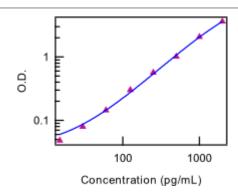


An Affymetrix Company

Mouse IL-21 ELISA Ready-SET-Go!®

Catalog Number: 88-8210 Also known as: Interleukin-21

RUO: For Research Use Only. Not for use in diagnostic procedures.



Standard curve of Mouse IL-21 ELISA Ready-SET-Go!®

Product Information

Contents: Mouse IL-21 ELISA Ready-SET-

Go!®

Catalog Number: 88-8210 Sensitivity: 16 pg/mL

Standard Curve Range: 16 - 2000 pg/mL



Temperature Limitation: Store at 2-8°C except standard which should be stored at less than or

equal to -70°C.



Batch Code: Refer to vial
Use By: Refer to vial



This mouse IL-21 ELISA Ready-SET-Go! reagent set (with or without high-affinity binding microwell plates) contains the necessary reagents, buffers and diluents for performing quantitative enzyme linked immunosorbent assays (ELISA). This ELISA reagent set is specifically engineered for accurate and precise measurement of mouse IL-21 protein levels from samples including serum, and supernatants from cell culture.

IL-21 is a 17 kDa immunomodulatory cytokine produced mainly by NKT, T helper (Th) 17 and T follicular helper (T_{FH}) cells. In T_{FH} cells, IL-21 expression leads to autocrine signaling through the IL-21 receptor (IL-21R) and STAT3, which leads to additional transcriptional activation by Bcl6. As with IFNγ for Th1 and IL-4 for Th2 cells, IL-21 is critical for T_{FH} cell development and effector function. This cytokine plays a role in T cell-dependent B cell differentiation into plasma cells and memory cells, stimulation of IgG production and induction of apoptotic signaling in naïve B cells.

In Th17 cells, IL-21 expression and autocrine feedback through STAT3, IRF4 and RORγt lead to upregulation of the IL-23R, thereby preparing Th17 cells for maturation and maintenance by the inflammatory cytokine IL-23. While upregulating IRF4 and RORγt, IL-21 also mediates the downregulation of Foxp3. High levels of IL-21 are present in chemically-induced colitis models. IL-21-deficient mice are protected from developing colitis upon chemical treatment by their inability to upregulate Th17-associated molecules.

Special Note: To ensure optimal results from this ELISA Ready-SET-Go! Set, please only use the components included in the set. Exchanging of components is not recommended as a change in signal may occur.

Components

Capture Antibody. Pre-titrated, purified antibody

Detection Antibody. Pre-titrated, biotin-conjugated antibody

Standard. Recombinant cytokine for generating standard curve and calibrating samples

Coating Buffer. 10X PBS ELISA Coating Buffer

Assay Diluent. 5X concentrated

Detection enzyme. Pre-titrated Avidin-HRP

Substrate Solution. Tetramethylbenzidine (TMB) Substrate Solution



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Certificate of Analysis. Lot-specific instructions for dilution of antibodies and standards 96 Well Plate. NUNC Maxisorp flat-bottom (included with product Cat. #'s ending in suffixes -22, -44, -76, -86)

Applications Reported

Mouse IL-21 ELISA Ready-SET-Go! has been reported for use in ELISAset.

References

Eddahri F, Denanglaire S, Bureau F, Spolski R, Leonard WJ, Leo O, Andris F. Interleukin-6/STAT3 signaling regulates the ability of naive T cells to acquire B-cell help capacities. Blood. 2009 Mar 12;113(11):2426-33.

Konforte D, Paige CJ. Interleukin-21 regulates expression of the immediate-early lytic cycle genes and proteins in Epstein-Barr Virus infected B cells. Virus Res. 2009 Sep;144(1-2):339-43.

Pot C, Jin H, Awasthi A, Liu SM, Lai CY, Madan R, Sharpe AH, Karp CL, Miaw SC, Ho IC, Kuchroo VK. Cutting edge: IL-27 induces the transcription factor c-Maf, cytokine IL-21, and the costimulatory receptor ICOS that coordinately act together to promote differentiation of IL-10-producing Tr1 cells. J Immunol. 2009 Jul 15;183(2):797-801.

