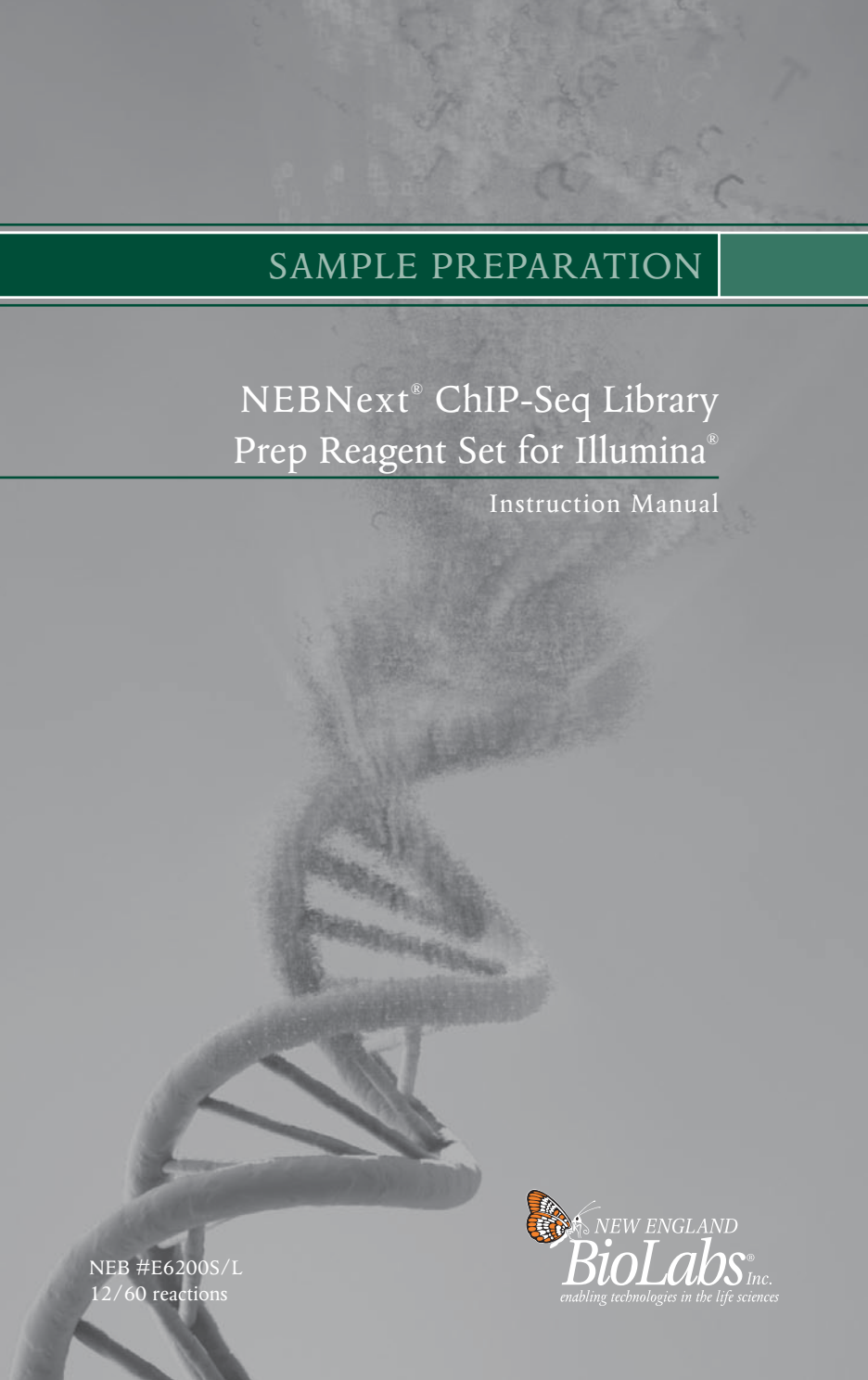
SAMPLE PREPARATION

NEBNext® ChIP-Seq Library Prep Reagent Set for Illumina®

Instruction Manual



NEB #E6200S/L
12/60 reactions

NEW ENGLAND BioLabs® Inc.
enabling technologies in the life sciences

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nqa ISO 9001 Registered Quality Management

nqa ISO 14001 Registered Environmental Management

nqa ISO 13485 Registered Medical Devices

Cloned at NEBiolabs | Recombinant Enzyme | NEB 2 Optimum Buffer | BSA Requires BSA | Yes Heat Inactivation

Table of Contents:

The Reagent Set Includes:

The volumes provided are sufficient for preparation of up to 12 reactions (NEB #E6200S) and 60 reactions (NEB #E6200L). (All reagents should be stored at –20°C).

Phosphorylation Reaction Buffer (10X)

Deoxynucleotide Solution Mix (10 mM each dNTP)

T4 DNA Polymerase RR

DNA Polymerase I, Large (Klenow) Fragment RR

T4 Polynucleotide Kinase RR

Deoxyadenosine 5´- Triphosphate (dATP) (1.0 mM)

Klenow Fragment (3´→5´ exo$^-$) RR

NEBuffer 2 for Klenow Fragment (3´→5´ exo$^-$) (10X)

Quick T4 DNA Ligase RR

Quick Ligation Reaction Buffer (2X)

Phusion High-Fidelity PCR Master Mix with HF Buffer (2X)

NEBNext High-Fidelity 2X PCR Master Mix (SAMPLE)

Applications:

The NEBNext ChIP-Seq Library Prep Reagent Set for Illumina contains enzymes and buffers that are ideally suited for sample preparation for next-generation sequencing of Chromatin Immunoprecipitated DNA. Each of these components must pass rigorous quality control standards and are Lot Controlled, both individually and as a set of reagents.

Lot Control: The lots provided in the NEBNext ChIP-Seq Library Prep Reagent Set for Illumina are managed separately and are qualified by additional functional validation. Individual reagents undergo standard enzyme activity and quality control assays, and also meet stringent criteria in the additional quality controls listed on each individual component page.

Functionally Validated: Each set of reagents is functionally validated together through construction and sequencing of a ChIP DNA library on an Illumina Sequencer (Illumina, Inc.).

For larger volume requirements, customized and bulk packaging is available by purchasing through the OEM/Bulks department at NEB. Please contact OEM@neb.com for further information.

Protocol for Use With NEBNext Singleplex (#E7350) or Multiplex Oligos for Illumina (#E7335, #E7500):

Starting Material: 10 ng of chromatin-immunoprecipitated (ChIP) qPCR verified or control DNA, in ≤ 40 µl of water or elution buffer

End Repair of ChIP DNA

1. Dilute DNA Polymerase I, Large (Klenow) Fragment by mixing 1 µl of enzyme with 4 µl of sterile water in a fresh microfuge tube.
2. In a sterile microfuge tube mix the following components:

ChIP DNA	1–40 µl
Phosphorylation Reaction Buffer (10X)	5 µl
dNTP mix	2 µl
T4 DNA Polymerase	1 µl
Diluted DNA Pol I Klenow Fragment	1 µl
T4 Polynucleotide Kinase	1 µl
Sterile H_2O	variable
Total volume	50 µl

3. Incubate in a thermal cycler for 30 minutes at 20°C.

Clean Up Using AMPure® XP Beads (Beckman Coulter, Inc.)

