic and inflammatory diseases.

For literature update refer to **www.eBioscience.com**

3 Principles of the Test

An anti-human sTNF-R (60kDa) monoclonal coating antibody is adsorbed onto microwells. Human sTNF-R (60kDa) present in the sample or standard binds to antibodies adsorbed to the microwells; an HRP-conjugated monoclonal anti-human sTNF-R (60kDa) antibody binds to human sTNF-R (60kDa) captured by the first antibody. Following incubation unbound enzyme conjugated anti-human sTNF-R (60kDa) is removed during a wash step and substrate solution reactive with HRP is added to the wells.

Figure 1

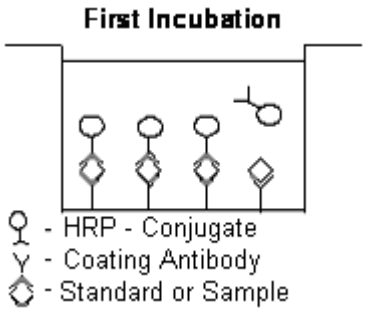
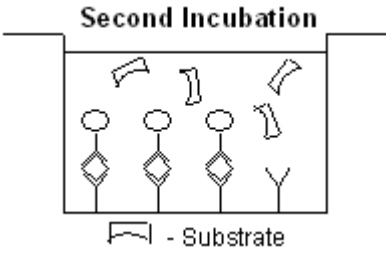
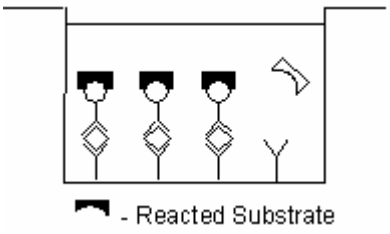


Figure 2



A coloured product is formed in proportion to the amount of human sTNF-R (60kDa) present in the sample. The reaction is terminated by addition of acid and absorbance is measured at 450 nm. A standard curve is prepared from seven human sTNF-R (60kDa) standard dilutions and human sTNF-R (60kDa) sample concentration determined.

Figure 3



4 Reagents Provided

1 aluminium pouch with **a Microwell Plate coated with Monoclonal Antibody** (murine) to human sTNF-R (60 kDa) and **HRP-Conjugate** (anti-sTNF-R (60 kDa) monoclonal (murine) antibody) lyophilized

2 aluminium pouches with a **human sTNF-R (60kDa) Standard curve (coloured)**

1 bottle (25 ml) **Wash Buffer Concentrate** 20x
(phosphate-buffered saline with 1% Tween 20)

1 vial (15 ml) **Substrate Solution** (tetramethyl-benzidine)

1 vial (5 ml) **Assay Buffer Concentrate** 20x
(Use when an external predilution of the samples is needed)

1 vial (15 ml) **Stop Solution** (1M Phosphoric acid)

2 adhesive **Plate Covers**

5 Storage Instructions

Store ELISA plate and Standard curves or whole kit at -20°C. The plate and the standard curves can also be removed, stored at -20°C, remaining kit reagents can be stored between 2° and 8°C. Expiry of the kit and reagents is stated on labels.
The expiry of the kit components can only be guaranteed if the components are stored properly, and if, in case of repeated use of one component, the reagent is not contaminated by the first handling.

6 Specimen Collection

Cell in cell culture supernatants, human serum and plasma were tested with this assay. Other biological samples might be suitable for use in the assay. Remove the serum or plasma from the clot or red cells as soon as possible after clotting and separation.

Samples containing a visible precipitate must be clarified prior to use in the assay. Do not use grossly hemolyzed or lipemic specimens.

Samples must be stored frozen at -20°C to avoid loss of bioactive human sTNF-R (60kDa). If samples are to be run within 24 hours, they may be stored at 2° to 8°C (for sample stability refer to 13).
Avoid repeated freeze-thaw cycles. Prior to assay, frozen serum or plasma should be brought to room temperature slowly and mixed gently.