Elsaesser H, Sauer K, Brooks DG. IL-21 is required to control chronic viral infection. Science. 2009 Jun 19;324(5934):1569-72.

Bauquet AT, Jin H, Paterson AM, Mitsdoerffer M, Ho IC, Sharpe AH, Kuchroo VK. The costimulatory molecule ICOS regulates the expression of c-Maf and IL-21 in the development of follicular T helper cells and TH-17 cells. Nat Immunol. 2009 Feb;10(2):167-75.

Nurieva R, Yang XO, Martinez G, Zhang Y, Panopoulos AD, Ma L, Schluns K, Tian Q, Watowich SS, Jetten AM, Dong C. Essential autocrine regulation by IL-21 in the generation of inflammatory T cells. Nature. 2007 Jul 26;448(7152):480-3.

Related Products

12-7211 Anti-Mouse IL-21 PE (FFA21)

14-8211 Mouse IL-21 Recombinant Protein

34-8211 Mouse IL-21 Recombinant Protein Carrier-Free

51-7213 Anti-Mouse IL-21 Alexa Fluor® 647 (To Be Discontinued. Refer to Cat. No. 50-7213) (mhalx21)

88-7216 Human IL-21 ELISA Ready-SET-Go!® (To Be Discontinued. Refer to Cat. No. 88-8218)



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Protocol: ELISA Ready-Set-Go!

The following protocol is a general guideline for the Ready-SET-Go! Sets

Materials Provided

Please refer to the Certificate of Analysis (C of A) for components

Other Materials Needed

- Buffers*
 - Wash Buffer: 1x PBS, 0.05% Tween-20 (or eBioscience ELISA Wash Buffer Powder, Cat. No. 00-0400)
 - Stop Solution: 1M H₃PO₄ (recommended) or 2N H₂SO₄
- Pipettes
- Refrigerator & frost-free -20°C freezer
- 96-well plate (Corning Costar 9018 or NUNC Maxisorp[®])

NOTE: The use of ELISA plates which are not high affinity protein binding plates will result in suboptimal performance, e.g., low signal or inconsistent data. Do not use tissue culture plates or low protein absorption plates. Use only the Corning Costar 9018 or NUNC Maxisorp (Cat. No. 44-2404) 96-well plates

- 96-well ELISA plate reader (microplate spectrophotometer)
- ELISA plate washer (highly recommended)

NOTE: To ensure optimal results from this ELISA Ready-SET-Go! Set, please only use the components included in the set. Exchanging of components is not recommended as a change in signal may occur.

Time Requirements

- 1 overnight incubation
- 4½-hour incubations
- 1 hour washing and analyzing samples

Experimental Procedure

- 1. Coat Corning Costar 9018 (or Nunc Maxisorp®) ELISA plate with 100 μ L/well of capture antibody in Coating Buffer (dilute as noted on C of A, which is included with the reagent set). Seal the plate and incubate overnight at 4°C.
- 2. Aspirate wells and wash 3 times with >250 μ L/well Wash Buffer*. Allowing time for soaking (~ 1 minute) during each wash step increases the effectiveness of the washes. Blot plate on absorbent paper to remove any residual buffer.
- 3. Dilute 1 part 5X concentrated Assay Diluent with 4 parts DI water.* Block wells with 200 µL/well of 1X Assay Diluent. Incubate at room temperature for 1 hour.
- 4. Optional: Aspirate and wash at least once with Wash Buffer.
- Using 1X Assay Diluent*, dilute standards as noted on the C of A to prepare the top concentration of the standard. Add 100 µL/well of top standard concentration to the appropriate



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wells. Perform 2-fold serial dilutions of the top standards to make the standard curve for a total of 8 points. Add 100 μ L/well of your samples to the appropriate wells. Seal the plate and incubate at room temperature for 2 hours (or overnight at 4°C for maximal sensitivity).