1. Vortex beads to resuspend.
2. Add 90 µl (1.8X) of resuspended AMPure XP beads to the reaction (~ 50 µl). Mix well on a vortex mixer or by pipetting up and down at least 10 times.
3. Incubate for 5 minutes at room temperature.
4. Put the tube/PCR plate on an appropriate magnetic stand to separate beads from supernatant. After the solution is clear (about 5 minutes), carefully remove and discard the supernatant. Be careful not to disturb the beads that contain DNA targets.
5. Add 200 µl of 80% ethanol to the tube/PCR plate while in the magnetic stand. Incubate at room temperature for 30 seconds, and then carefully remove and discard the supernatant.
6. Repeat Step 5 once.
7. Air dry beads for 10 minutes while the tube/PCR plate is on the magnetic stand with the lid open.
8. Elute DNA target from beads into 36 µl of 0.1X TE. Mix well on a vortex mixer or by pipetting up and down, and put the tube/PCR plate in the magnetic stand until the solution is clear.
9. Transfer 34 µl of the supernatant to a clean LoBind® (Eppendorf AG) tube, and store at –20°C.

Alternatively, purify DNA sample on one purification column and elute in 34 µl of sterile dH_2O or elution buffer.

dA-Tailing of End Repaired DNA

1. Mix the following components in a sterile microfuge tube:

End Repaired DNA	34 µl
NEBuffer 2	5 µl
dATP	10 µl
Klenow Fragment (3´→5´ exo⁻)	1 µl
Total volume	50 µl

2. Incubate at 37°C for 30 minutes.

Clean up Using AMPure XP Beads

1. Vortex beads to resuspend.
2. Add 90 µl (1.8X) of resuspended AMPure XP beads to the dA tailing reaction(~ 50 µl). Mix well on a vortex mixer or by pipetting up and down at least 10 times.
3. Incubate for 5 minutes at room temperature.
4. Put the tube/PCR plate on an appropriate magnetic stand to separate beads from supernatant. After the solution is clear (about 5 minutes), carefully remove and discard the supernatant. Be careful not to disturb the beads that contain DNA targets.
5. Add 200 µl of 80% ethanol to the tube/PCR plate while in the magnetic stand. Incubate at room temperature for 30 seconds, and then carefully remove and discard the supernatant.
6. Repeat Step 5 once.
7. Air dry beads for 10 minutes while the tube/PCR plate is on the magnetic stand with the lid open.
8. Elute DNA target from beads into 12 µl of 0.1X TE. Mix well on a vortex mixer or by pipetting up and down, and put the tube/PCR plate in the magnetic stand until the solution is clear.
9. Transfer 10 µl of the supernatant to a clean LoBind tube, and store at –20°C.

Alternatively, purify DNA sample on one purification column and elute in 10 µl of sterile dH_2O or elution buffer.

Adaptor Ligation of dA-Tailed DNA

Note: Dilute the NEBNext Adaptor (15 µM) to 1.5 µM in Nuclease Free water for immediate use.

1. Mix the following components in a sterile microfuge tube:

End Repaired, dA-Tailed DNA	10 µl
Quick Ligation Reaction Buffer (2X)	15 µl
Diluted NEBNext Adaptor (1.5 µM)	1 µl
Quick T4 DNA Ligase	4 µl
Total volume	30 µl

2. Incubate at 20°C for 15 minutes.
3. Add 3 µl of USER™ enzyme, mix by pipetting up and down, and incubate at 37°C for 15 minutes.

Clean Up Using AMPure XP Beads

1. Vortex AMPure XP beads to resuspend
2. Add 54 µl (1.8X) of resuspended AMPure XP beads to the ligation reaction (~ 30 µl). Mix well on a vortex mixer or by pipetting up and down at least 10 times.
3. Incubate for 5 minutes at room temperature.
4. Put the tube/PCR plate on an appropriate magnetic stand to separate beads from supernatant. After the solution is clear (about 5 minutes), carefully remove and discard the supernatant. Be careful not to disturb the beads that contain DNA targets.
5. Add 200 µl of 80% ethanol to the tube/PCR plate while in the magnetic stand. Incubate at room temperature for 30 seconds, and then carefully remove and discard the supernatant.
6. Repeat Step 5 once.
7. Air dry beads for 10 minutes while the tube/PCR plate is on the magnetic stand with the lid open.
8. Elute DNA target from beads into 105 µl of dH_2O. Mix well on a vortex mixer or by pipetting up and down, and put the tube/PCR plate in the magnetic stand until the solution is clear.
9. Transfer 100 µl of the supernatant to a clean tube/PCR plate.

Alternatively, purify DNA sample on a single purification column and elute in the appropriate volume of sterile dH_2O or elution buffer for desired size selection.

Size selection of Adaptor Ligated DNA Using AMPure XP Beads

Note: (X) refers to original sample volume of 100 µl.

1. Add 90 µl (0.9X) resuspended AMPure XP beads to 100 µl DNA solution. Mix well on a vortex mixer or by pipetting up and down at least 10 times.
2. Incubate for 5 minutes at room temperature.
3. Place the tube/PCR plate on an appropriate magnetic stand to separate beads from supernatant. After the solution is clear (about 5 minutes), carefully transfer the supernatant to a new tube/well **(Caution: do not discard the supernatant).** Discard beads that contain the large fragments.
4. Add 20 µl (0.2X) resuspended AMPure XP beads to the supernatant, mix well and incubate for 5 minutes at room temperature.
5. Put the tube/PCR plate on an appropriate magnetic stand to separate beads from supernatant. After the solution is clear (about 5 minutes), carefully remove and discard the supernatant. Be careful not to disturb the beads that contain DNA targets **(Caution: do not discard beads).**
6. Add 200 µl of 80% freshly prepared ethanol to the tube/PCR plate while in the magnetic stand. Incubate at room temperature for 30 seconds, and then carefully remove and discard the supernatant.
7. Repeat Step 6 once.
8. Air dry beads for 10 minutes while the tube/PCR plate is on the magnetic stand with lid open.
9. Elute DNA target from beads into 25 µl water or 0.1X TE buffer. Mix well on a vortex mixer or by pipetting up and down, and put the tube/PCR plate in the magnetic stand until the solution is clear.
10. Transfer 23 µl of the supernatant to a clean PCR tube and proceed to enrichment.

Alternatively, size selection can be performed using a number of methods including E-Gel® (Life Technologies, Inc.) size select gels or standard 2% agarose gels. NEB's 100 bp ladder (NEB #N3231) can be used to determine the size of the fragments. Isolate library fragments in the 175–225 base pair range.

Purify the DNA on one purification column and elute in 25 µl of sterile water or elution buffer.

PCR Enrichment of Adaptor Ligated DNA

1. Mix the following components in a sterile microfuge tube:

Adaptor ligated DNA	23 µl
NEBNext High-Fidelity 2X PCR Master Mix**	25 µl
Universal PCR Primer (25 µM)	1 µl
Index 1 Primer* (25 µM)	1 µl
Total volume	50 µl

* If you are using the NEBNext Multiplex Oligos for Illumina (#E7335, #E7500), for each reaction, only one of the 12 PCR primer indices is used during the PCR step.

** NEBNext High-Fidelity 2X PCR Master Mix will be replacing Phusion High-Fidelity PCR Master Mix. Both vials will be supplied for a limited time only.

