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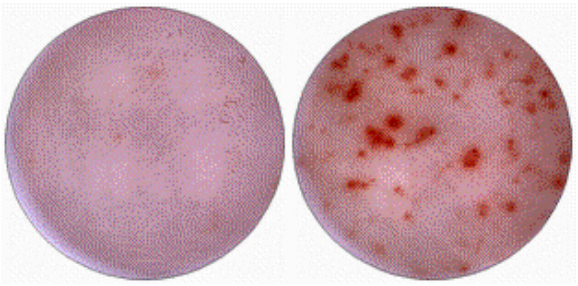
Mouse/Rat IL-17A ELISPOT Ready-SET-Go!®

Catalog Number: 88-7370
Also Known As: Interleukin-17A, IL17A, IL-17AA
RUO: For Research Use Only. Not for use in diagnostic procedures.



Splenocytes were activated with PMA and Ionomycin for 24 hrs in a Mouse IL-17A ELISPOT assay (right). Left well is medium alone control (no mitogen).

Product Information

Contents: Mouse/Rat IL-17A ELISPOT Ready-SET-Go!®
REF **Catalog Number:** 88-7370

Temperature Limitation: Store at 2-8°C.
LOT **Batch Code:** Refer to Vial
Use By: Refer to Vial
Caution, contains Azide

Description

This Mouse IL-17A ELISPOT Ready-SET-Go! reagent set contains the necessary reagents for performing enzyme linked immunosorbent spot (ELISPOT) assays for high resolution frequency analysis of IL-17A-secreting cells. This ELISPOT reagent set is pre-titrated for optimal spot development.

Components

Capture Antibody. Pre-titrated, Functional Grade (low endotoxin) purified antibody
Detection Antibody. Pre-titrated, biotin-conjugated antibody
ELISA/ELISPOT Coating Buffer Powder. This Ready-Set-Go! ELISPOT Set may contain ELISA/ELISPOT Coating Buffer Powder (Reconstitute to 1L with dH20 and filter (0.22 uM)) or 10X PBS ELISPOT Coating Buffer (Dilute 1 part 10X Buffer into 9 parts dH20 and filter with 0.22 uM).
Assay Diluent. 5X concentrated
Detection enzyme. Pre-titrated Avidin-HRP
Certificate of Analysis. Lot-specific instructions for dilution of antibodies and enzyme

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ELISPOT Set Protocol

Research Use Only

Additional Materials Needed

- 96-Well PVDF Membrane ELISPOT Plates (Millipore, Cat. No. MAIPS4510)
- AEC (3-amino-9-ethyl carbazole) Substrate (Sigma, Cat. No. A-5754)
- Distilled water
- ELISPOT Wash Buffer: 1X PBS, with 0.05% Tween-20 (or eBioscience ELISA Wash Buffer Powder, cat 00-0400)

Instruments

- Pipettes and pipettors
- Refrigerator
- Incubator
- Laminar Flow Hood
- Plate Washer: Wash bottle or automated wash machine
- ELISPOT plate reader or dissecting microscope for visual inspection

Experiment Duration

- 1 overnight antibody incubation
- 1-2 day cell activation
- 3-5 hr washing, antibody incubation, color development

ELISPOT Method

Aseptic Procedure: (Use sterile buffers and aseptic conditions; use laminar flow hood for procedures.)

1. Dilute Functional Grade purified capture antibody in sterile ELISPOT Coating Buffer, as noted on Certificate of Analysis which is included with the reagent set. Coat ELISPOT plate with 100 µl/well of capture antibody solution. Incubate at 4°C overnight.
2. Decant or aspirate coating antibody from plate.
3. Wash plates 2 times with 200 µl/well sterile ELISPOT Coating Buffer. Decant.
4. Block plate with 200 µl/well of complete RPMI-1640 at room temperature for 1 hour. Decant or aspirate plate.
5. Aliquot mitogen, antigen or controls diluted in complete RPMI-1640 medium to appropriate wells at 100 µl/well. Aliquot cells at desired densities (e.g., $1x10^5$/ml - $2x10^6$/ml) at 100 µl/well and incubate at 37°C, 5% CO_2 humidified incubator for 24-48 hours.
   ***Note**: Kinetics and cell densities vary with target cytokine, treatment, and cell type and must be empirically determined. See references. Cells can be diluted in a sterile tissue culture plate starting at $2x10^6$/well in triplicate wells with a series of 1:3 or 1:4 serial dilutions down the plate, and then transferred to the ELISPOT plate.*

