

## CO<sub>2</sub> Independent Medium without L-glutamine

**CAUTION:** Human origin materials are non-reactive (donor level) for anti-HIV 1 & 2, anti-HCV, and HB<sub>s</sub>Ag. Handle in accordance with established bio-safety practices.

Cat. No.: 18045 500 mL

Storage Condition: 2 to 8°C, in the dark.

### Background

Traditionally cell culture media have relied on sodium bicarbonate as the primary buffering component.<sup>1</sup> When utilizing a bicarbonate buffer system, a carbon dioxide (CO<sub>2</sub>) rich atmosphere of five to ten percent in air is required to maintain the *in vitro* environment at physiological pH.<sup>1,2</sup> However, such a system does not allow for work outside of the CO<sub>2</sub> regulated atmosphere for extended periods of time due to extreme fluctuations in pH, which can affect cellular function.<sup>3,4</sup> As an alternative to bicarbonate and synthetic buffering systems, GIBCO has developed a CO<sub>2</sub> Independent Medium<sup>5,6</sup> which is capable of maintaining long term pH stability under atmospheric CO<sub>2</sub> (0.04% Figure 1). GIBCO CO<sub>2</sub> Independent Medium contains a unique buffering system composed of mono and dibasic sodium phosphate and β-glycerophosphate. A small amount of sodium bicarbonate has been included in the formulation to meet essential bicarbonate dependent functions.<sup>7,8,9</sup> No synthetic buffers are utilized, thus eliminating any cytotoxic effects associated with such buffering systems.<sup>10,11</sup> Additionally, GIBCO CO<sub>2</sub> Independent Medium has been formulated with components that enhance cellular production and utilization of CO<sub>2</sub> such that an exogenous source of CO<sub>2</sub> is not required for the maintenance of CO<sub>2</sub> dependent cellular functions.<sup>2,8,9,12</sup>

pH Stability of CO<sub>2</sub> Independent Medium

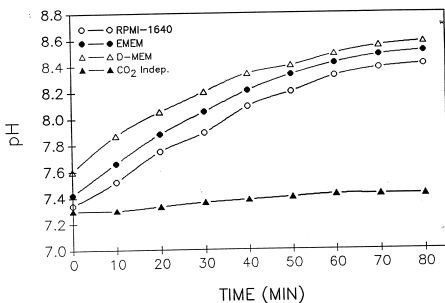


Figure 1. pH stability of CO<sub>2</sub> Independent Medium as compared to other media which utilize a NaHCO<sub>3</sub> buffering system. CO<sub>2</sub> Independent Medium has been found to maintain a stable pH over a 72-hour test period in atmospheric CO<sub>2</sub> without the presence of cells.

This product offers the user long term pH stability which allows for the maintenance of normal cellular function during routine culture procedures. Additionally, this medium provides versatility in that it can be utilized in either an open or closed culture system, the latter being ideally suited for use in toxicological and/or virological procedures where there is a risk of aerosol contamination or infection. It is also anticipated that this product would be beneficial for large scale cell culture procedures and for tissue transport.

### Quality Control Testing

CO<sub>2</sub> Independent Medium is quantitatively performance tested using a growth assay which monitors the 7-day growth kinetics of WI-38, a human diploid lung fibroblast cell line. Each production lot is further evaluated for pH stability over a 72-hour test period in atmospheric CO<sub>2</sub>. Additional standard evaluations are pH, osmolarity, endotoxin and tests for the presence of bacterial and fungal contaminants. See the available Certificate of Analysis for lot-specific test results and product specifications.

### Instructions for Use

CO<sub>2</sub> Independent Medium is supplied without L-glutamine and must be supplemented with 4 mM L-glutamine either as powder (584.6 mg/L) or liquid (20 mL of 100X stock, Cat. No. 25030). If a liquid L-glutamine source is used, an equivalent amount of medium should be removed to equal the volumetric amount of L-glutamine added. For the best performance of this product, it is recommended that an equivalent amount of medium also be removed to compensate for the addition of serum. The addition of L-glutamine and serum on top of the original volume may affect product performance.

### Applications

For the development of CO<sub>2</sub> Independent Medium a variety of anchorage dependent and suspension cell lines have been utilized. As summarized in Table 1, growth kinetic analysis demonstrated that all of the cell lines tested proliferated at rates equal to or greater than control cultures when grown in a closed, 0% CO<sub>2</sub> culture system. When utilizing an open, 0% CO<sub>2</sub> culture environment, some anchorage dependent cell lines (BHK-21, HT-29 and MDBK) may require adaptation to CO<sub>2</sub> Independent Medium prior to initiation of experiments. If desired, cultures can be placed in a 5% CO<sub>2</sub> incubator utilizing either an open or closed culture system without any deleterious effects to cellular growth.

Table 1: Summary of growth performance analysis of unadapted anchorage dependent and suspension cell lines cultured in CO<sub>2</sub> Independent Medium.

Cell Line	Culture Environment	
	Open System, 0% CO <sub>2</sub>	Closed System, 0% CO <sub>2</sub>
AE-1	ND	++
BHK-21	+/-	++
CHO	+	++
CHO*	ND	+
HT-29	+/-	+
MDBK**	+/-	++
VERO	+	+
WI-38	+	+

\* Adapted to suspension growth in CHO-S-SFM.

\*\* Display normal growth at low seeding density.

+ Cell growth is equal to the control.

++ Cell growth exceeds the control.

+/- Cell growth is slightly lower than control.

ND Not determined.

### Adaptation of Cells to CO<sub>2</sub> Independent Medium

For maximum growth performance, some cell lines may require either direct or sequential adaptation to CO<sub>2</sub> Independent Medium. In either case, the pre-adapted cell line should be in mid-logarithmic growth phase with high (>90%) viability. Success of the adaptation procedure will depend on the cell line being used and the culture conditions employed. It is recommended that the user first evaluate this product with unadapted cells since not all cell lines will require adaptation. If the growth assays employed are conducted in a closed culture system, stock cultures can be directly adapted and maintained in CO<sub>2</sub> Independent Medium using a closed, 0% CO<sub>2</sub> system.

#### A. Direct Adaptation

For direct adaptation of selected cell lines, stock cultures should be inoculated at normal seeding densities and incubated using a closed cap in a humidified (37°C) incubator with 0% CO<sub>2</sub>. Growth should be monitored daily and cells subcultured at 80-90% confluency. Subsequent passages should utilize a humidified (37°C) 0% CO<sub>2</sub> atmosphere with an open cap vessel. If the cell cultures fail to maintain acceptable growth and viability over 3-5 passages during direct adaptation, the sequential adaptation method should be used.

#### B. Sequential Adaptation

For this procedure cells are initially inoculated into a 50:50 ratio (v/v) of CO<sub>2</sub> Independent Medium and the currently utilized medium. Stock cultures should be maintained under an open cap in a humidified (37°C) incubator with 0% CO<sub>2</sub>. Flasks should be subcultured at 80-90% confluency and inoculated with a 75:25 ratio (v/v) of CO<sub>2</sub> Independent Medium and the currently utilized medium. Subsequent subculturing should use 100% CO<sub>2</sub> Independent Medium and maintained as described above.

### References:

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- Chang, R.S., Liepins, H. and Margolish, M. Carbon dioxide requirement and nucleic acid metabolism of HeLa and conjunctival cells. *Proc. Soc. Exper. BiolMed.* **106**:149-152 (1961).

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You may also contact your Invitrogen Sales Representative or our World Wide Web site at [www.invitrogen.com](http://www.invitrogen.com).

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**CAUTION: Not intended for human or animal diagnostic or therapeutic uses**