2. PCR cycling conditions

CYCLE STEP	TEMP	TIME	CYCLES
Initial Denaturation	98°C	30 sec	1
Denaturation Annealing Extension	98°C 65°C 72°C	10 sec 30 sec 30 sec	15
Final Extension	72°C 4°C	5 min hold	1

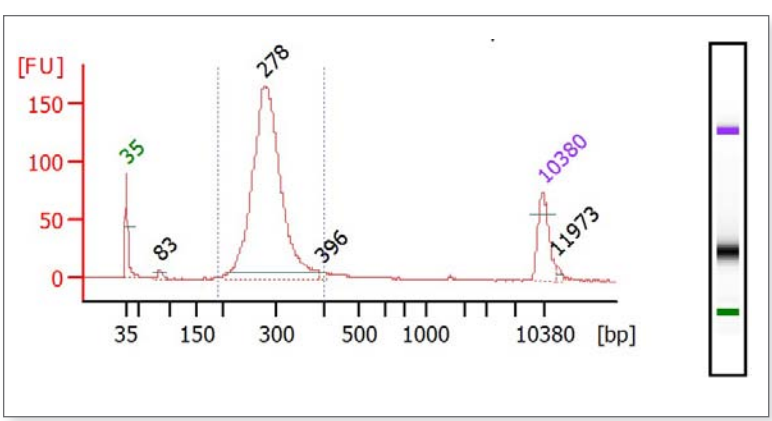
Clean Up Using AMPure XP Beads

1. Vortex beads to resuspend.
2. Add 50 µl (1X) of resuspended AMPure XP beads to the PCR reactions (~ 50 µl). Mix well on a vortex mixer or by pipetting up and down at least 10 times.
3. Incubate for 5 minutes at room temperature.
4. Put the tube/PCR plate on an appropriate magnetic stand to separate beads from supernatant. After the solution is clear (about 5 minutes), carefully remove and discard the supernatant. Be careful not to disturb the beads that contain DNA targets.
5. Add 200 µl of 80% ethanol to the tube/PCR plate while in the magnetic stand. Incubate at room temperature for 30 seconds, and then carefully remove and discard the supernatant.
6. Repeat Step 5 once.
7. Air dry beads for 10 minutes while the tube/PCR plate is on the magnetic stand with the lid open.
8. Elute DNA target from beads into 17 µl of 0.1X TE. Mix well on a vortex mixer or by pipetting up and down, and put the tube/PCR plate in the magnetic stand until the solution is clear.
9. Transfer 15 µl of the supernatant to a clean LoBind tube, and store at –20°C.

Alternatively, purify sample on one purification column and elute in 15 µl of sterile water or elution buffer.

Dilute the library 20 fold with nuclease free water, and assess the library quality on a Bioanalyzer® (Agilent high sensitivity chip) (Agilent Technologies, Inc.). Check that the electropherogram shows a narrow distribution with a peak size around 275 bp is expected (example shown below).

Figure 1: Bioanalyzer traces of final library.



Protocol for Use With End User Supplied Primers and Adaptors:

Starting Material: 10 ng of chromatin-immunoprecipitated (ChIP) qPCR verified or control DNA, in ≤ 40 µl of water or elution buffer

End Repair of ChIP DNA

1. Dilute DNA Polymerase I, Large (Klenow) Fragment by mixing 1 µl of enzyme with 4 µl of sterile water in a fresh microfuge tube.
2. In a sterile microfuge tube mix the following components:

ChIP DNA	1–40 µl
Phosphorylation Reaction Buffer (10X)	5 µl
dNTP mix	2 µl
T4 DNA Polymerase	1 µl
Diluted DNA Pol I Klenow Fragment	1 µl
T4 Polynucleotide Kinase	1 µl
Sterile H_2O	variable
Total volume	50 µl

3. Incubate in a thermal cycler for 30 minutes at 20°C.

Clean Up Using AMPure XP Beads

1. Vortex beads to resuspend.
2. Add 90 µl (1.8X) of resuspended AMPure XP beads to the reaction (~ 50 µl). Mix well on a vortex mixer or by pipetting up and down at least 10 times.
3. Incubate for 5 minutes at room temperature.
4. Put the tube/PCR plate on an appropriate magnetic stand to separate beads from supernatant. After the solution is clear (about 5 minutes), carefully remove and discard the supernatant. Be careful not to disturb the beads that contain DNA targets.
5. Add 200 µl of 80% ethanol to the tube/PCR plate while in the magnetic stand. Incubate at room temperature for 30 seconds, and then carefully remove and discard the supernatant.
6. Repeat Step 5 once.
7. Air dry beads for 10 minutes while the tube/PCR plate is on the magnetic stand with the lid open.
8. Elute DNA target from beads into 36 µl of 0.1X TE. Mix well on a vortex mixer or by pipetting up and down, and put the tube/PCR plate in the magnetic stand until the solution is clear.
9. Transfer 34 µl of the supernatant to a clean LoBind tube, and store at –20°C.

Alternatively, purify DNA sample on one purification column and elute in 34 µl of sterile dH_2O or elution buffer.

dA-Tailing of End Repaired DNA

1. Mix the following components in a sterile microfuge tube:

End Repaired DNA	34 µl
NEBuffer 2	5 µl
dATP	10 µl
Klenow Fragment (3´→5´ exo$^-$)	1 µl
Total volume	50 µl

2. Incubate at 37°C for 30 minutes.

Clean up Using AMPure XP Beads

1. Vortex beads to resuspend.
2. Add 90 µl (1.8X) of resuspended AMPure XP beads to the dA tailing reaction (~ 50 µl). Mix well on a vortex mixer or by pipetting up and down at least 10 times.
3. Incubate for 5 minutes at room temperature.
4. Put the tube/PCR plate on an appropriate magnetic stand to separate beads from supernatant. After the solution is clear (about 5 minutes), carefully remove and discard the supernatant. Be careful not to disturb the beads that contain DNA targets.
5. Add 200 µl of 80% ethanol to the tube/PCR plate while in the magnetic stand. Incubate at room temperature for 30 seconds, and then carefully remove and discard the supernatant.
6. Repeat Step 5 once.
7. Air dry beads for 10 minutes while the tube/PCR plate is on the magnetic stand with the lid open.
8. Elute DNA target from beads into 12 µl of 0.1X TE. Mix well on a vortex mixer or by pipetting up and down, and put the tube/PCR plate in the magnetic stand until the solution is clear.
9. Transfer 10 µl of the supernatant to a clean LoBind tube, and store at –20°C.

Alternatively, purify DNA sample on one purification column and elute in 10 µl of sterile dH_2O or elution buffer.

Adaptor Ligation of dA-Tailed DNA

1. Mix the following components in a sterile microfuge tube:

End Repaired, dA-Tailed DNA	10 µl
Quick Ligation Reaction Buffer (2X)	15 µl
1.5 µM DNA Adaptors*	1 µl
Quick T4 DNA Ligase	4 µl
Total volume	30 µl

* Adaptors are not included, use adaptors appropriate to specific application. If necessary adjust the adaptor concentration to obtain a final adaptor to DNA molar ratio of 10:1.

