Thermo
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PRODUCT INFORMATION

BamHI

#ER0051 4000 u

Lot: Expiry Date:

5'...**G↓G A T C C**...3'
3'...**C C T A G↑G**...5'

Concentration: 10 u/µl
Source: *E.coli* that carries the cloned *bamHIR* gene from *Bacillus amyloliquefaciens* H
Supplied with: 2 x 1 ml of 10X Buffer BamHI
1 ml of 10X Buffer Tango

Store at -20°C

In total 4 vials. BSA included

www.thermoscientific.com/fermentas

RECOMMENDATIONS

1X Buffer BamHI (for 100% BamHI digestion)
10 mM Tris-HCl (pH 8.0), 5 mM $MgCl_2$, 100 mM KCl, 0.02% Triton X-100, 0.1 mg/ml BSA.

Incubation temperature
37°C.

Unit Definition
One unit is defined as the amount of BamHI required to digest 1 µg of lambda DNA-Bsp120I fragments in 1 hour at 37°C in 50 µl of recommended reaction buffer.

Dilution
Dilute with Dilution Buffer (#B19): 10 mM Tris-HCl (pH 7.4 at 25°C) 100 mM KCl, 1 mM EDTA, 1 mM DTT, 0.2 mg/ml BSA and 50% glycerol.

Double Digests
Thermo Scientific Tango Buffer is provided to simplify buffer selection for double digests. 98% of Thermo Scientific restriction enzymes are active in a 1X or 2X concentration of Tango™ Buffer. Please refer to to www.fermentas.com/doubledigest to choose the best buffer for your experiments.
1X Tango Buffer: 33 mM Tris-acetate (pH 7.9 at 37°C), 10 mM magnesium acetate, 66 mM potassium acetate, 0.1 mg/ml BSA.

Storage Buffer

BamHI is supplied in: 10 mM Tris-HCl (pH 7.4 at 25°C), 200 mM NaCl, 1 mM DTT, 0.1 mM EDTA, 0.15% Triton X-100, 0.2 mg/ml BSA and 50% glycerol.

Recommended Protocol for Digestion

- Add:

nuclease-free water	16 µl
10X Buffer BamHI	2 µl
DNA (0.5-1 µg/µl)	1 µl
BamHI	0.5-2 µl***,****

- Mix gently and spin down for a few seconds.
- Incubate at 37°C for 1-16 hours******.

The digestion reaction may be scaled either up or down.

Recommended Protocol for Digestion of PCR Products Directly after Amplification

- Add:

PCR reaction mixture	10 µl (~0.1-0.5 µg of DNA)
nuclease-free water	18 µl
10X Buffer BamHI	2 µl
BamHI	1-2 µl***,****

- Mix gently and spin down for a few seconds.
- Incubate at 37°C for 1-16 hours******.

***** This volume of the enzyme is recommended for preparations of standard concentrations (10 u/µl), whereas HC enzymes (50 u/µl) should be diluted with Dilution Buffer to obtain 10 u/µl concentration.

****** *See* Overdigestion Assay.

Thermal Inactivation

Only small amounts of BamHI (up to 10 units) can be inactivated at 80°C in 20 min.

Inactivation Procedure

- To prepare the digested DNA for electrophoresis:
  – stop the digestion reaction by adding 0.5 M EDTA, pH 8.0 (#R1021), to achieve a 20 mM final concentration. Mix thoroughly, add an electrophoresis loading dye and load onto gel.
- To prepare DNA suitable for further enzymatic reactions:
  – extract with phenol/chloroform, precipitate with ethanol or isopropanol, wash the pellet with 75% cold ethanol and air-dry;
  – dissolve DNA in either nuclease-free water, TE buffer, or a buffer suitable for further applications;
  – check the DNA concentration in the solution.

For **ENZYME PROPERTIES** and **CERTIFICATE OF ANALYSIS** *see* back page

ENZYME PROPERTIES

Enzyme Activity in Thermo Scientific REase Buffers, %

BamHI	B	G	O	R	Tango	2X Tango
100	20-50**	100	20-50	50-100**	100**	50-100

**Star activity appears at a greater than 5-fold overdigestion (5 u x 1h).

Methylation Effects on Digestion

Dam: completely overlaps – no effect.
Dcm: may overlap – no effect.
CpG: may overlap – no effect.
EcoKI: never overlaps – no effect.
EcoBI: never overlaps – no effect.

Stability during Prolonged Incubation

A minimum of 0.5 units of the enzyme is required for complete digestion of 1 µg of lambda DNA in 16 hours at 37°C.

Digestion of Agarose-embedded DNA

A minimum of 5 units of the enzyme is required for complete digestion of 1 µg of agarose-embedded lambda DNA in 16 hours.

Compatible Ends

BclI, BglII, Bsp143I, MboI, PsuI

Number of Recognition Sites in DNA

λ	ΦX174	pBR322	pUC57	pUC18/19	pTZ19R/U	M13mp18/19
5	0	1	1	1	1	1

CERTIFICATE OF ANALYSIS

Overdigestion Assay

No detectable change in the specific fragmentation pattern is observed after an 80-fold overdigestion with BamHI (5 u/µg lambda DNA x 16 hours).

Ligation/Recutting Assay

After a 50-fold overdigestion (3 u/µg DNA x 17 hours) with BamHI, more than 95% of the digested DNA fragments can be ligated at a 5'-termini concentration of 0.05 µM. More than 95% of these sites can be recut.

Labeled Oligonucleotide (LO) Assay

No detectable degradation of single-stranded or double-stranded labeled oligonucleotides occurred during incubation with 10 units of BamHI for 4 hours.

Blue/White Cloning Assay

pUC57 was incubated with 10 units of BamHI for 16 hours. After religation and transformation, the background level of white colonies was <1%.

Quality authorized by:  Jurgita Zilinskiene

PRODUCT USE LIMITATION

This product is developed, designed and sold exclusively *for research purposes and in vitro use only.* The product was not tested for use in diagnostics or for drug development, nor is it suitable for administration to humans or animals.

Please refer to www.thermoscientific.com/fermentas for Material Safety Data Sheet of the product.