

# Human IFN gamma ELISPOT Ready-SET-Go!®

Catalog Number: 88-7386 Also Known As:Interferon-gamma, IFN-g, IFNg RUO: For Research Use Only. Not for use in diagnostic procedures.



Normal human peripheral blood cells were activated with PMA/Ionomycin in human IFN gamma ELISPOT assay. Control well is medium alone.

## **Product Information**

Contents: Human IFN gamma ELISPOT Ready-SET-Gol® REF Catalog Number: 88-7386

**Temperature Limitation:** Store at 2-8°C.

LOT Batch Code: Refer to Vial

Use By: Refer to Vial

▲ Caution, contains Azide

## Description

This Human IFN-γ ELISPOT Ready-SET-Go! reagent set contains the necessary reagents for performing enzyme linked immunosorbent spot (ELISPOT) assays for high resolution frequency analysis of IFN-γ-secreting cells. This ELISPOT reagent set is pre-titrated for optimal spot development.

#### Components

Capture Antibody. Pre-titrated, Functional Grade (low endotoxin) purified antibody

### Detection Antibody. Pre-titrated, biotin-conjugated antibody

**ELISA/ELISPOT Coating Buffer**This Ready-Set-Go! ELISPOT Set may contain ELISA/ELISPOT Coating Buffer Powder (Reconstitute to 1L with dH20 and filter (0.22 uM)) or 10X PBS ELISPOT Coating Buffer (Dilute 1 part 10X Buffer into 9 parts dH20 and filter with 0.22 uM).

Assay Diluent. 5X concentrated

Detection enzyme. Pre-titrated Avidin-HRP

Certificate of Analysis. Lot-specific instructions for dilution of antibodies and enzyme

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## **ELISPOT Set Protocol**

### Research Use Only Additional Materials Needed

- 96-Well PVDF Membrane ELISPOT Plates (Millipore, Cat. No. MAIPS4510)
- AEC (3-amino-9-ethyl carbazole) Substrate (Sigma, Cat. No. A-5754)
- Distilled water
- ELISPOT Wash Buffer: 1X PBS, with 0.05% Tween-20 (or eBioscience ELISA Wash Buffer Powder, cat 00-0400)

#### Instruments

- Pipettes and pipettors
- Refrigerator
- Incubator
- Laminar Flow Hood
- Plate Washer: Wash bottle or automated wash machine
- ELISPOT plate reader or dissecting microscope for visual inspection

#### **Experiment Duration**

- 1 overnight antibody incubation
- 1-2 day cell activation
- 3-5 hr washing, antibody incubation, color development

### **ELISPOT Method**

Aseptic Procedure: (Use sterile buffers and aseptic conditions; use laminar flow hood for procedures.)

- Dilute Functional Grade purified capture antibody in sterile ELISPOT Coating Buffer, as noted on Certificate of Analysis which is included with the reagent set. Coat ELISPOT plate with 100 μl/well of capture antibody solution. Incubate at 4°C overnight.
- 2. Decant or aspirate coating antibody from plate.
- 3. Wash plates 2 times with 200 µl/well sterile ELISPOT Coating Buffer. Decant.
- 4. Block plate with 200 µl/well of complete RPMI-1640 at room temperature for 1 hour. Decant or aspirate plate.
- Aliquot mitogen, antigen or controls diluted in complete RPMI-1640 medium to appropriate wells at 100 μl/well. Aliquot cells at desired densities (e.g., 1x10<sup>5</sup>/ml - 2x10<sup>6</sup>/ml) at 100 μl/well and incubate at 37°C, 5% CO<sub>2</sub> humidified incubator for 24-48 hours.

**Note**: Kinetics and cell densities vary with target cytokine, treatment, and cell type and must be empirically determined. See references. Cells can be diluted in a sterile tissue culture plate starting at  $2x10^6$ /well in triplicate wells with a series of 1:3 or 1:4 serial dilutions down the plate, and then transferred to the ELISPOT plate.



