

M0208S

2,000 units 40,000 U/ml Lot: 0071209 RECOMBINANT Store at -20°C Exp: 9/14

Description: Tag DNA Ligase catalyzes the formation of a phosphodiester bond between juxtaposed 5' phosphate and 3' hydroxyl termini of two adjacent oligonucleotides which are hybridized to a complementary target DNA. The ligation will occur only if the oligonucleotides are perfectly paired to the complementary target DNA and have no gaps between them; therefore, a single-base substitution can be detected. *Tag* DNA Ligase is active at elevated temperatures (45-65°C) (1,2).



M0208S

R: 164

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Lot: 0071209 2,000 units 40,000 U/ml RECOMBINANT Store at -20°C Exp: 9/14

Description: Taq DNA Ligase catalyzes the formation of a phosphodiester bond between juxtaposed 5' phosphate and 3' hydroxyl termini of two adjacent oligonucleotides which are hybridized to a complementary target DNA. The ligation will occur only if the oligonucleotides are perfectly paired to the complementary target DNA and have no gaps between them; therefore, a single-base substitution can be detected. *Tag* DNA Ligase is active at elevated temperatures (45-65°C) (1,2).

Source: Purified from an E. coli strain containing the cloned ligase gene from *Thermus aquaticus* HB8 (1)

Supplied in: 50 mM KCI. 10 mM Tris-HCI (pH 7.4). 0.1 mM EDTA, 1 mM DTT, 200 µg/ml BSA and 50% alvcerol.

Applications:

R: 164

- Allele-specific gene detection using Ligase Detection Reaction and Ligase Chain Reaction (1.3).
- Mutagenesis by incorporation of a phosphorylated oligonucleotide during PCR amplification (4).

Reagents Supplied with Enzyme: 10X Tag DNA Ligase Reaction Buffer and 5 µg control DNA (BstEII-digested λ DNA).

Reaction Conditions: Incubate DNA and enzyme in 1X Taq DNA Ligase Buffer at 45°C for 15 minutes or in a thermocycler with a program suited to the reaction described by Barany (1991) Genetic Disease Detection and DNA Amplification Using Cloned Thermostable Ligase. Proc. Natl. Acad. Sci. USA 88, 189–193, "The reaction is stopped with a mixture of 50% glycerol, 50 mM EDTA, bromphenol blue."

Source: Purified from an *E. coli* strain containing the cloned ligase gene from *Thermus aquaticus* HB8 (1)

Supplied in: 50 mM KCl, 10 mM Tris-HCl (pH 7.4), 0.1 mM EDTA, 1 mM DTT, 200 µg/ml BSA and 50% alvcerol.

Applications:

- Allele-specific gene detection using Ligase Detection Reaction and Ligase Chain Reaction (1,3).
- Mutagenesis by incorporation of a phosphorylated oligonucleotide during PCR amplification (4).

Reagents Supplied with Enzyme: 10X Tag DNA Ligase Reaction Buffer and 5 µg control DNA (BstEII-digested λ DNA).

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1X Tag DNA Ligase Reaction Buffer:

20 mM Tris-HCl 25 mM potassium acetate 10 mM magnesium acetate 10 mM dithiothreitol 1 mM NAD 0.1% Triton X-100 (pH 7.6 @ 25°C)

Requires NAD⁺ as a cofactor. NAD⁺ is supplied in the 10X Tag DNA Ligase Reaction Buffer; the buffer should be stored at -70°C to extend the half life of the NAD⁺ cofactor.

Unit Definition: (Cohesive End Unit) One unit is defined as the amount of enzyme required to give 50% ligation of the 12-base pair cohesive ends of 1 μ g of BstEll-digested λ DNA in a total reaction volume of 50 µl in 15 minutes at 45°C.

