

PRODUCT INFORMATION

Klenow Fragment

#EP0051 300 u

Lot: **Expiry Date:**

Concentration: 10 u/μl
Supplied with: 1 ml of 10X Reaction Buffer

Store at -20°C

In total 2 vials.

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Description

Klenow Fragment is the Large Fragment of DNA Polymerase I, *E.coli*. It exhibits 5'→3' polymerase activity and 3'→5' exonuclease (proofreading) activity, but lacks 5'→3' exonuclease activity of DNA Polymerase I.

Applications

- DNA blunting by fill-in of 5'-overhangs. (1), see protocols on back page.
- Random-primed DNA labeling (2-4).
- Labeling by fill-in 5'-overhangs of dsDNA.
- DNA sequencing by the Sanger method (5).
- Site-specific mutagenesis of DNA with synthetic oligonucleotides (6).
- Second strand synthesis of cDNA (7).

Source

E.coli cells with a cloned fragment of the *polA* gene.

Molecular Weight

68 kDa monomer.

Definition of Activity Unit

One unit of the enzyme catalyzes the incorporation of 10 nmol of deoxyribonucleotides into a polynucleotide fraction (adsorbed on DE-81) in 30 min at 37°C. Enzyme activity is assayed in the following mixture: 50 mM Tris-HCl (pH 8.0 at 25°C), 5 mM MgCl₂, 1 mM DTT, 0.033 mM dNTP, 0.4 M Bq/ml [³H]-dTTP and 62.5 μg/ml activated salmon milt DNA.

Rev.7



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Storage Buffer

The enzyme is supplied in: 25 mM Tris-HCl (pH 7.5), 0.1 mM EDTA, 1 mM DTT and 50% (v/v) glycerol.

10X Reaction Buffer

500 mM Tris-HCl (pH 8.0 at 25°C), 50 mM MgCl₂, 10 mM DTT.

Inhibition and Inactivation

- Inhibitors: metal chelators, PP_i, P_i (at high concentrations) (8).
- Inactivated by heating at 75°C for 10 min or by addition of EDTA.

Note

- Activity of Klenow Fragment in Thermo Scientific buffers (in comparison to activity in assay buffer):


| Buffers | Activity, % |
|--|-------------|
| for restriction enzymes: | |
| Thermo Scientific FastDigest, FastDigest® Green, O, R, 1X Thermo Scientific Tango, 2X Tango™, BamHI, EcoRI | 100 |
| Ecl136II, PacI, SacI, KpnI | 50-75 |
| B | 25-50 |
| G | 20-50 |
| for PCR buffers: | |
| Taq buffer with KCl, Taq buffer with (NH ₄) ₂ SO ₄ , Pfu buffer | 100 |
| RT buffers | 100 |

CERTIFICATE OF ANALYSIS

Endodeoxyribonuclease Assay

No conversion of covalently closed circular DNA to nicked DNA was detected after incubation of 20 units of Klenow Fragment with 1 μg of pUC19 DNA for 4 hours at 37°C.

Quality authorized by:

 Jurgita Zilinskiene

(continued on back page)

Protocol for DNA 3'-end labeling by fill-in of 5'-overhangs

1. Prepare the following reaction mixture:

| | |
|--|---|
| Linear DNA | 0.1-4 µg |
| 10X reaction buffer for Klenow Fragment | 2 µl |
| [α-³²P]-dNTP, ~15-30 TBq/mmol (400-800 Ci/mmol) <i>or</i> | 0.74 MBq (20 µCi) |
| [α-³²P]-dNTP, ~110 TBq/mmol (3000 Ci/mmol) | 2.96 MBq (80 µCi) |
| 3 dNTP Mix, 2 mM each (without a labeled dNTP) | 2.5 µl (0.25 mM final concentration) |
| Klenow Fragment | 0.1 µl (1 u) |
| Water, nuclease-free (#R0581) | to 20 µl |
| Total volume | 20 µl |

2. Incubate at 37°C for 15 min.

3. Stop the reaction by heating at 75°C for 10 min.

Note

This protocol is suitable for labeling of the following Fermentas DNA markers, composed of DNA fragments with 5'-overhangs:

Lambda DNA EcoRI Marker, #SM0281

Lambda DNA HindIII Marker, #SM0101

Lambda DNA EcoRI/HindIII Marker, #SM0191

Lambda DNA Eco91I Marker, #SM0111

ΦX174 DNA HinfI Marker, #SM0261

- The modified version of this protocol can be used for nonradioactive labeling of DNA markers. Substitute a part of dTTP with a modified nucleotide (e.g. Biotin-11-dUTP or Fluorescein-12-dUTP) at a molar ratio of 1:2.

Protocol for Fill-in of 5'-overhangs

1. Prepare the following reaction mixture:

| | |
|--|--------------------------------------|
| Linear DNA | 10-15 µl (0.1-4 µg) |
| 10X reaction buffer for Klenow Fragment | 2 µl |
| dNTP Mix, 2mM each (#R0241) | 0.5 µl (0.05 mM final concentration) |
| Klenow Fragment | 0.1-0.5 µl (1-5 u) |
| Water, nuclease-free (#R0581) | to 20 µl |
| Total volume | 20 µl |

2. Mix thoroughly, spin briefly and incubate at 37°C for 10 min.

3. Stop the reaction by heating at 75°C for 10 min.

Note

The enzyme incorporates modified nucleotides (e.g. biotin-, digoxigenin-, fluorescently-labeled nucleotides).

References

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4. Yu, H., et al., Cyanine dye dUTP analogs for enzymatic labeling of DNA probes, *Nucleic Acids Res.*, 22, 3226-3232, 1994.
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8. Eun, H.-M., *Enzymology Primer for Recombinant DNA Technology*, Academic Press, Inc., 1996.

PRODUCT USE LIMITATION

This product is developed, designed and sold exclusively *for research purposes and in vitro use only*. The product was not tested for use in diagnostics or for drug development, nor is it suitable for administration to humans or animals. Please refer to www.thermoscientific.com/fermentas for Material Safety Data Sheet of the product.