

# Mouse IL-17E (IL-25) ELISA Ready-SET-Go!®

Catalog Number: 88-7002 Also known as: Interleukin-17E, Interleukin-25 RUO: For Research Use Only. Not for use in diagnostic procedures.



### Description

This Mouse IL-17E ELISA Ready-SET-Go! reagent set contains all of the necessary buffers and reagents to perform enzyme-linked immunosorbent assays. The set has been optimized for the accurate and precise measurement of mouse IL-17E in serum and tissue culture supernatant samples. Serum samples must be diluted 8-fold in the provided sample diluent prior to evaluation in this assay.

IL-17E, or IL-25, was first identified as an IL-17 family member on the basis of sequence homology. It was originally believed to be a Th2 cytokine based on the detection of mRNA transcripts in polarized Th2 cells, but expression was later observed in epithelial cells, eosinophils, and mast cells. Later studies confirmed the role of IL-17E as a potent inducer of type 2 immune responses. IL-17E signaling induces the differentiation of CD4+ T cells towards the Th2 lineage and upregulates expression of Th2 cytokines such as IL-4, IL-5, and IL-13. Administration of IL-17E also promotes symptoms of allergic asthma, such as eosinophil infiltration, airway hyperreactivity, and high circulating levels of IgE. IL-17E binds to IL-17BR, a receptor shared with IL-17B, and activates NF kappa-B pathways.

### Components

Capture Antibody. Pre-titrated, purified antibody Detection Antibody. Pre-titrated, biotin-conjugated antibody Standard. Recombinant cytokine for generating standard curve and calibrating samples Coating Buffer. 10X PBS ELISA Coating Buffer Assay Diluent. 5X Concentrated Sample Diluent B. 12 mL of a 1X solution per plate Detection Enzyme. Pre-titrated Avidin-HRP Substrate Solution. Tetramethylbenzidine (TMB) Solution Certificate of Analysis. Lot-specific instructions for dilution of antibodies and standards. 96-well Plates. Corning Costar flat-bottom plates (included with Cat. #s ending in -22 and -86)

### **Applications Reported**

This ELISA set is for the quantitative detection of mouse IL-17E in serum, plasma, and tissue culture supernatant

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samples.

#### **Applications Tested**

This assay has been validated for the detection of endogenous mouse IL-17E with supernatant collected from PMA and lonomycin-stimulated culture of Balb/c bone marrow-derived mast cells. No detectable levels were observed in unstimulated cultures.

This assay was evaluated for specificity on a panel of 72 recombinant cytokines at 100 ng/mL. No significant crossreactivity was observed, including to Human IL-17E Recombinant Protein, or to other recombinant proteins of the IL-17 family.

It is recommended that serum samples be diluted at least 8-fold with the included Sample Diluent B in order to ensure optimal recovery. Sample Diluent B is also compatible with tissue culture supernatant samples, although it is not necessary to use this buffer for their evaluation. Plasma samples have not been tested in this assay.

#### References

Dong C. Regulation and pro-inflammatory function of interleukin-17 family cytokines. Immunol Rev. 2008 Dec;226:80-6.

Kang CM, Jang AS, Ahn MH, Shin JA, Kim JH, Choi YS, Rhim TY, Park CS. Interleukin-25 and interleukin-13 production by alveolar macrophages in response to particles. Am J Respir Cell Mol Biol. 2005 Sep;33(3):290-6.

Ikeda K, Nakajima H, Suzuki K, Kagami S, Hirose K, Suto A, Saito Y, Iwamoto I. Mast cells produce interleukin-25 upon Fc epsilon RI-mediated activation. Blood. 2003 May 1;101(9):3594-6.

Hurst SD, Muchamuel T, Gorman DM, Gilbert JM, Clifford T, Kwan S, Menon S, Seymour B, Jackson C, Kung TT, Brieland JK, Zurawski SM, Chapman RW, Zurawski G, Coffman RL. New IL-17 family members promote Th1 or Th2 responses in the lung: in vivo function of the novel cytokine IL-25. J Immunol. 2002 Jul 1;169(1):443-53.

