





1-800-632-7799 info@neb.com www.neb.com

# **R3136S**



10,000 units 20,000 U/ml Lot: 0061208 RECOMBINANT Store at -20°C Exp: 8/14

# **Recognition Site:**

5′... G A T C C ... 3′ 3′... C C T A G<sub>4</sub>G ... 5′

**Note:** BamHI-HF<sup>™</sup> has the same specificity as BamHI (NEB #R0136), but it has been engineered for reduced star activity.

## **More Units**













BioLabs

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5′... G<sup>T</sup>G A T C C ... 3′ 3′... C C T A G<sub>1</sub>G...5′

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#### **More Units**

**Source:** An *E. coli* strain that carries the cloned and modified (E163A/E167T)) BamHI gene from Bacillus amyloliquefaciens H (ATCC 49763)

Supplied in: 50 mM KCl. 10 mM Tris-HCl (pH 7.4). 0.1 mM EDTA, 1 mM dithiothreitol, 200 µg/ml BSA and 50% glycerol.

Reagents Supplied with Enzyme: 10X NEBuffer 4.

Reaction Conditions: 1X NEBuffer 4. Incubate at 37°C.

### 1X NEBuffer 4:

50 mM potassium acetate 20 mM Tris-acetate 10 mM magnesium acetate 1 mM DTT pH 7.9 at 25°C

Unit Definition: One unit is defined as the amount of enzyme required to digest 1  $\mu$ g of  $\lambda$  DNA in 1 hour at 37°C in a total reaction volume of 50 µl.

**Diluent Compatibility:** Diluent Buffer A 50 mM KCl. 10 mM Tris-HCl. 0.1 mM EDTA. 1 mM dithiothreitol, 200 µg/ml BSA and 50% glycerol (pH 7.4 @ 25°C).

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## **Quality Controls**

**Ligation:** After 50-fold overdigestion with BamHI-HF. > 95% of the DNA fragments can be ligated with T4 DNA Ligase (at a 5' termini concentration of 1–2  $\mu$ M) at 16°C. Of these ligated fragments, > 95% can be recut.

16-Hour Incubation: A 50 ul reaction containing 1 µg of DNA and 100 units of BamHI-HF incubated for 16 hours resulted in the same pattern of DNA bands as a reaction incubated for 1 hour with 1 unit of enzyme.

Exonuclease Activity: Incubation of 100 units of BamHI-HF with 1 µg sonicated 3H DNA (105 cpm/ ug) for 4 hours at 37°C in 50 ul reaction buffer released < 0.1% radioactivity.

Endonuclease Activity: Incubation of 100 units of BamHI-HF with 1 ug  $\phi$ X174 RF I DNA for 4 hours at 37°C in 50 µl reaction buffer resulted in < 10% conversion to RF II.

Blue/White Screening Assay: An appropriate vector is digested at a unique site within the lac $Z^{\alpha}$ gene with a 10-fold excess of enzyme. The vector DNA is then ligated, transformed, and plated onto Xgal/IPTG/Amp plates. Successful expression of β-galactosidase is a function of how intact its gene remains after cloning, an intact gene gives rise to a blue colony, removal of even a single base gives rise to a white colony. Enzyme preparations must produce fewer than 3% white colonies to be Blue/White certified.

## **Enzyme Properties**

Activity in NEBuffers: NEBuffer 1 100% NEBuffer 2 50% NEBuffer 3 10% NEBuffer 4 100%

When using a buffer other than the optimal (supplied) NEBuffer, it may be necessary to add more enzyme to achieve complete digestion.

(see other side)

CERTIFICATE OF ANALYSIS

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CERTIFICATE OF ANALYSIS

**Survival in a Reaction:** Intermediate activity. Suitable for extended digestion, but < 8 hours.

Heat Inactivation: No

Plasmid Cleavage: Number of units required to cleave 1 μg of supercoiled plasmid DNA in one hour: pUC19 = 5 units, pBR322 = 1 unit, LITMUS = 5 units.

**Notes:** Not sensitive to *dam, dcm* or mammalian CpG methylation.

Requires 5 units to cut pUC19 DNA plasmids or plasmids with similar flanking sequences.

The increased specificity for the BamHI-HF cut site has increased binding of the enzyme to the DNA and the enzyme may remain attached to the DNA during gel electrophoresis. To disrupt binding, add SDS to a final concentration of 0.5%–1% or purify DNA before electrophoresis.

New icons (see www.neb.com for details)

= Time-Saver™ Qualified

e = indicates that the enzyme has been engineered

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