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Non-Aseptic Procedure:

1. Decant cells and medium from plates. Wash plate 3 times with ELISPOT Wash Buffer.
2. Dilute biotinylated detection antibody in Assay Diluent according to instructions on the Certificate of Analysis provided with the reagent set. Add 100 µl/well to plate microwells and incubate at room temperature for 2 hr (or 4°C overnight).
3. Decant antibody solution. Wash 4 times with ELISPOT Wash Buffer. Allow wells to soak for 1 minute for each wash.
4. Dilute Avidin-HRP reagent in Assay Diluent according to instructions on the Certificate of Analysis provided with the reagent set. Add 100 µl/well of Avidin-HRP and incubate at room temperature for 45 minutes.
5. Decant Avidin-HRP solution. Wash plate 3 times with ELISPOT Wash Buffer, and then 2 times with 1X PBS (no Tween-20).
6. Add 100 µl/well of freshly-prepared AEC Substrate Solution and develop at room temperature for 10-60 minutes; monitor development of spots.
7. Stop the substrate reaction by washing wells 3 times with 200 µl/well distilled water.
8. Air-dry the plate. Count spots using a dissecting microscope or automated ELISPOT plate reader. Store plates in the dark prior to reading.

Cytokine ELISPOT Buffers

- ELISA/ELISPOT Coating Buffer Powder
  - Reconstitute powder to 1 L in dH_20; filter sterilize using a 0.22 µM filter
- Complete RPMI-1640:
  - RPMI-1640 with 10% FBS and 1% Pen/Strep/L-Glu
- Assay Diluent (supplied as 5X):
  - Dilute 5X solution to 1X in DI H_2O
- ELISPOT Wash Buffer:
  - 1X PBS with 0.05% Tween-20 (0.5 ml Tween-20 in 1 L PBS) or eBioscience ELISA Wash Buffer Powder (Cat # 00-0400)
- 1X PBS:
  - 80.0 g NaCl
  - 11.6 g Na_2HPO_2
  - 2.0 g KH_2PO_4
  - 2.0 g KCl
  - Qs with DI H_20 up to 10.0 L, pH to 7.0

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- AEC (3-amino-9-ethyl carbazole) Substrate Solution:
  - AEC Stock Solution: Dissolve 100 mg of AEC in 10 ml of N,N Dimethylformamide (DMF; Pierce, Cat. No. 20672)
  - Add 333 µl of AEC Stock Solution to 10 ml of 0.1 M Acetate Solution (pH 5.0) (see below for recipe). Filter through a 0.45 µm filter.
  - Just before use, add 5 µl of 30% H_2O_2. Mix and use immediately.

- 0.1 M Acetate Solution (pH 5.0):
  - Combine 148 ml 0.2 M acetic acid (11.55 ml glacial acetic acid per 1 L dH_2O) with 352 ml 0.2 M sodium acetate (27.2 g sodium acetate per 1 L dH_2O).
  - Qs to 1 L with dH_2O. Adjust pH to 5.0.

Selected References

1. Gebauer BS, et al. 2002. Evolution of the enzyme-linked immunosorbent spot assay for post-transplant alloreactivity as a potentially useful immune monitoring tool. Am. J. Transplant. 9: 857-866.
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4. Ott PA, et al. 2004. CD28 costimulation enhances the sensitivity of the ELISPOT assay for detection of antigen-specifi c memory effector CD4 and CD8 cell populations in human diseases. J. Immunol. Methods. 285: 223-235.
5. Smith JG, et al. 2001. Development and validation of a gamma interferon ELISPOT assay for quantitation of cellular immune responses to varicella-zoster virus. Clin. Diag. Lab. Immunol. 8: 871-879.
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7. Rininsland F, et al. 2000. Granzyme B ELISPOT assay for ex vivo measurements of T cell immunity. J. Immunol. Meth. 240:143-155.

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