### **Related Products**

14-8175 Mouse IL-17E (IL-25) Recombinant Protein



### Research Use Only

**Other Materials Needed** 

- Buffers
  - Wash Buffer: 1 x PBS, 0.05% Tween-20 (or eBioscience ELISA Wash Buffer Powder, cat 00-0400)
  - Stop Solution: 1M H<sub>3</sub>PO<sub>4</sub> or 2N H<sub>2</sub>SO<sub>4</sub>
- Pipettes and pipettors
- Page Refrigerator
- 96-well plate (Corning Costar 9018)
  - NOTE: The use of ELISA plates which are not high affinity protein binding plates will result in suboptimal performance, e.g., low signal or inconsistent data. Do not use tissue culture plates or low protein absorption plates. Use only the Corning Costar 9018 or NUNC Maxisorp 96 well plates provided or suggested.
- 96-well ELISA plate reader (microplate spectrophotometer)
- ELISA plate washer

NOTE: To ensure optimal results from this ELISA Ready-SET-Go! set, please only use the components included in the set. Exchanging of components is not recommended as a change in signal may occur.

### Stability

This ELISA set is guaranteed to perform as specified at least 12 months from date of receipt if stored and handled as instructed according to this datasheet and the Certificate of Analysis, which is included with the reagents.

### **Storage Instructions for Cytokine Standards**

The frozen cytokine standard is already aliquoted at 20  $\mu$ l per vial. Upon receipt, frozen cytokine standard should be immediately stored at -80°C; stable for at least 12 months. After thawing, quick-spin vial prior to opening. Do not re-aliquot into smaller fractions. These are single use vials. Use one time and discard. For dilution of the standard, please see instructions on the Certificate of Analysis and follow these as written.

**Storage Instructions for Other Set Reagents** 

Store at 4°C.

**Time Requirements** 

- 1 overnight incubation
- 4½-hour incubations
- 1 hour washing and analyzing samples



Research Use Only Experimental Procedure

- Coat Corning Costar 9018 ELISA plate with 100 μl/well of capture antibody in Coating Buffer (dilute as noted on Certificate of Analysis, which is included with the reagent set). Seal the plate and incubate overnight at 4°C.
- 2. Aspirate wells and wash 5 times with >250 μl/well Wash Buffer\*. Allowing time for soaking (~ 1 minute) during each wash step increases the effectiveness of the washes. Blot plate on absorbent paper to remove any residual buffer.
- Dilute 1 part 5X concentrated Assay Diluent with 4 parts DI water.\* Block wells with 200 μl/well of 1X Assay Diluent. Incubate at room temperature for 1 hour.
- 4. Aspirate/wash as in step 2. Repeat for a total of 5 washes.
- 5. Using Sample Diluent B, dilute standards as noted on the Certificate of Analysis (C of A). Add 100 μl/well of standard to the appropriate wells. Perform 2-fold serial dilutions of the top standards to make the standard curve. Add 100 μl/well of your samples to the appropriate wells. Serum and plasma samples must be diluted at least 8-fold in Sample Diluent B prior to evaluation in this assay. Cover or seal the plate and incubate at room temperature for 2 hours (or overnight at 4°C for maximal sensitivity).
- 6. Aspirate/wash as in step 2. Repeat for a total of 5 washes.
- Add 100 μl/well of detection antibody diluted in 1X Assay Diluent\* (dilute as noted on C of A). Seal the plate and incubate at room temperature for 1 hour.
- 8. Aspirate/wash as in step 2. Repeat for a total of 5 washes.
- 9. Add 100 μl/well of Avidin-HRP\* diluted in 1X Assay Diluent (dilute as noted on C of A). Seal the plate and incubate at room temperature for 30 minutes.
- 10. Aspirate and wash as in step 2. In this wash step, soak wells in Wash Buffer\* for 1 to 2 minutes prior to aspiration. Repeat for a total of 7 washes.
- 11. Add 100 μl/well of Substrate Solution to each well. Incubate plate at room temperature for 15 minutes.
- 12. Add 50  $\mu$ l of Stop Solution to each well.
- 13. Read plate at 450 nm. If wavelength subtraction is available, subtract the values of 570 nm from those of 450 nm and analyze data.