Unit Assay Conditions: 1X Tag DNA Ligase Reaction Buffer and DNA (20 µg/ml). After incubation at 45°C for 15 minutes, the reaction is terminated by addition of stop dye (50% glycerol, 50 mM EDTA and bromophenol blue), heated at 70°C for 10 minutes and then loaded on a 0.7% agarose gel. Due to the presence of ligase, the cos ends of BstEll-digested λ DNA will stay together after 70°C heat treatment.

1X Tag DNA Ligase Reaction Buffer:

20 mM Tris-HCI 25 mM potassium acetate 10 mM magnesium acetate 10 mM dithiothreitol 1 mM NAD 0.1% Triton X-100 (pH 7.6 @ 25°C)

Requires NAD⁺ as a cofactor. NAD⁺ is supplied in the 10X Tag DNA Ligase Reaction Buffer; the buffer should be stored at -70°C to extend the half life of the NAD⁺ cofactor.

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Heat Inactivation: No.

Quality Control Assays

Exonuclease Activity: Incubation of 1,200 units for 4 hours at 37°C in 50 µl of thermostable ligase buffer containing 1 ug sonicated ³H DNA $(10^5 \text{ cpm/}\mu\text{g})$ gave < 0.1% acid soluble counts.

Nuclease Activity: Incubation of 4,000 units for 16 hours at 37°C in the recommended assav buffer plus NAD with Smal-Sall fragments of λ DNA does not alter the expected banding pattern on agarose gels. Incubation of HindIII fragments of λ DNA with 80 units of enzyme for 16 hours at 37°C in the recommended assav buffer without NAD does not alter the λ HindIII banding pattern on agarose gels.

Endonuclease Activity: Incubation of 1,500 units of enzyme for 4 hours at 37°C in 50 µl of assay buffer without NAD and 1 µg ϕ X174 RF I DNA gave < 10% conversion to RF II.

(see other side)

CERTIFICATE OF ANALYSIS

Heat Inactivation: No

Quality Control Assays

Exonuclease Activity: Incubation of 1,200 units for 4 hours at 37°C in 50 µl of thermostable ligase buffer containing 1 ug sonicated ³H DNA (10^5 cpm/µg) gave < 0.1% acid soluble counts.

Nuclease Activity: Incubation of 4,000 units for 16 hours at 37°C in the recommended assav buffer plus NAD with Smal-Sall fragments of λ DNA does not alter the expected banding pattern on agarose gels. Incubation of HindIII fragments of λ DNA with 80 units of enzyme for 16 hours at 37°C in the recommended assav buffer without NAD does not alter the λ HindIII banding pattern on agarose gels.

Endonuclease Activity: Incubation of 1,500 units of enzyme for 4 hours at 37°C in 50 µl of assay buffer without NAD and 1 µg ϕ X174 RF I DNA gave < 10% conversion to RF II.

(see other side)



References:

- Barany, F. (1991) Proc. Natl. Acad. Sci. USA 88, 189-193.
- Takahashi, M. et al. (1984) *J. Biol. Chem.* 259, 10041–10047.
- Barany, F. (1991) *The Ligase Chain Reaction in a* PCR World (pp. 5–16). Cold Spring Harbor: Cold Spring Harbor Laboratory Press.
- 4. Michael, S.F. (1994) *Biotechniques* 16, 411–412.

Taq DNA Ligase: Notice to Purchaser: This product is designed to ligate DNA fragments at temperatures requiring a thermoactive and thermostable enzyme. The seller is aware that the product may be used in the Ligase Chain Reaction[™] (LCR[™]) process covered by one of more claims of a pending patent application or issued patent assigned to Cornell Research Foundation Inc., or Cornell Research Foundation, Inc. and the California Institute of Technology. LCR[™] license inquires should be directed to Cornell Research Foundation, Inc.

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References:

- 1. Barany, F. (1991) *Proc. Natl. Acad. Sci. USA* 88, 189-193.
- Takahashi, M. et al. (1984) J. Biol. Chem. 259, 10041–10047.
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