7 Materials Required But Not Provided

- 5 ml and 10 ml graduated pipettes
- 5 µl to 1000 µl adjustable single channel micropipettes with disposable tips
- 50 µl to 300 µl adjustable multichannel micropipette with disposable tips
- Multichannel micropipette reservoir
- Beakers, flasks, cylinders necessary for preparation of reagents
- Device for delivery of wash solution (multichannel wash bottle or automatic wash system)
- Microwell strip reader capable of reading at 450 nm (620 nm as optional reference wave length)
- Glass-distilled or deionized water
- Statistical calculator with program to perform linear regression analysis

8 Precautions for Use

- All chemicals should be considered as potentially hazardous. We therefore recommend that this product is handled only by those persons who have been trained in laboratory techniques and that it is used in accordance with the principles of good laboratory practice. Wear suitable protective clothing such as laboratory overalls, safety glasses and gloves. Care should be taken to avoid contact with skin or eyes. In the case of contact with skin or eyes wash immediately with water. See material safety data sheet(s) and/or safety statements(s) for specific advice.
- Reagents are intended for research use only and are not for use in diagnostic or therapeutic procedures.
- Do not mix or substitute reagents with those from other lots or other sources.
- Do not use kit reagents beyond expiration date on label.
- Do not expose kit reagents to strong light during storage or incubation.
- Do not pipette by mouth.
- Do not eat or smoke in areas where kit reagents or samples are handled.
- Avoid contact of skin or mucous membranes with kit reagents or specimens.
- Rubber or disposable latex gloves should be worn while handling kit reagents or specimens.
- Avoid contact of substrate solution with oxidizing agents and metal.
- Avoid splashing or generation of aerosols.
- In order to avoid microbial contamination or cross-contamination of reagents or specimens which may invalidate the test use disposable pipette tips and/or pipettes.
- Use clean, dedicated reagent trays for dispensing substrate reagent.

- Glass-distilled water or deionized water must be used for reagent preparation.
- Substrate solution must be at room temperature prior to use.
- Decontaminate and dispose specimens and all potentially contaminated materials as they could contain infectious agents. The preferred method of decontamination is autoclaving for a minimum of 1 hour at 121.5°C.
- Liquid wastes not containing acid and neutralized waste may be mixed with sodium hypochlorite in volumes such that the final mixture contains 1.0% sodium hypochlorite. Allow 30 minutes for effective decontamination. Liquid waste containing acid must be neutralized prior to the addition of sodium hypochlorite.

9 Preparation of Reagents and Samples

Buffer concentrate should be brought to room temperature and diluted before starting the test procedure. If crystals have formed in the buffer concentrate, warm it gently until crystals have completely dissolved.

9.1 Wash Buffer (1x)

Pour entire contents (25 ml) of the Wash Buffer Concentrate (20x) into a clean 500 ml graduated cylinder. Bring to final volume to 500 ml with glass-distilled or deionized water. Mix gently to avoid foaming.

Transfer to a clean wash bottle and store at 2° to 25°C. Please note that Wash Buffer (1x) is stable for 30 days.

9.2 Assay Buffer (1x)

Pour the entire contents (5ml) of the **Assay Buffer Concentrate** into a clean 100 ml graduated cylinder. Bring to final volume of 100 ml with distilled water. Mix gently.

10 Test Protocol

- **Use plate immediately after removal from -20°C!**
- **Do not wait until pellets have completely dissolved before applying samples - the binding reaction in the standard strips starts immediately after addition of water!**
- **Do not try to dissolve pellets by pipetting up and down in the wells - some parts of the pellet could stick to the tip creating high variation of results**
- **Perform the washing step with at least 400 µl of washing buffer as stated in the manual or fill the wells completely - otherwise any pellet residues sticking to the rim of the well will not be removed and create high variation of results**
- **Allow the washing buffer to sit in the wells for a few seconds before aspiration**
- **Remove covers of the standard strips carefully in order that all the lyophilised pellets remain in the wells**

a. Determine the number of Microwell Strips required to test the desired number of samples plus Microwell Strips for blanks and standards (coloured). Each sample, standard, blank, and optional control sample should be assayed in duplicate. Remove extra Microwell Strips from holder and store in foil bag with the desiccant provided at -20°C sealed tightly. Place microwell strips containing the standard curve in position A1/A2 to H1/H2 (see Table 1).

b. Add **distilled water** to all **standard and blank wells** as indicated on the label of the standard strips (A1, A2 to H1, H2).

c. Add 140 µl of **distilled water** to the **sample wells**.