\*NOTE: Be certain that no sodium azide is present in the solutions used in this assay, as this inhibits HRP enzyme activity.



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**Standard Calibration** 

The standard of the Ready-SET-Go! is calibrated against NIBSC standards:

Table of Standard Calibration					
Cytokine	ng of eB standard	ng of NIBSC standard	U of NIBSC standard	NIBSC Lot #	
hIL-2	1	1.1	14.6	86/564	
hIL-4	1	2.2	22	88/656	
hIL-5	1	2.2	22	90/586	
hIL-6	1	1.7	170	89/548	
hIL-10	1	0.8	4	93/722	
hIL-12	1	0.8	8	95/544	
hIFN-g	1	1.1	22	87/586	
hTNF-a	1	0.9	36	87/650	
mIL-2	1	3.1	310	93/566	
mIL-4	1	3	30	91/656	
mIL-6	1	8.5	850	93/730	
mIFN-g*	1		4.5	Gg02-901-533	
mTNF-a	1	1.7	340	88/532	
* Mouse IEN g is calibrated using NIH standard (Let $C_{0}$ 001 E22) and is measured in Units (U)					

\* Mouse IFN-g is calibrated using NIH standard (Lot Gg02-901-533) and is measured in Units (U)

ELISA Troubleshooting Guide				
Problem	Possibility	Solution		
A. High Background	1. Improper and inefficient washing	1. Improve efficiency of washing. Fill plates completely, soak for 1 minute per wash, as directed		
	2. Cross contamination from other specimens or positive control	2. Repeat ELISA, be careful when washing and pipetting		
	3. Contaminated substrate	3. Substrate should be colorless		
	4. Incorrect dilutions, e.g., conjugate concentration was too high	4. Repeat test using correct dilutions; check with the recommendations of the antibody manufacturer		
B. No signal	1. Improper, low protein binding capacity plates were used	1. Repeat ELISA, using recommended high binding capacity plates		
	2. Wrong substrate was used	2. Repeat ELISA, use the correct substrate		
	3. Enzyme inhibitor present in buffers; e.g., sodium azide in the washing buffer and Assay Diluent inhibits peroxidase activity	3. Repeat ELISA, make sure your system contains no enzyme inhibitor		





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C. Very weak signal	1. Improper and inefficient washing	1. Make sure washing procedure is done correctly
	2. Incorrect dilutions of standard	2. Follow recommendations of standard handling exactly as written on the certificate of analysis
	3. Insufficient incubation time	3. Repeat ELISA, follow the protocol carefully for each step' s incubation time
	4. Incorrect storage of reagents	4. Store reagents in the correct temperature, avoid freeze and thaw, avoid using the "frost free" freezer
	5. Wrong filter in ELISA reader was used	5. Use the correct wavelength setting
	6. Wrong plate used	6. Use the recommended Corning Costar 9018 or NUNC Maxisorp flat bottom 96 well plates
D. Variation amongst replicates	1. Improper and inefficient washing	1. Make sure washing procedure is done correctly; see certificate of analysis
	2. Poor mixing of samples	2. Mix samples and reagents gently and equilibrate to proper temperature
	3. Plates not clean	3. Plates should be wiped on bottom before measuring absorbance
	4. Improper, low binding capacity plates were used	4. Use recommended high binding capacity plates
	5. Reagents have expired	5. Do not use if past expiration date