

# PmeI



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R0560S 016121214121

## R0560S



500 units 10,000 U/ml Lot: 0161212

RECOMBINANT Store at -20°C Exp: 12/14

### Recognition Site:

5'...GTTT<sup>▼</sup>AAAC...3'  
3'...CAAAT<sup>▲</sup>TTG...5'

**Source:** An *E. coli* strain that carries the cloned PmeI gene from *Pseudomonas mendocina* (B. Zhou)

Supplied in: 100 mM NaCl, 10 mM Tris-HCl (pH 7.4), 0.1 mM EDTA, 1 mM dithiothreitol, 200 µg/ml BSA and 50% glycerol.

### Reagents Supplied with Enzyme:

10X NEBuffer 4, 100X BSA.

### Reaction Conditions:

1X NEBuffer 4, supplemented with 100 µg/ml BSA. Incubate at 37°C.

### 1X NEBuffer 4:

50 mM potassium acetate  
20 mM Tris-acetate  
10 mM magnesium acetate  
1 mM dithiothreitol  
pH 7.9 @ 25°C

**Unit Definition:** One unit is defined as the amount of enzyme required to digest 1 µg of λ DNA in 1 hour at 37°C in a total reaction volume of 50 µl.

**Diluent Compatibility:** Diluent Buffer A  
50 mM KCl, 10 mM Tris-HCl, 0.1 mM EDTA, 1 mM dithiothreitol, 200 µg/ml BSA and 50% glycerol (pH 7.4 @ 25°C)

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### Quality Control Assays

**Ligation:** After 10-fold overdigestion with PmeI, > 95% of the DNA fragments can be ligated with T4 DNA Ligase (at a 5' termini concentration of 1–2 µM) at 16°C. Of these ligated fragments, > 95% can be recut.

**16-Hour Incubation:** A 50 µl reaction containing 1 µg of DNA and 10 units of enzyme incubated for 16 hours resulted in the same pattern of DNA bands as a reaction incubated for 1 hour with 1 unit of enzyme.

**Exonuclease Activity:** Incubation of 100 units of enzyme with 1 µg sonicated [<sup>3</sup>H] DNA (10<sup>5</sup> cpm/µg) for 4 hours at 37°C in 50 µl reaction buffer released 0.05% radioactivity.

**Endonuclease Activity:** Incubation of 16 units of enzyme with 1 µg φX174 RF I DNA for 4 hours at 37°C in a 50 µl reaction buffer resulted in 5% conversion to RF II.

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**Endonuclease Activity:** Incubation of 16 units of enzyme with 1 µg φX174 RF I DNA for 4 hours at 37°C in a 50 µl reaction buffer resulted in 5% conversion to RF II.

**Blue/White Screening Assay:** This enzyme has been tested to determine the integrity of the DNA ends produced after digestion with an excess of enzyme. An appropriate vector is digested at a unique site within *lacZ<sup>α</sup>* gene with a 10-fold excess of enzyme, ligated, transformed and plated on XGal/IPTG/Amp plates. Successful expression of β-galactosidase is a function of how intact its gene remains after cloning, an intact gene gives rise to a blue colony, an interrupted gene (i.e. degraded DNA end) gives rise to a white colony. Enzymes must produce fewer than 3% white colonies to be Blue/White Certified.

### Enzyme Properties

#### Activity in NEBuffers:

NEBuffer 1	0%
NEBuffer 2	50%
NEBuffer 3	10%
NEBuffer 4	100%

When using a buffer other than the optimal (supplied) NEBuffer, it may be necessary to add more enzyme to achieve complete digestion.

(See other side)

CERTIFICATE OF ANALYSIS

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CERTIFICATE OF ANALYSIS

**Survival in a Reaction:** A minimum of 1.00 unit is required to digest 1 µg of substrate DNA in 16 hours.

**Heat Inactivation:** 20 units of enzyme were inactivated by incubation at 65°C for 20 minutes.

**Plasmid Cleavage:** Number of units required to cleave 1 µg of supercoiled plasmid DNA in one hour: pNEB193 = 4 units.


**Notes:** PmeI is an octanucleotide recognizing restriction endonuclease.

Cleavage of mammalian genomic DNA is blocked by some combinations of overlapping CpG methylation.

pNEB193 Vector (NEB #N3051), a cloning vector which contains single sites for NEB's unique 8-base cutters Ascl, PacI and PmeI is available.

Four units of PmeI are required for complete digestion of 1 µg of pNEB193.

For extended digests, incubation at 25°C increases enzyme stability.

 = Time-Saver™ Qualified (See [www.neb.com](http://www.neb.com) for details).

U.S. Patent No. 5,196,330 and 5,945,228

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