Table 1

Table depicting an example of the arrangement of blanks, standards and samples in the microwell strips:

	1	**2**	**3**	**4**
A	Standard 1 (5.000 ng/ml)	Standard 1 (5.000 ng/ml)	Sample 1	Sample 1
B	Standard 2 (2.500 ng/ml)	Standard 2 (2.500 ng/ml)	Sample 2	Sample 2
C	Standard 3 (1.250 ng/ml)	Standard 3 (1.250 ng/ml)	Sample 3	Sample 3
D	Standard 4 (0.625 ng/ml)	Standard 4 (0.625 ng/ml)	Sample 4	Sample 4
E	Standard 5 (0.313 ng/ml)	Standard 5 (0.313 ng/ml)	Sample 5	Sample 5
F	Standard 6 (0.156 ng/ml)	Standard 6 (0.156 ng/ml)	Sample 6	Sample 6
G	Standard 7 (0.078 ng/ml)	Standard 7 (0.078 ng/ml)	Sample 7	Sample 7
H	Blank	Blank	Sample 8	Sample 8

d. Add 10 µl of each **Sample**, in duplicate, to the designated wells and mix the contents.

e. Cover with a **Plate Cover** and incubate at room temperature (18ºC to 25ºC) for 2 hours, if available on a microplate shaker at 400 rpm.

f. Remove **Plate Cover** and empty wells. Wash the microwell strips 3 times with approximately 400 µl Wash Buffer per well with thorough aspiration of microwell contents between washes. Take care not to scratch the surface of the microwells.

   After the last wash, tap microwell strips on absorbent pad or paper towel to remove excess Wash Buffer. Use the microwell strips immediately after washing or place upside down on a wet absorbent paper for no longer than 15 minutes. Do not allow wells to dry.

g. Pipette 100 µl of **TMB Substrate Solution** to all wells, including the blank wells.

h. Incubate the microwell strips at room temperature (18° to 25°C) for about 10 minutes. Avoid direct exposure to intense light.

   The colour development on the plate should be monitored and the substrate reaction stopped (see point i. of this protocol) before positive wells are no longer properly recordable. Determination of the ideal time period for colour development has to be done individually for each assay.
   It is recommended to add the stop solution when the highest standard has developed a dark blue colour.
   Alternatively the colour development can be monitored by the ELISA reader at 620 nm. The substrate reaction should be stopped as soon as Standard 1 has reached an OD of 0.9 – 0.95.

i. Stop the enzyme reaction by quickly pipetting 100 µl of **Stop Solution** into each well, including the blank wells. It is important that the Stop Solution is spread quickly and uniformly throughout the microwells to completely inactivate the enzyme. Results must be read immediately after the Stop Solution is added or within one hour if the microwell strips are stored at 2 - 8°C in the dark.

j. Read absorbance of each microwell on a spectro-photometer using 450 nm as the primary wave length (optionally 620 nm as the

reference wave length; 610 nm to 650 nm is acceptable). Blank the plate reader according to the manufacturer's instructions by using the blank wells. Determine the absorbance of both the samples and the human sTNF-R (60kDa) standards.

Note: In case of incubation without shaking the obtained O.D. values may be lower than indicated below. Nevertheless the results are still valid.

11 Calculation of Results

- Calculate the average absorbance values for each set of duplicate standards and samples. Duplicates should be within 20 per cent of the mean.
- Create a standard curve by plotting the mean absorbance for each standard concentration on the ordinate against the human sTNF-R (60kDa) concentration on the abscissa. Draw a best fit curve through the points of the graph.
- To determine the concentration of circulating human sTNF-R (60kDa) for each sample, first find the mean absorbance value on the ordinate and extend a horizontal line to the standard curve. At the point of intersection, extend a vertical line to the abscissa and read the corresponding human sTNF-R (60kDa) concentration.
- ***Samples have been diluted 1:10, thus the concentration read from the standard curve must be multiplied by the dilution factor (x 10).**
- **Calculation of samples with a concentration exceeding standard 1 may result in incorrect, low human sTNF-R (60kDa) levels). Such samples require further external predilution according to expected human sTNF-R (60kDa) values with Assay Buffer in order to precisely quantitate the actual human sTNF-R (60kDa).**
- It is suggested that each testing facility establishes a control sample of known human sTNF-R (60kDa) concentration and runs this additional control with each assay. If the values obtained are not within the expected range of the control, the assay results may be invalid.
- A representative standard curve is shown in Figure 4. This curve cannot be used to derive test results. Every laboratory must prepare a standard curve for each group of microwell strips assayed.