2. Incubate at 20°C for 15 minutes.

Clean Up Using AMPure XP Beads

1. Vortex AMPure XP beads to resuspend
2. Add 54 µl (1.8X) of resuspended AMPure XP beads to the ligation reaction (~ 30 µl). Mix well on a vortex mixer or by pipetting up and down at least 10 times.
3. Incubate for 5 minutes at room temperature.
4. Put the tube/PCR plate on an appropriate magnetic stand to separate beads from supernatant. After the solution is clear (about 5 minutes), carefully remove and discard the supernatant. Be careful not to disturb the beads that contain DNA targets.
5. Add 200 µl of 80% ethanol to the tube/PCR plate while in the magnetic stand. Incubate at room temperature for 30 seconds, and then carefully remove and discard the supernatant.
6. Repeat Step 5 once.
7. Air dry beads for 10 minutes while the tube/PCR plate is on the magnetic stand with the lid open.
8. Elute DNA target from beads into 105 µl of dH_2O. Mix well on a vortex mixer or by pipetting up and down, and put the tube/PCR plate in the magnetic stand until the solution is clear.
9. Transfer 100 µl of the supernatant to a clean tube/PCR plate.

Alternatively, purify DNA sample on one purification column and elute in appropriate volume of sterile dH_2O or elution buffer for desired size selection.

Size selection of Adaptor Ligated DNA Using AMPure XP Beads

Note: (X) refers to original sample volume of 100 µl.

1. Add 90 µl (0.9X) resuspended AMPure XP beads to 100 µl DNA solution. Mix well on a vortex mixer or by pipetting up and down at least 10 times.
2. Incubate for 5 minutes at room temperature.
3. Place the tube/PCR plate on an appropriate magnetic stand to separate beads from supernatant. After the solution is clear (about 5 minutes), carefully transfer the supernatant to a new tube/well **(Caution: do not discard the supernatant).** Discard beads that contain the large fragments.
4. Add 20 µl (0.2X) resuspended AMPure XP beads to the supernatant, mix well and incubate for 5 minutes at room temperature.
5. Put the tube/PCR plate on an appropriate magnetic stand to separate beads from supernatant. After the solution is clear (about 5 minutes), carefully remove and discard the supernatant. Be careful not to disturb the beads that contain DNA targets **(Caution: do not discard beads).**
6. Add 200 µl of 80% freshly prepared ethanol to the tube/PCR plate while in the magnetic stand. Incubate at room temperature for 30 seconds, and then carefully remove and discard the supernatant.
7. Repeat Step 6 once.
8. Air dry beads for 10 minutes while the tube/PCR plate is on the magnetic stand with lid open.
9. Elute DNA target from beads into 25 µl water or 0.1X TE buffer. Mix well on a vortex mixer or by pipetting up and down, and put the tube/PCR plate in the magnetic stand until the solution is clear.
10. Transfer 23 µl of the supernatant to a clean PCR tube and proceed to enrichment.

Alternatively, size selection can be performed using a number of methods including E-Gel size select gels or standard 2% agarose gels. NEB's 100 bp ladder (NEB #N3231) can be used to determine the size of the fragments. Isolate library fragments in the 175–225 base pair range.

Purify the DNA on one purification column and elute in 25 µl of sterile water or elution buffer.

PCR Enrichment of Adaptor Ligated DNA

1. Mix the following components in a sterile microfuge tube:

Adaptor ligated DNA	23 µl
NEBNext High-Fidelity 2X PCR Master Mix**	25 µl
Primer 1* (25 µM)	1 µl
Primer 2* (25 µM)	1 µl
Total volume	50 µl

* Primers are not included, use primers appropriate to specific application.

** NEBNext High-Fidelity 2X PCR Master Mix will be replacing Phusion High-Fidelity PCR Master Mix. Both vials will be supplied for a limited time only.

2. PCR cycling conditions

CYCLE STEP	TEMP	TIME	CYCLES
Initial Denaturation	98°C	30 sec	1
Denaturation Annealing Extension	98°C 65°C 72°C	10 sec 30 sec 30 sec	15
Final Extension	72°C 4°C	5 min hold	1

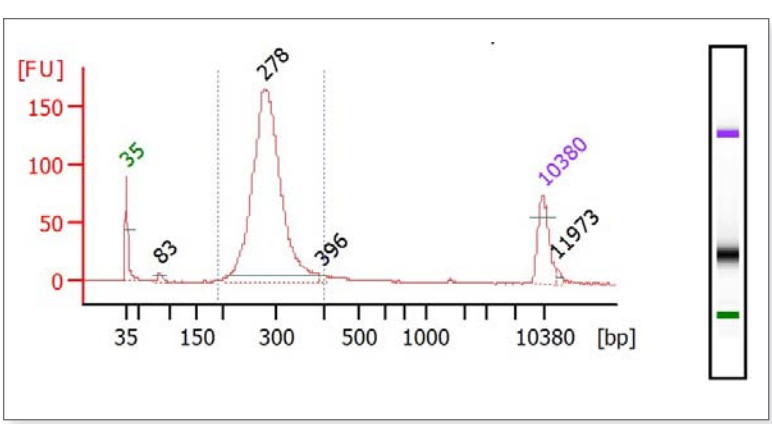
Clean Up Using AMPure XP Beads

1. Vortex beads to resuspend.
2. Add 50 µl (1X) of resuspended AMPure XP beads to the PCR reactions (~ 50 µl). Mix well on a vortex mixer or by pipetting up and down at least 10 times.
3. Incubate for 5 minutes at room temperature.
4. Put the tube/PCR plate on an appropriate magnetic stand to separate beads from supernatant. After the solution is clear (about 5 minutes), carefully remove and discard the supernatant. Be careful not to disturb the beads that contain DNA targets.
5. Add 200 µl of 80% ethanol to the tube/PCR plate while in the magnetic stand. Incubate at room temperature for 30 seconds, and then carefully remove and discard the supernatant.
6. Repeat Step 5 once.
7. Air dry beads for 10 minutes while the tube/PCR plate is on the magnetic stand with the lid open.
8. Elute DNA target from beads into 17 µl of 0.1X TE. Mix well on a vortex mixer or by pipetting up and down, and put the tube/PCR plate in the magnetic stand until the solution is clear.
9. Transfer 15 µl of the supernatant to a clean LoBind tube, and store at –20°C.

Alternatively, purify sample on one purification column and elute in 15 µl of sterile water or elution buffer.

Dilute the library 20 fold with nuclease free water, and assess the library quality on a Bioanalyzer (Agilent high sensitivity chip). Check that the electropherogram shows a narrow distribution with a peak size around 275 bp is expected (example shown below).

Figure 2: Bioanalyzer traces of final library.



Phosphorylation Reaction Buffer

#E6201A: 0.06 ml **Concentration: 10X**
#E6201AA: 0.3 ml

Store at –20°C

1X Phosphorylation Reaction Buffer:
50 mM Tris-HCl
10 mM $MgCl_2$
10 mM DTT
1 mM ATP
pH 7.5 @ 25°C

Quality Control Assays

16-Hour Incubation: 50 µl reactions containing this reaction buffer at 1X concentration and 1 µg of HindIII digested Lambda DNA incubated for 16 hours at 37°C results in no detectable non-specific nuclease degradation as determined by agarose gel electrophoresis. 50 µl reactions containing this reaction buffer at 1X concentration and 1 µg T3 DNA incubated for 16 hours at 37°C also results in no detectable non-specific nuclease degradation as determined by agarose gel electrophoresis.