## **ELISPOT Set Protocol**

### **Research Use Only**

Non-Aseptic Procedure:

- 1. Decant cells and medium from plates. Wash plate 3 times with ELISPOT Wash Buffer.
- Dilute biotinylated detection antibody in Assay Diluent according to instructions on the Certificate of Analysis provided with the reagent set. Add 100 μl/well to plate microwells and incubate at room temperature for 2 hr (or 4°C overnight).
- 3. Decant antibody solution. Wash 4 times with ELISPOT Wash Buffer. Allow wells to soak for 1 minute for each wash.
- 4. Dilute Avidin-HRP reagent in Assay Diluent according to instructions on the Certificate of Analysis provided with the reagent set. Add 100 μl/well of Avidin-HRP and incubate at room temperature for 45 minutes.
- Decant Avidin-HRP solution. Wash plate 3 times with ELISPOT Wash Buffer, and then 2 times with 1X PBS (no Tween-20).
- Add 100 μl/well of freshly-prepared AEC Substrate Solution and develop at room temperature for 10-60 minutes; monitor development of spots.
- 7. Stop the substrate reaction by washing wells 3 times with 200 µl/well distilled water.
- 8. Air-dry the plate. Count spots using a dissecting microscope or automated ELISPOT plate reader. Store plates in the dark prior to reading.

## **Cytokine ELISPOT Buffers**

- ELISA/ELISPOT Coating Buffer Powder
  - Reconstitute powder to 1 L in dH<sub>2</sub>0; filter sterilize using a 0.22 μM filter
- Complete RPMI-1640:
  - RPMI-1640 with 10% FBS and 1% Pen/Strep/L-Glu
- Assay Diluent (supplied as 5X):
  - Dilute 5X solution to 1X in DI H<sub>2</sub>O
- ELISPOT Wash Buffer:
  - 1X PBS with 0.05% Tween-20 (0.5 ml Tween-20 in 1 L PBS) or eBioscience ELISA Wash Buffer Powder (Cat # 00-0400)
- 1X PBS:
  - 80.0 g NaCl
  - 11.6 g Na<sub>2</sub>HPO<sub>2</sub>
  - 2.0 g KH<sub>2</sub>PO<sub>4</sub>
  - 2.0 g KCl
  - Qs with DI H<sub>2</sub>0 up to 10.0 L, pH to 7.0



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- AEC (3-amino-9-ethyl carbazole) Substrate Solution:
  - AEC Stock Solution: Dissolve 100 mg of AEC in 10 ml of N,N Dimethylformamide (DMF; Pierce, Cat. No. 20672)
  - Add 333 μl of AEC Stock Solution to 10 ml of 0.1 M Acetate Solution (pH 5.0) (see below for recipe). Filter through a 0.45 μm filter.
  - Just before use, add 5  $\mu$ l of 30% H<sub>2</sub>O<sub>2</sub>. Mix and use immediately.
- 0.1 M Acetate Solution (pH 5.0):
  - Combine 148 ml 0.2 M acetic acid (11.55 ml glacial acetic acid per 1 L dH<sub>2</sub>O) with 352 ml 0.2 M sodium acetate (27.2 g sodium acetate per 1 L dH<sub>2</sub>O).
  - Qs to 1 L with dH<sub>2</sub>O. Adjust pH to 5.0.

## **Selected References**

- 1. Gebauer BS, et al. 2002. Evolution of the enzyme-linked immunosorbent spot assay for post-transplant alloreactivity as a potentially useful immune monitoring tool. Am. J. Transplant. 9: 857-866.
- Guerkov RE, et al. 2003. Detection of low-frequency antigen-specifi c IL-10-producing CD4(+) T cells via ELISPOT in PBMC: cognate vs. nonspecifi c production of the cytokine. J. Immunol. Methods. 279: 111-121.
- 3. Kreher CR, et al. 2003. CD4+ and CD8+ cells in cryopreserved human PBMC maintain full functionality in cytokine ELISPOT assays. J. Immunol. Methods. 278: 79-93.
- 4. Ott PA, et al. 2004. CD28 costimulation enhances the sensitivity of the ELISPOT assay for detection of antigen-specific memory effector CD4 and CD8 cell populations in human diseases. J. Immunol. Methods. 285: 223-235.
- 5. Smith JG, et al. 2001. Development and validation of a gamma interferon ELISPOT assay for quantitation of cellular immune responses to varicella-zoster virus. Clin. Diag. Lab. Immunol. 8: 871-879.
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- 7. Rininsland F, et al. 2000. Granzyme B ELISPOT assay for ex vivo measurements of T cell immunity. J. Immunol. Meth. 240:143-155.