* N.B: There is a common dilution factor for samples due to the conjugate which must then be included in the calculation. The samples contribute 100 µl to the final volume per well. These 100 µl are composed of 90 µl of sample diluent plus 10 µl of the sample. This is a 1:10 dilution.
The remaining 50 µl to give 150 µl are due to the addition of 50 µl conjugate to all wells.
90 µl sample diluent and 50 µl conjugate results in 140 µl reconstitution volume, addition of 10 µl sample (90 µl + 10 µl = 1:10 dilution)

Figure 4

Representative standard curve for human sTNF-R (60kDa) Instant ELISA. Human sTNF-R (60kDa) was diluted in serial 2-fold steps in Assay Buffer, each symbol represents the mean of 3 parallel titrations. Do not use this standard curve to derive test results. A standard curve must be run for each group of microwell strips assayed.

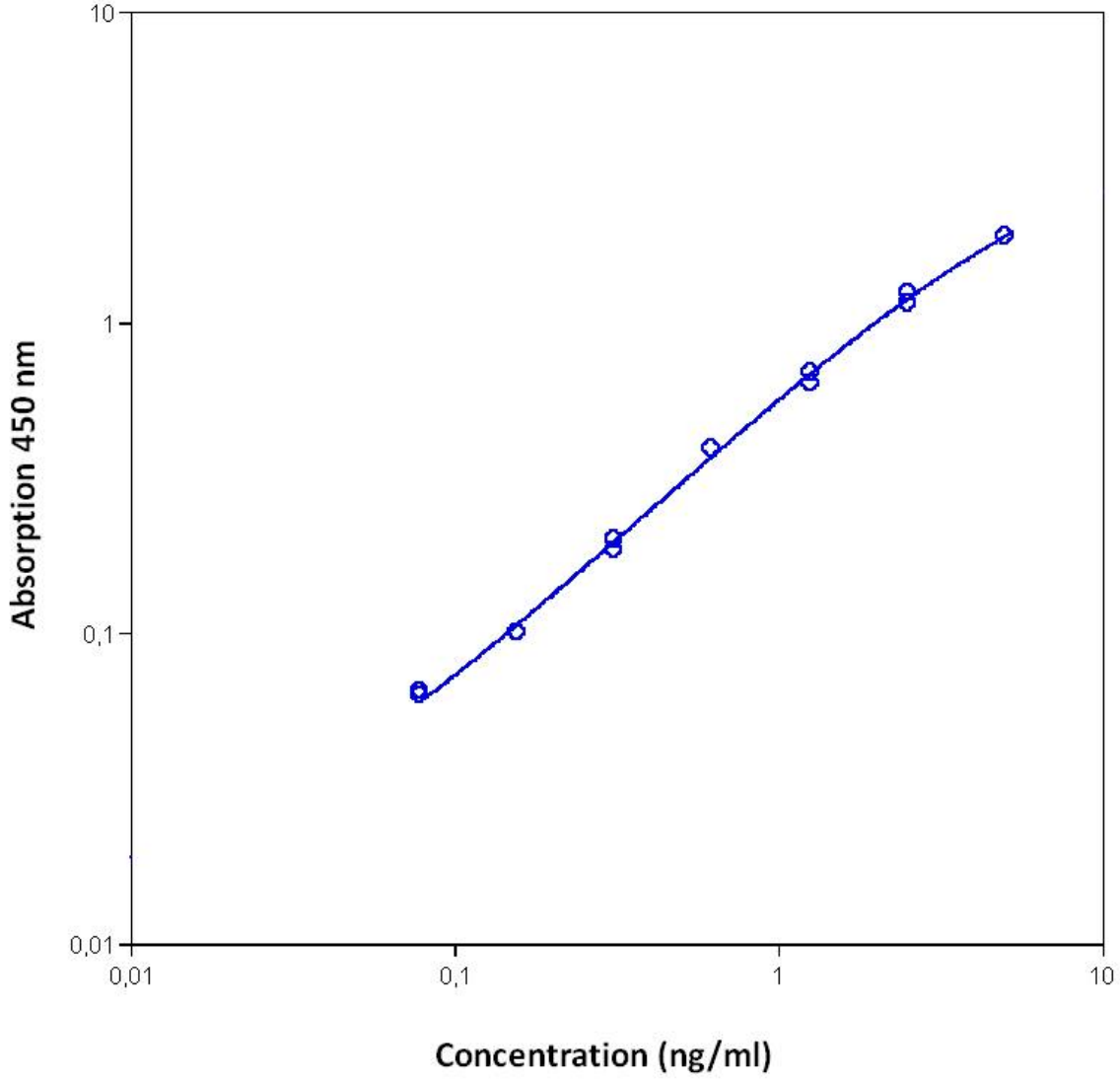


Table 2

Typical data using the human sTNF-R (60kDa) INSTANT ELISA
Measuring wavelength: 450 nm
Reference wavelength: 620 nm

Standard	human sTNF-R (60kDa) Concentration (ng/ml)	O.D. (450 nm)	O.D. Mean	C.V. (%)
1	5.000	1.876 1.895	1.886	0.7
2	2.500	1.144 1.243	1.194	5.9
3	1.250	0.635 0.690	0.663	5.9
4	0.625	0.395 0.392	0.394	0.5
5	0.313	0.199 0.184	0.192	5.5
6	0.156	0.101 0.100	0.101	0.7
7	0.078	0.063 0.065	0.064	2.2
Blank	0.000	0.013 0.014	0.014	5.2

The OD values of the standard curve may vary according to the conditions of assay performance (e.g. operator, pipetting technique, washing technique or temperature effects). Furthermore shelf life of the kit may affect enzymatic activity and thus colour intensity. Values measured are still valid.

12 Limitations

- Since exact conditions may vary from assay to assay, a standard curve must be established for every run.
- Bacterial or fungal contamination of either screen samples or reagents or cross-contamination between reagents may cause erroneous results.
- Disposable pipette tips, flasks or glassware are preferred, reusable glassware must be washed and thoroughly rinsed of all detergents before use.
- Improper or insufficient washing at any stage of the procedure will result in either false positive or false negative results. Empty wells completely before dispensing fresh wash solution, fill with Wash Buffer as indicated for each wash cycle and do not allow wells to sit uncovered or dry for extended periods.