Endonuclease Activity: Incubation of this reaction buffer at a 1X concentration with 1 µg of ϕX174 RF I DNA for 4 hours at 37°C in 50 µl reactions results in less than 10% conversion to RF II as determined by agarose gel electrophoresis.

RNase Activity: Incubation of this reaction buffer at 1X concentration with 40 ng of a FAM-labeled RNA transcript for 16 hours at 37°C results in no detectable RNase activity as determined by polyacrylamide gel electrophoresis.

Phosphatase Activity: Incubation of this reaction buffer at a 1X concentration in protein phosphatase assay buffer (1 M diethanolamine @ pH 9.8 and 0.5 mM $MgCl_2$) containing 2.5 mM *p*-nitrophenyl phosphate at 37°C for 4 hours yields no detectable *p*-nitrophenylene anion as determined by spectrophotometric analysis at 405 nm.

Lot Controlled

Deoxynucleotide Solution Mix

#E6202A: 0.024 ml 10 mM each dNTP
#E6202AA: 0.120 ml

Store at –20°C

Description: Deoxynucleotide Solution Mix is an equimolar solution of ultrapure dATP, dCTP, dGTP and dTTP.

Supplied in: Milli-Q® water (Millipore Corporation) as a sodium salt at pH 7.5.

Concentration: Each nucleotide is supplied at a concentration of 10 mM. (40 mM total nucleotide concentration).

Quality Assurance: Nucleotide solutions are certified free of nucleases and phosphatases.

Notes: Storing nucleotide triphosphates in solutions containing magnesium promotes triphosphate degradation.

Quality Control Assays

16-Hour Incubation: 50 µl reactions containing a minimum of 2 mM dNTPs and 1 µg of HindIII digested Lambda DNA incubated for 16 hours at 37°C results in no detectable non-specific nuclease degradation as determined by agarose gel electrophoresis. 50 µl reactions containing a minimum of 2 mM dNTPs and 1 µg T3 DNA incubated for 16 hours at 37°C also results in no detectable non-specific nuclease degradation as determined by agarose gel electrophoresis.

RNase Activity: Incubation of 1 mM dNTPs with 40 ng of a FAM-labeled RNA transcript for 16 hours at 37°C results in no detectable RNase activity as determined by polyacrylamide gel electrophoresis.

Phosphatase Activity: Incubation of a minimum of 5 mM dNTPs in protein phosphatase assay buffer (1 M diethanolamine @ pH 9.8 and 0.5 mM $MgCl_2$) containing 2.5 mM *p*-nitrophenyl phosphate at 37°C for 4 hours yields no detectable *p*-nitrophenylene anion as determined by spectrophotometric analysis at 405 nm.

HPLC: dNTP purity is determined by HPLC to be > 99%.

Functional Activity (PCR): The dNTPs are tested in 25 cycles of PCR amplification generating 0.5 kb, 2 kb, and 5kb amplicons from lambda DNA.

Lot Controlled

T4 DNA Polymerase

#E6203A: 0.015 ml
#E6203AA: 0.06 ml

RR BSA Yes

Store at –20°C

Description: T4 DNA Polymerase catalyzes the synthesis of DNA in the 5´→ 3´ direction and requires the presence of template and primer. This enzyme has a 3´→ 5´ exonuclease activity which is much more active than that found in DNA Polymerase I. Unlike *E. coli* DNA Polymerase I, T4 DNA Polymerase does not have a 5´→ 3´ exonuclease function.

Source: Purified from a strain of *E. coli* that carries a T4 DNA Polymerase overproducing plasmid.

Supplied in: 100 mM $KP0_4$ (pH 6.5), 1 mM DTT and 50% glycerol.

Quality Control Assays

SDS-PAGE Purity: SDS-PAGE analysis of this enzyme indicates > 95% enzyme purity.

Endonuclease Activity: Incubation of a minimum of 50 units of this enzyme with 1 µg of ϕX174 RF I DNA in assay buffer for 4 hours at 37°C in 50 µl reactions results in less than 10% conversion to RF II as determined by agarose gel electrophoresis.

Phosphatase Activity: Incubation of a minimum of 30 units of this enzyme in protein phosphatase assay buffer (1 M diethanolamine @ pH 9.8 and 0.5 mM $MgCl_2$) containing 2.5 mM *p*-nitrophenyl phosphate at 37°C for 4 hours yields no detectable *p*-nitrophenylene anion as determined by spectrophotometric analysis at 405 nm.

Functional Activity (Nucleotide Incorporation): One unit of this enzyme incorporates 10 nmol of dNTP into acid-precipitable material in a total reaction volume of 50 µl in 30 minutes at 37°C in 1X T4 DNA Polymerase Reaction Buffer with 33 µM dNTPs including [^{3}H]-dTTP, 70 µg/ml denatured herring sperm DNA and 50 µg/ml BSA.

Lot Controlled

References:

1. Tabor, S. and Struhl, K. (1989). DNA-Dependent DNA Polymerases. In F. M. Ausebel, R. Brent, R. E. Kingston, D. D. Moore, J. G. Seidman, J. A. Smith and K. Struhl (Eds.), *Current Protocols in Molecular Biology* (pp. 3.5.10–3.5.12). New York: John Wiley & Sons Inc.
2. Sambrook, J. et al. (1989). *Molecular Cloning: A Laboratory Manual,* (2nd ed.), (pp. 5.44–5.47). Cold Spring Harbor: Cold Spring Harbor Laboratory Press.

DNA Polymerase I, Large (Klenow) Fragment

#E6004A: 0.015 ml
#E6004AA: 0.06 ml

Store at –20°C

Description: DNA Polymerase I, Large (Klenow) Fragment is a proteolytic product of *E. coli* DNA Polymerase I which retains polymerization and 3´→ 5´ exonuclease activity, but has lost 5´→ 3´ exonuclease activity. Klenow retains the polymerization fidelity of the holoenzyme without degrading 5´ termini.

Source: A genetic fusion of the *E. coli pol*A gene, that has its 5´→ 3´ exonuclease domain genetically replaced by maltose binding protein (MBP). Klenow Fragment is cleaved from the fusion and purified away from MBP. The resulting Klenow fragment has the identical amino and carboxy termini as the conventionally prepared Klenow fragment.

Supplied in: 25 mM Tris-HCl (pH 7.4), 0.1 mM EDTA, 1 mM dithiothreitol and 50% glycerol.

Quality Control Assays

SDS-PAGE Purity: SDS-PAGE analysis of this enzyme indicates > 95% enzyme purity.

16-Hour Incubation: 50 µl reactions containing a minimum of 5 units of this enzyme and 1 µg of HindIII digested Lambda DNA incubated for 16 hours at 37°C results in no detectable non-specific nuclease degradation as determined by agarose gel electrophoresis. 50 µl reactions containing a minimum of 5 units of this enzyme and 1 µg T3 DNA incubated for 16 hours at 37°C also results in no detectable non-specific nuclease degradation as determined by agarose gel electrophoresis.

Endonuclease Activity: Incubation of a minimum of 50 units of this enzyme with 1 µg of ϕX174 RF I DNA in assay buffer for 4 hours at 37°C in 50 µl reactions results in less than 10% conversion to RF II as determined by agarose gel electrophoresis.