- 6. Aspirate/wash as in step 2. Repeat for a total of 3-5 washes**.
- Add 100 μL/well of detection antibody diluted in 1X Assay Diluent* (dilute as noted on C of A).
 Seal the plate and incubate at room temperature for 1 hour.
- 8. Aspirate/wash as in step 2. Repeat for a total of 3-5 washes**.
- 9. Add 100 μL/well of Avidin-HRP* diluted in 1X Assay Diluent (dilute as noted on C of A). Seal the plate and incubate at room temperature for 30 minutes.
- 10. Aspirate and wash as in step 2. In this wash step, soak wells in Wash Buffer* for 1 to 2 minutes prior to aspiration. Repeat for a total of 5-7 washes**.
- 11. Add 100 μ L/well of Substrate Solution to each well. Incubate plate at room temperature for 15 minutes.
- 12. Add 50 µL of Stop Solution to each well.
- 13. Read plate at 450 nm. If wavelength subtraction is available, subtract the values of 570 nm from those of 450 nm and analyze data.

NOTES:

- * Be certain that no sodium azide is present in the solutions used in this assay, as this inhibits HRP enzyme activity.
- **The number of washes in the protocol was adapted to an automatic plate washer. This can be decreased when using other methods but should be tested empirically. Allowing time for soaking (~ 1 minute) during each wash step increases the effectiveness of the washes.



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Quick Guide: Standard Calibration

The following table indicates the protein standard contained in the Ready-SET-Go! is calibrated against NIBSC standards.

Table of Standard Calibration					
Cytokine	ng of eB standard	ng of NIBSC standard	U of NIBSC standard	NIBSC Lot #	
hIL-2	1	1.1	14.6	86/564	
hIL-4	1	2.2	22	88/656	
hIL-5	1	2.2	22	90/586	
hIL-6	1	1.7	170	89/548	
hIL-8	1	1.8	180	89/520	
hIL-10	1	0.8	4	93/722	
hIL-12	1	0.8	8	95/544	
hIL-17A	1	0.9	9000	01/420	
hIFN-g	1	1.1	22	87/586	
hTNF-a	1	0.9	36	87/650	
mIL-2	1	3.1	310	93/566	
mIL-4	1	3	30	91/656	
mIL-6	1	8.5	850	93/730	
mIFN-g*	1		4.5	Gg02-901-533	
mTNF-a	1	1.7	340	88/532	
* Mouse IFN-g is calibrated using NIH standard (Lot Gg02-901-533) and is measured in Units (U)					

ELISA Troubleshooting Guide				
Problem	Possibility	Solution		
A. High background	Improper and inefficient washing	Improve efficiency of washing. Fill plates completely, soak for 1 minute per wash, as directed		
	Cross contamination from other specimens or positive control	Repeat ELISA being careful when washing and pipetting		
	3. Contaminated substrate	3. Substrate should be colorless. Replace		
	4. Incorrect dilutions, e.g., conjugate concentration was too high	4. Repeat using correct dilutions		
B. No signal	Improper, low protein binding capacity plates were used	Repeat ELISA using recommended high binding capacity plates		
	2. Wrong substrate was used	2. Repeat ELISA using the correct substrate		
	3. Enzyme inhibitor present in buffers; e.g., sodium azide in the washing buffer and Assay Diluent inhibits peroxidase activity	3. Repeat ELISA making no enzyme inhibitor is present in any buffers.		



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	4. Coated capture antibody in Assay Diluent rather than Coating Buffer	4. Repeat ELISA using Coating Buffer contained in the set as the diluent for the capture antibody.
C. Very weak signal	Improper and inefficient washing	1. Make sure washing procedure is done correctly, with a soak time.
	2. Incorrect dilutions of standard	2. Follow recommendations of standard preparation exactly as written on the C of A
	3. Insufficient incubation time	3. Repeat ELISA following the protocol carefully for each step
	4. Incorrect storage of reagents	4. Store reagents at the correct temperature asn indicated on the Technical Data Sheet. Freezing certain components will severely impact results. Do not re-use the standards.
	5. Wrong filter in ELISA reader was used	5. Use the correct wavelength setting
	6. Wrong plate used	6. Use the recommended Corning Costar 9018 or NUNC Maxisorp flat bottom 96 well plates
D. Variation amongst replicates	Improper and inefficient washing	1. Make sure washing procedure is done correctly; see C of A. Edge effects can be avoided by moving samples and standards in from the edge of the plate.
	2. Poor mixing of samples	2. Mix samples and reagents gently and equilibrate to proper temperature
	3. Plates not clean	3. Plates should be wiped on bottom before measuring absorbance
	4. Reagents have expired	4. Order a new Ready-Set-Go ELISA.