- The use of radioimmunotherapy has significantly increased the number of patients with human anti-mouse IgG antibodies (HAMA). HAMA may interfere with assays utilizing murine monoclonal antibodies leading to both false positive and false negative results. Serum samples containing antibodies to murine immunoglobulins can still be analysed in such assays when murine immunoglobulins (serum, ascitic fluid, or monoclonal antibodies of irrelevant specificity) are added to the Sample.

13 Performance Characteristics

13.1 Sensitivity

The limit of detection of human sTNF-R (60kDa) defined as the analyte concentration resulting in an absorbance significantly higher than that of the dilution medium (mean plus 2 standard deviations) was determined to be 0.053 ng/ml (mean of 6 independent assays).

13.2 Reproducibility

13.2.1 Intra-assay

Reproducibility within the assay was evaluated in 3 independent experiments. Each assay was carried out with 6 replicates of 5 serum samples containing different concentrations of human sTNF-R (60kDa). 2 standard curves were run on each plate. Data below show the mean human sTNF-R (60kDa) concentration and the coefficient of variation for each sample (see Table 3). The calculated overall intra-assay coefficient of variation was 4.1%.

Table 3

The Mean human sTNF-R (60kDa) concentration and the coefficient of variation for each sample.

Positive Sample	Experiment	human sTNF-R (60kDa) Concentration (ng/ml)	Coefficient of Variation (%)
1	1	8.9	1.2
	2	8.7	2.1
	3	10.3	1.8
2	1	7.8	2.8
	2	7.7	2.5
	3	8.7	1.1
3	1	9.2	1.9
	2	8.4	1.9
	3	10.3	6.1
4	1	12.4	2.8
	2	10.8	11.1
	3	11.1	11.1
5	1	11.4	2.0
	2	10.1	8.6
	3	10.7	13.5

13.2.2 Inter-assay

Assay to assay reproducibility within one laboratory was evaluated in 3 independent experiments by 3 technicians. Each assay was carried out with 6 replicates of 5 serum samples containing different concentrations of human sTNF-R (60kDa). 2 standard curves were run on each plate. Data below (see Table 4) show the mean human sTNF-R (60kDa) concentration and the coefficient of variation calculated on 18 determinations of each sample. The calculated overall coefficient of variation was 8.2%.