Phosphatase Activity: Incubation of a minimum of 50 units of this enzyme in protein phosphatase assay buffer (1 M diethanolamine @ pH 9.8 and 0.5 mM $MgCl_2$) containing 2.5 mM *p*-nitrophenyl phosphate at 37°C for 4 hours yields no detectable *p*-nitrophenylene anion as determined by spectrophotometric analysis at 405 nm.

RNase Activity: Incubation of a minimum of 5 units of this enzyme with 40 ng of a FAM-labeled RNA transcript for 16 hours at 37°C results in no detectable RNase activity as determined by polyacrylamide gel electrophoresis.

Functional Activity (Nucleotide Incorporation): One unit of this enzyme incorporates 10 nmol of dNTP into acid-precipitable material in a total reaction volume of 50 µl in 30 minutes at 37°C in 1X NEBuffer 2 with 33 µM dNTPs including [^{3}H]-dTTP, 70 µg/ml denatured herring sperm DNA and 50 µg/ml BSA.

Lot Controlled

Reference:

1. Sambrook, J. et al. (1989). *Molecular Cloning: A Laboratory Manual,* (2nd ed.), (pp. 5.40–5.43). Cold Spring Harbor: Cold Spring Harbor Laboratory Press.

T4 Polynucleotide Kinase

#E6205A: 0.015 ml
#E6205AA: 0.06 ml

Store at –20°C

Description: Catalyzes the transfer and exchange of P_i from the γ position of ATP to the 5´-hydroxyl terminus of polynucleotides (double- and single-stranded DNA and RNA) and nucleoside 3´-monophosphates. Polynucleotide Kinase also catalyzes the removal of 3´-phosphoryl groups from 3´-phosphoryl polynucleotides, deoxynucleoside 3´-monophosphates and deoxynucleoside 3´-diphosphates (1).

Source: An *E. coli* strain that carries the cloned T4 Polynucleotide Kinase gene. T4 Polynucleotide Kinase is purified by a modification of the method of Richardson (1).

Supplied in: 50 mM KCl, 10 mM Tris-HCl (pH 7.4), 0.1 mM EDTA, 1 mM dithiothreitol, 0.1 µM ATP and 50% glycerol.

Quality Assurance: Free of exonuclease, phosphatase, endonuclease and RNase activities. Each lot is tested under 5´-end-labeling conditions to assure maximal transfer of [^{32}P].

Quality Control Assays

SDS-PAGE Purity: SDS-PAGE analysis of this enzyme indicates > 95% enzyme purity.

16-Hour Incubation: 50 µl reactions containing a minimum of 10 units of this enzyme and 1 µg of HindIII digested Lambda DNA incubated for 16 hours at 37°C results in no detectable non-specific nuclease degradation as determined by agarose gel electrophoresis. 50 µl reactions containing a minimum of 10 units of this enzyme and 1 µg T3 DNA incubated for 16 hours at 37°C also results in no detectable non-specific nuclease degradation as determined by agarose gel electrophoresis.

Endonuclease Activity: Incubation of a minimum of 200 units of this enzyme with 1 µg of ϕX174 RF I DNA in assay buffer for 4 hours at 37°C in 50 µl reactions results in less than 10% conversion to RF II as determined by agarose gel electrophoresis.

Phosphatase Activity: Incubation of a minimum of 100 units of this enzyme in protein phosphatase assay buffer (1 M diethanolamine @ pH 9.8 and 0.5 mM $MgCl_2$) containing 2.5 mM *p*-nitrophenyl phosphate at 37°C for 4 hours yields no detectable *p*-nitrophenylene anion as determined by spectrophotometric analysis at 405 nm.

RNase Activity: Incubation of a minimum of 100 units of this enzyme with 2 µg MS2 phage RNA for 1 hour at 37°C in 50 µl 1X T4 Polynucleotide Kinase Reaction Buffer followed by agarose gel electrophoresis shows no degradation. Incubation of 10 units of this enzyme with 40 ng of a FAM- labeled RNA transcript for 16 hours at 37°C results in no detectable RNase activity as determined by polyacrylamide gel electrophoresis.

Exonuclease Activity: Incubation of 300 units of enzyme with 1 µg sonicated [^{3}H]DNA (10^5 cpm/µg) for 4 hours at 37°C in 50 µl reaction buffer released < 0.1% radioactivity.

Functional Activity (Labeling): ^{32}P end labeling of 5´-hydroxyl terminated $d(T)_8$ with a minimum of 50 units of this enzyme for 30 minutes at 37°C in 50 µl 1X T4 Polynucleotide Kinase Buffer followed by 20% acrylamide gel electrophoresis reveals that less than 1% of the product has been degraded by exonuclease or phosphatase activities.

Lot Controlled

References:

1. Richardson, C.C. (1981). In P.D. Boyer (Ed.), *The Enzymes* Vol. 14, (pp. 299–314). San Diego: Academic Press.
2. Sambrook, J. et al. (1989) *Molecular Cloning: A Laboratory Manual*, (2nd ed.), (pp. 10.59–10.67, 11.31–11.33). Cold Spring Harbor: Cold Spring Harbor Laboratory Press.

Deoxyadenosine 5´- Triphosphate (dATP)

#E6006A: 0.120 ml **Concentration: 1.0 mM**
#E6006AA: 0.600 ml

Store at –20°C

Supplied in: Milli-Q water as a sodium salt at pH 7.5.

Concentration: dATP is supplied at a concentration of 1 mM.

Quality Assurance: Nucleotide solutions are certified free of nucleases and phosphatases.

Notes: Storing nucleotide triphosphates in solutions containing magnesium promotes triphosphate degradation. Nucleotide concentrations are determined by measurements of absorbance.

Quality Control Assays

Phosphatase Activity: Incubation of a minimum of 1 mM dATP in protein phosphatase assay buffer (1M diethanolamine @ pH 9.8 and 0.5 mM $MgCl_2$) containing 2.5 mM *p*-nitrophenyl phosphate at 37°C for 4 hours yields no detectable *p*-nitrophenylene anion as determined by spectrophotometric analysis at 405 nm.

16-Hour Incubation: 50 µl reactions containing a minimum of 0.2 mM dATP and 1 µg of HindIII digested Lambda DNA incubated for 16 hours results in no detectable non-specific nuclease degradation as determined by agarose gel electrophoresis. 50 µl reactions containing 0.2 mM dATP and 1 µg T3 DNA incubated for 16 hours also results in no detectable non-specific nuclease degradation as determined by agarose gel electrophoresis.

RNase Activity: Incubation of a minimum of 0.1 mM dATP with 40 ng of a FAM-labeled RNA transcript for 16 hours at 37°C results in no detectable RNase activity as determined by polyacrylamide gel electrophoresis.

HPLC: dATP purity is determined by HPLC to be > 99%.

Functional Activity (PCR): This dATP in a pool of dNTPs is tested in 25 cycles of PCR amplification generating 0.5 kb, 2 kb, and 5kb amplicons from lambda DNA.

Lot Controlled

Klenow Fragment (3´→5´ exo⁻)

#E6207A: 0.015 ml
#E6207AA: 0.06 ml

Store at –20°C

Description: Klenow Fragment (3´→ 5´ exo⁻) is an N-terminal truncation of DNA Polymerase I which retains polymerase activity, but lacks the 5´→ 3´ exonuclease activity and contains mutations (D355A, E357A), which abolish the 3´→ 5´ exonuclease activity (1).

