

500 units 20,000 U/ml Lot: 0011207 RECOMBINANT Store at -20°C Exp: 7/14

Recognition Site:

5′... G C A T G C ... 3′ 3′... C,G T A C G ... 5′

Note: SphI-HF[™] has the same specificitiv as SphI (NEB #R0182), but it has been engineered for reduced star activity.



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Source: An E. coli strain that carries the cloned and modified (K100A) Sphl gene from Streptomyces phaeochromogenes (NRRL B-3559)

Supplied in: 200 mM NaCl. 10 mM Tris-HCl (pH 7.4), 0.1 mM EDTA, 1 mM DTT, 100 µg/ml BSA and 50% glycerol.

Reagents Supplied with Enzyme: 10X NEBuffer 4.

Reaction Conditions: 1X NEBuffer 4. Incubate at 37°C.

1X NEBuffer 4:

50 mM potassium acetate 20 mM Tris-acetate 10 mM magnesium acetate 1 mM DTT pH 7.9 @ 25°C

Unit Definition: One unit is defined as the amount of enzyme required to digest 1 μ g of λ DNA in 1 hour at 37°C in a total reaction volume of 50 µl.

Diluent Compatibility: Diluent Buffer B 300 mM NaCl. 10 mM Tris-HCl. 0.1 mM EDTA. 1 mM DTT. 500 µg/ml BSA and 50% glycerol. (pH 7.4 @ 25°C).

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Quality Controls

Ligation: After 20-fold overdigestion with SphI-HF. > 95% of the DNA fragments can be ligated with T4 DNA Ligase (at a 5' termini concentration of 1–2 μ M) at 16°C. Of these ligated fragments, > 95% can be recut.

16-Hour Incubation: A 50 µl reaction containing 1 µg of DNA and 25 units of enzyme incubated for 16 hours resulted in the same pattern of DNA bands as a reaction incubated for 1 hour with 1 unit of enzyme.

Exonuclease Activity: Incubation of 100 units of enzyme with 1 µg sonicated ³H DNA (10⁵ cpm/µg) for 4 hours at 37°C in 50 µl reaction buffer released < 0.1% radioactivity

Endonuclease Activity: Incubation of 100 units of enzyme with 1 ug ϕ X174 RF I DNA for 4 hours at 37°C in 50 µl reaction buffer resulted in < 20% conversion to RF II.

Blue/White Screening Assay: This enzyme has been tested to determine the integrity of the DNA ends produced after digestion with an excess of enzyme. An appropriate vector is digested at a unique site within $lacZ^{\alpha}$ gene with a 10-fold excess of enzyme, ligated, transformed and plated on XGal/IPTG/Amp plates. Successful expression of B-galactosidase is a function of how intact its gene remains after cloning, an intact gene gives rise to a blue colony, an interrupted gene (i.e. degraded DNA end) gives rise to a white colony. Enzymes must produce fewer than 3% white colonies to be Blue/White Certified.

Enzyme Properties

Activity in NEBuffers:	
NEBuffer 1	50%
NEBuffer 2	25%
NEBuffer 3	10%
NEBuffer 4	100 %

When using a buffer other than the optimal (supplied) NEBuffer, it may be necessary to add more enzyme to achieve complete digestion.

(see other side)

CERTIFICATE OF ANALYSIS

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(see other side)

info@neb.com www.neb.com 🗙 RX e 🥝 ★- NEB4 37° ₩₩ @

Survival in a Reaction: A minimum of 1.0 units is required to digest 1 μ g of substrate DNA in 16 hours.

Heat Inactivation: 70 units of enzyme were inactivated by incubation at 65°C for 20 minutes.

Plasmid Cleavage: Number of units required to cleave 1 μ g of supercoiled plasmid DNA in one hour: pBR322 = 3 units pUC 19 = 3 units.

Note: Cleaves to leave a 3' CATG extension which can be efficiently ligated to DNA fragments generated by NlaIII.

Not sensitive to *dam, dcm* or mammalian CpG methylation.

New icons (see www.neb.com for details)

🕐 = Time-Saver™ Qualified

 $\boldsymbol{\mathcal{C}}$ = indicates that the enzyme has been engineered

 \star = indicates that the enzyme has reduced star activity

U.S. Patent No. 5,262,318

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Note: Cleaves to leave a 3[°] CATG extension which can be efficiently ligated to DNA fragments generated by NIaIII.

Not sensitive to *dam, dcm* or mammalian CpG methylation.

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U.S. Patent No. 5,262,318