# **SmaI**



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**R0141S** 



2,000 units 20,000 U/ml Lot: 0841209 RECOMBINANT Store at -20°C Exp: 9/14

**Recognition Site:** 

5′...CCC GGG...3′ 3′...GGGCCC...5′

**Source:** An *E. coli* strain that carries the cloned Smal gene from *Serratia marcescens* (ATCC 49779)

Supplied in: 50 mM KCl, 10 mM Tris-HCl (pH 7.4), 0.1 mM EDTA, 1 mM DTT, 200  $\mu g/ml$  BSA and 50% alverol.

**Reagents Supplied with Enzyme:** 10X NEBuffer 4.

Reaction Conditions: 1X NEBuffer 4. Incubate at 25°C.

1X NEBuffer 4:

50 mM potassium acetate 20 mM Tris-acetate 10 mM magnesium acetate 1 mM DTT pH 7.9 @ 25°C

Unit Definition: One unit is defined as the amount of enzyme required to digest 1  $\mu g$  of  $\lambda$  DNA (HindIII digest) in 1 hour at 25°C in a total reaction volume of 50  $\mu l$ .

Diluent Compatibility: Diluent Buffer A 50 mM KCl, 10 mM Tris-HCl, 0.1 mM EDTA, 1 mM DTT, 200 μg/ml BSA and 50% glycerol (pH 7.4 @ 25°C)

## **Quality Control Assays**

**Ligation:** After 10-fold overdigestion with Smal, approximately 50% of the DNA fragments can be ligated with T4 DNA Ligase (at a 5' termini

concentration of 1–2  $\mu$ M) at 16°C. Of these ligated fragments, > 95% can be recut.

16-Hour Incubation: A 50 µl reaction containing 1 µg of DNA and 40 units of enzyme incubated for 16 hours resulted in the same pattern of DNA bands as a reaction incubated for 1 hour with 1 unit of enzyme.

**Exonuclease Activity:** Incubation of 80 units of enzyme with 1  $\mu$ g sonicated <sup>3</sup>H DNA (10<sup>5</sup> cpm/ $\mu$ g) for 4 hours at 37°C in 50  $\mu$ l reaction buffer released < 0.1% radioactivity.

Endonuclease Activity: Incubation of 50 units of enzyme with 1  $\mu$ g  $\phi$ X174 RF I DNA for 4 hours at 37°C in 50  $\mu$ l reaction buffer resulted in < 15% conversion to RF II.

Blue/White Screening Assay: This enzyme has been tested to determine the integrity of the DNA ends produced after digestion with an excess of enzyme. An appropriate vector is digested at a unique site within  $lacZ^{\alpha}$  gene with a 5-fold excess of enzyme, ligated, transformed and plated on XGal/IPTG/Amp plates. Successful expression of  $\beta$ -galactosidase is a function of how intact its

gene remains after cloning, an intact gene gives rise to a blue colony, an interrupted gene (i.e. degraded DNA end) gives rise to a white colony. Enzymes must produce fewer than 3% white colonies to be Blue/White Certified.

# **Enzymes Properties**

**Activity in NEBuffers:** 

NEBuffer 1 0% NEBuffer 2 0% NEBuffer 3 0% NEBuffer 4 **100%** 

When using a buffer other than the optimal (supplied) NEBuffer, it may be necessary to add more enzyme to achieve complete digestion.

Survival in a Reaction: A minimum of 0.13 unit is required to digest 1  $\mu g$  of substrate DNA in 16 hours.

**Heat Inactivation:** 50 units of enzyme were inactivated by incubation at 65°C for 20 minutes.

Notes: Smal is an isoschizomer of Xmal.

(See other side)

CERTIFICATE OF ANALYSIS

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CERTIFICATE OF ANALYSIS

Smal produces blunt-ended fragments whereas Xmal produces a 5' extension. Smal has a half-life of 15 minutes at 37°C. Incubation at 37°C results in 50% activity.

Cleavage of mammalian genomic DNA is blocked by CpG methylation.

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