Source: An *E. coli* strain containing a plasmid with a fragment of the *E.coli* polA (D355A, E357A) gene starting at codon 324.

Supplied in: 25 mM Tris-HCl (pH 7.4), 0.1 mM EDTA, 1 mM DTT and 50% glycerol.

Quality Control Assays

SDS-PAGE Purity: SDS-PAGE analysis of this enzyme indicates > 95% enzyme purity.

16-Hour Incubation: 50 µl reactions containing a minimum of 5 units of this enzyme and 1 µg of HindIII digested Lambda DNA incubated for 16 hours at 37°C results in no detectable non-specific nuclease degradation as determined by agarose gel electrophoresis. 50 µl reactions containing a minimum of 5 units of this enzyme and 1 µg T3 DNA incubated for 16 hours at 37°C also results in no detectable non-specific nuclease degradation as determined by agarose gel electrophoresis.

Endonuclease Activity: Incubation of a minimum of 50 units of this enzyme with 1 µg of ϕX174 RF I DNA in assay buffer for 4 hours at 37°C in 50 µl reactions results in less than 10% conversion to RF II as determined by agarose gel electrophoresis.

Phosphatase Activity: Incubation of a minimum of 50 units of this enzyme in protein phosphatase assay buffer (1 M diethanolamine @ pH 9.8 and 0.5 mM $MgCl_2$) containing 2.5 mM *p*-nitrophenyl phosphate at 37°C for 4 hours yields no detectable *p*-nitrophenylene anion as determined by spectrophotometric analysis at 405 nm.

RNase Activity: Incubation of a minimum of 5 units of this enzyme with 40 ng of a FAM- labeled RNA transcript for 16 hours at 37°C results in no detectable RNase activity as determined by polyacrylamide gel electrophoresis.

Exonuclease Activity: Incubation of a minimum of 200 units of this enzyme with 1 µg sonicated [^{3}H] DNA (10^5 cpm/µg) for 4 hours at 37°C in 50 µl reaction buffer releases < 0.1% radioactivity.

3´→5´ Exonuclease Activity: Incubation of a minimum of 50 units of enzyme in 20 µl of a 10 nM solution of a fluorescent 5´-FAM labeled oligonucleotide for 30 minutes at 37°C yields no detectable 3´→5´ degradation as determined by capillary electrophoresis.

Functional Activity (Nucleotide Incorporation): One unit of this enzyme incorporates 10 nmol of dNTP into acid-precipitable material in a total reaction volume of 50 µl in 30 minutes at 37°C in 1X NEBuffer 2 with 33 µM dNTPs including [^{3}H]-dTTP, 70 µg/ml denatured herring sperm DNA and 50 µg/ml BSA.

Reference:

1. Derbyshire, V. et al. (1988) *Science* 240, 199–201.

NEBuffer 2 for Klenow Fragment (3´→5´ exo⁻)

#E6008A: 0.06 ml **Concentration: 10X**
#E6008AA: 0.300 ml

Store at –20°C

1X NEBuffer 2:
50 mM NaCl
10 mM Tris-HCl
10 mM $MgCl_2$
1 mM DTT
pH 7.9 @ 25°C

Quality Control Assays

16-Hour Incubation: 50 µl reactions containing this reaction buffer at 1X concentration and 1 µg of HindIII digested Lambda DNA incubated for 16 hours at 37°C results in no detectable non-specific nuclease degradation as determined by agarose gel electrophoresis. 50 µl reactions containing this reaction buffer at 1X concentration and 1 µg T3 DNA incubated for 16 hours at 37°C also results in no detectable non-specific nuclease degradation as determined by agarose gel electrophoresis.

Endonuclease Activity: Incubation of this reaction buffer at a 1X concentration with 1 µg of ϕX174 RF I DNA for 4 hours at 37°C in 50 µl reactions results in less than 10% conversion to RF II as determined by agarose gel electrophoresis.

RNase Activity: Incubation of this reaction buffer at 1X concentration with 40 ng of a FAM-labeled RNA transcript for 16 hours at 37°C results in no detectable RNase activity as determined by polyacrylamide gel electrophoresis.

Phosphatase Activity: Incubation of this reaction buffer at a 1X concentration in protein phosphatase assay buffer (1 M diethanolamine @ pH 9.8 and 0.5 mM $MgCl_2$) containing 2.5 mM *p*-nitrophenyl phosphate at 37°C for 4 hours yields no detectable *p*-nitrophenylene anion as determined by spectrophotometric analysis at 405 nm.

Lot Controlled

Quick T4 DNA Ligase

#E6209A: 0.048 ml
#E6209AA: 0.24 ml

Store at –20°C

Source: Purified from *E. coli* C600 pcl857 pPLc28 lig8 (2).

Quality Control Assays

SDS-PAGE Purity: SDS-PAGE analysis of this enzyme indicates > 95% enzyme purity.

16-Hour Incubation: 50 µl reactions containing a minimum of 2,000 units of this enzyme and 1 µg of HindIII digested Lambda DNA incubated for 16 hours at 37°C results in no detectable non-specific nuclease degradation as determined by agarose gel electrophoresis. 50 µl reactions containing a minimum of 2,000 units of this enzyme and 1 µg T3 DNA incubated for 16 hours at 37°C also results in no detectable non-specific nuclease degradation as determined by agarose gel electrophoresis.

Endonuclease Activity: Incubation of a minimum of 3,200 units of this enzyme with 1 µg of ϕX174 RF I DNA in assay buffer for 4 hours at 37°C in 50 µl reactions results in less than 10% conversion to RF II as determined by agarose gel electrophoresis.

Phosphatase Activity: Incubation of a minimum of 20,000 units of this enzyme in protein phosphatase assay buffer (1 M diethanolamine @ pH 9.8 and 0.5 mM $MgCl_2$) containing 2.5 mM *p*-nitrophenyl phosphate at 37°C for 4 hours yields no detectable *p*-nitrophenylene anion as determined by spectrophotometric analysis at 405 nm.

RNase Activity: Incubation of a minimum of 2,000 units of this enzyme with 40 ng of a FAM-labeled RNA transcript for 16 hours at 37°C results in no detectable RNase activity as determined by polyacrylamide gel electrophoresis.

Exonuclease Activity: Incubation of a minimum of 3,200 units of this enzyme with 1 µg sonicated [^{3}H] DNA (10^5 cpm/µg) for 4 hours at 37°C in 50 µl reaction buffer releases < 0.1% radioactivity.

Functional Activity (Blunt End Ligation): 50 µl reactions containing a 0.5 µl Quick T4 DNA Ligase, 18 µg HaeIII digested ϕX174 and 1X T4 DNA Ligase Buffer incubated at 16°C for 7.5 min results in > 95% of fragments ligated as determined by agarose gel electrophoresis.

Functional Activity (Cohesive End Ligation): 20 µl reactions containing 0.5 µl Quick T4 DNA Ligase, 12 µg HindIII digested lambda DNA and 1X T4 DNA Ligase Buffer incubated at 37°C overnight results in > 95% of fragments ligated as determined by agarose gel electrophoresis. Redigestion of the ligated products, 50 µl reactions containing 6 µg of the ligated fragments, 40 units HindIII, and 1X NEBuffer 2 incubated at 37°C for 2 hours, results in no detectable undigested fragments as determined by agarose gel electrophoresis.