Table 4
The mean human sTNF-R (60kDa) concentration and the coefficient of variation calculated on 18 determinations of each sample.

Sample	human sTNF-R (60kDa) Concentration (ng/ml)	Coefficient of Variation (%)
1	9.3	9.7
2	8.1	7.1
3	9.3	10.6
4	11.5	7.6
5	10.8	6.2

13.3 Spike Recovery

The spike recovery was evaluated by spiking 2 levels of sTNF-R (60 kDa) into pooled normal human serum diluted 1:10. Recoveries were determined in 3 independent experiments with 8 replicates each. The unspiked serum (diluted 1:10) was used as blank in these experiments. The overall mean recovery was determined to be 73%.

13.4 Dilution Parallelism

4 serum samples with different levels of human sTNF-R (60kDa) were analysed at serial 2 fold dilutions with 4 replicates each. The recovery ranged between 87% and 112% with an overall recovery of 102%.

13.5 Sample Stability

13.5.1 Freeze-Thaw Stability

Aliquots of serum samples (unspiked or spiked) were stored at -70°C and thawed 5 times, and the human sTNF-R (60kDa) levels determined. There was no significant loss of human sTNF-R (60 kDa) immunoreactivity by freezing and thawing.

13.5.2 Storage Stability

Aliquots of serum samples (spiked or unspiked) were stored at -20°C, 2-8°C, room temperature (RT) and at 37°C, and the human sTNF-R (60kDa) level determined after 24 h. There was no significant loss of human sTNF-R (60 kDa) immunoreactivity during storage under above conditions.

13.6 Comparison of Serum and Plasma

From two individuals, serum as well as EDTA plasma, citrate plasma, and heparin plasma obtained at the same time point were evaluated. Human sTNF-R (60 kDa) concentrations were not significantly different and therefore all these body fluids are suitable for the assay.

13.7 Specificity

The interference of tumor necrosis factor alpha (TNFα) and beta (TNFβ) with the assay was evaluated by measuring 50% calf serum with a sTNF-R (60 kDa) level of 5 ng/ml spiked with different concentrations of above cytokines in the sTNF-R (60kDa) ELISA. No interference has been observed with TNFα up to 10 ng/ml and with TNFβ up to 100 µg/ml.

13.8 Expected Values

A panel of 37 randomly selected sera from healthy blood donors (males and females) was tested for human sTNF-R (60kDa). The detected human sTNF-R (60kDa) levels ranged between 1.47 and 4.16 ng/ml with a mean level of 2.67 ng/ml and a standard deviation of ±0.69 ng/ml.

14 Ordering Information

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* Customers outside North America and Europe may contact their eBioscience distributor listed on our website at www.eBioscience.com/distributors.

15 Reagent Preparation Summary

15.1 Wash Buffer (1x)

Add **Wash Buffer Concentrate** 20x (25 ml) to 475 ml distilled water

15.2 Assay Buffer (1x)

Add **Assay Buffer Concentrate** 20x (5 ml) to 95 ml distilled water

16 Test Protocol Summary

- Place standard strips in position A1/A2 to H1/H2.
- Add distilled water, in duplicate, to all standard and blank wells as indicated on the label of the standard strips.
- Add 140 µl distilled water to sample wells.
- Add 10 µl Sample to designated wells.
- Cover microwell strips and incubate 2 hours at room temperature (18° to 25°C) on a microplate shaker at 400 rpm.
- Empty and wash microwell strips 3 times with 400 µl Wash Buffer.
- Add 100 µl of TMB Substrate Solution to all wells including blank wells.
- Incubate the microwell strips for about 10 minutes at room temperature (18° to 25°C).
- Add 100 µl Stop Solution to all wells including blank wells.
- Blank microwell reader and measure colour intensity at 450 nm.

Note: Samples have been diluted 1:10, thus the concentration read from the standard curve must be multiplied by the dilution factor (x 10).