Functional Activity (Adaptor Ligation): 50 µl reactions containing 0.125 µl Quick T4 DNA Ligase, 8 nmol 12 bp adaptor, and 1X T4 DNA Ligase Buffer incubated at 16°C overnight results in no detectable unligated adaptor as determined by agarose gel electrophoresis.

Functional Activity (Transformation): After a five-minute ligation of linearized, dephosphorylated LITMUS™ 28 (containing either blunt [EcoRV] or cohesive [HindIII] ends) and a mixture of compatible insert fragments, transformation into chemically competent *E. coli* DH-5 alpha cells yields a minimum of 1 x 10^6 recombinant transformants per µg plasmid DNA.

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References:

1. Engler, M. J. and Richardson, C. C. (1982). In P. D. Boyer (Ed.), *The Enzymes* Vol. 5, (p. 3). San Diego: Academic Press.
2. Remaut, E., Tsao, H. and Fiers, W. (1983) *Gene* 22, 103–113.

Quick Ligation Reaction Buffer

#E6210A: 0.18 ml **Concentration: 2X**
#E6210AA: 0.900 ml

Store at –20°C

1X Quick Ligation Reaction Buffer:
66 mM Tris-HCl
10 mM $MgCl_2$
1 mM dithiothreitol
1 mM ATP
7.5% Polyethylene glycol (PEG 6000)
pH 7.6 @ 25°C

Quality Control Assays

16-Hour Incubation: 50 µl reactions containing this reaction buffer at 1X concentration and 1 µg of HindIII digested Lambda DNA incubated for 16 hours at 37°C results in no detectable non-specific nuclease degradation as determined by agarose gel electrophoresis. 50 µl reactions containing this reaction buffer at 1X concentration and 1 µg T3 DNA incubated for 16 hours at 37°C also results in no detectable non-specific nuclease degradation as determined by agarose gel electrophoresis.

Endonuclease Activity: Incubation of this reaction buffer at a 1X concentration with 1 µg of ϕX174 RF I DNA for 4 hours at 37°C in 50 µl reactions results in less than 10% conversion to RF II as determined by agarose gel electrophoresis.

RNase Activity: Incubation of this reaction buffer at 1X concentration with 40 ng of a FAM-labeled RNA transcript for 16 hours at 37°C results in no detectable RNase activity as determined by polyacrylamide gel electrophoresis.

Phosphatase Activity: Incubation of this reaction buffer at a 1X concentration in protein phosphatase assay buffer (1M diethanolamine @ pH 9.8 and 0.5 mM $MgCl_2$) containing 2.5 mM *p*-nitrophenyl phosphate at 37°C for 4 hours yields no detectable *p*-nitrophenylene anion as determined by spectrophotometric analysis at 405 nm.

Lot Controlled

Phusion High-Fidelity PCR Master Mix with HF Buffer

E6012A: **0.3 ml** **Concentration: 2X**
E6012AA: **1.5 ml**

Store at –20°C

Storage and Stability: Store at –20°C. After thawing the mix can be refrozen or optionally stored at 4°C for three months.

Component Specifications: 2X Phusion High-Fidelity Master Mix with HF Buffer contains 0.04 units/µl Phusion DNA Polymerase, 2X Phusion HF Buffer (provides 1.5 mM $MgCl_2$ in final reaction concentration), and 400 µM of each dNTP.

Manufactured and quality controlled by New England Biolabs, Inc., Thermo Scientific Phusion® High-Fidelity DNA Polymerase is purified from an *E. coli* strain expressing the cloned Phusion DNA Polymerase gene. This DNA polymerase possesses the following activities: 5´→3´ DNA polymerase activity and 3´→5´ exonuclease activity. Phusion DNA Polymerase is purified free of contaminating endo- and exonucleases.

Quality Control Assays

16-Hour Incubation: 50 µl reactions containing 1X Phusion Master Mix and 1 µg of HaeIII digested ϕX174 RF I DNA incubated for 16 hours at 37°C results in no detectable non-specific nuclease degradation as determined by agarose gel electrophoresis. 50 µl reactions containing 1X Phusion Master Mix and 1 µg T3 DNA incubated for 16 hours at 37°C also results in no detectable non-specific nuclease degradation as determined by agarose gel electrophoresis.

Phosphatase Activity: Incubation of 1X Phusion Master Mix in protein phosphatase assay buffer (1 M diethanolamine @ pH 9.8 and 0.5 mM $MgCl_2$) containing 2.5 mM *p*-nitrophenyl phosphate at 37°C for 4 hours yields no detectable *p*-nitrophenylene anion as determined by spectrophotometric analysis at 405 nm.

7.5 kb Genomic and 20 kb Lambda PCR: 30 cycles of PCR amplification in a 50 µl reaction containing 50 ng genomic DNA with 1X Phusion High-Fidelity PCR Master Mix with HF Buffer and 1.0 µM primers results in the expected 7.5 kb product.

20 kb Lambda DNA PCR: 22 cycles of PCR amplification in a 50 µl reaction containing 10 ng Lambda DNA with 1X Phusion High-Fidelity PCR Master Mix with HF Buffer and 1.0 µM primers results in the expected 20 kb product.

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Notice to Customer: Phusion® DNA Polymerase was developed by Finnzymes Oy, now a part of Thermo Fisher Scientific.

NEBNext High-Fidelity 2X PCR Master Mix

E6013A: 0.3 ml **Concentration: 2X**
E6013AA: 0.75 ml

Store at –20°C

Description: The NEBNext High-Fidelity 2X PCR Master Mix is specifically optimized for robust, high-fidelity amplification of next-generation sequencing (NGS) libraries, regardless of GC content. The polymerase component of the master mix, Q5™ High-Fidelity DNA Polymerase, is a novel thermostable DNA polymerase that possesses 3´→ 5´ exonuclease activity, and is fused to a processivity-enhancing Sso7d domain. Q5 High-Fidelity DNA Polymerase also has an ultra-low error rate (> 50-fold lower than that of *Taq* DNA Polymerase and 6-fold lower than that of *Pyrococcus furiosus* (Pfu) DNA Polymerase)

Quality Control Assays

16-Hour Incubation: A 50 µl reactions containing NEBNext High-Fidelity 2X PCR Master Mix and 1 µg of HindIII digested Lambda DNA incubated for 16 hours at 37°C results in no detectable non-specific nuclease degradation as determined by agarose gel electrophoresis. 50 µl reactions containing 100 units of NEBNext High-Fidelity 2X PCR Master Mix and 1 µg of T3 DNA incubated for 16 hours at 37°C results in no detectable non-specific nuclease degradation as determined by agarose gel electrophoresis.

Phosphatase Activity: Incubation of NEBNext High-Fidelity 2X PCR Master Mix in protein phosphatase assay buffer (1 M diethanolamine @ pH 9.8 and 0.5 mM $MgCl_2$) containing 2.5 mM *p*-nitrophenyl phosphate at 37°C for 4 hours yields no detectable *p*-nitrophenylene anion as determined by spectrophotometric analysis at 405 nm.

GC-Rich Genomic PCR: 30 cycles of PCR amplification of 20 ng genomic DNA in a 50 µl reaction containing 0.5 µM primers and 1X NEBNext High-Fidelity PCR Master Mix result in the expected 737 bp product.

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